

Workshop in Systems Immunology



Emanuele de Rinaldis
Magnus Fontes
Giorgio Gaglia
Shameer Khader

June 19th 2023

Today's Plan

8:00-8:10 am	Course Overview and Objectives – Emanuele de Rinaldis
8:10-9:00 am	Introduction to Systems Immunology – Emanuele de Rinaldis
9:00-9:45 am	Systems Immunology & Immune Oncology: A Data-Centric View – Magnus Fontes
9:45-10:00 am	Break
10:00-11:30 am	Deep Dive Into Selected Scientific Case Studies: From Systems Immunology to Novel Therapeutic Insights – Emanuele de Rinaldis
11:30 am-12:00 pm	Q/A and Panel Discussion
12:00-1:00 pm	Break for Lunch
1:00-2:00 pm	Spatial Biology Methods and Analytics for Immunology & Oncology – Giorgio Gaglia
2:00-2:15 pm	Break
2:15-3:30 pm	Artificial Intelligence – A Primer for Immunologists – Shameer Khader
3:30-3:45 pm	Break
3:45-4:45 pm	Interactive Data Analysis Session – Magnus Fontes
4:45-5:00 pm	Wrap Up Notes & Final Remarks

Introduction to Systems Immunology

Technologies, Methods and Applications

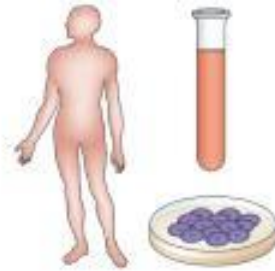
Emanuele de Rinaldis
FOCIS - June 19th, 2023



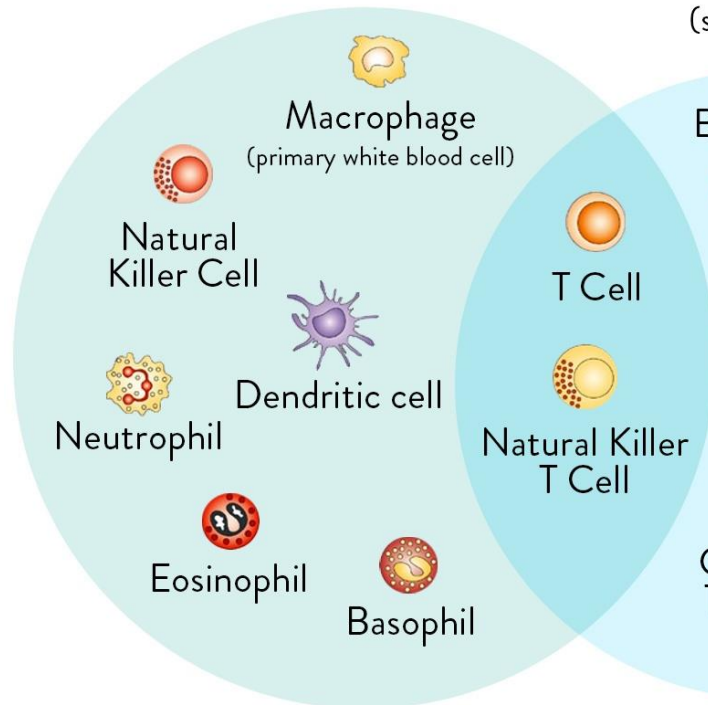
Outline

- ❑ Introduction to Systems Immunology Concepts
- ❑ Brief view/recap on immune profiling technologies and applications
 - ❑ Sequencing
 - ❑ Bulk vs Single-Cell
 - ❑ Application of Single-Cell sequencing
- ❑ Main techniques and rationales for data aggregation and visualization

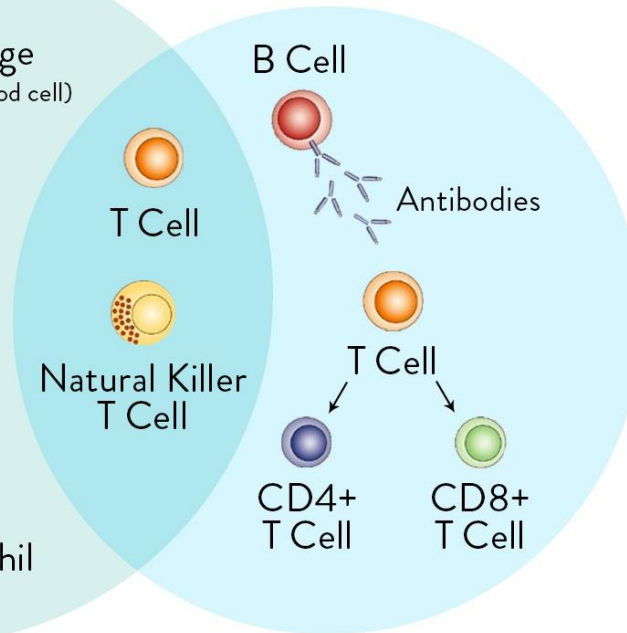
The Immune System



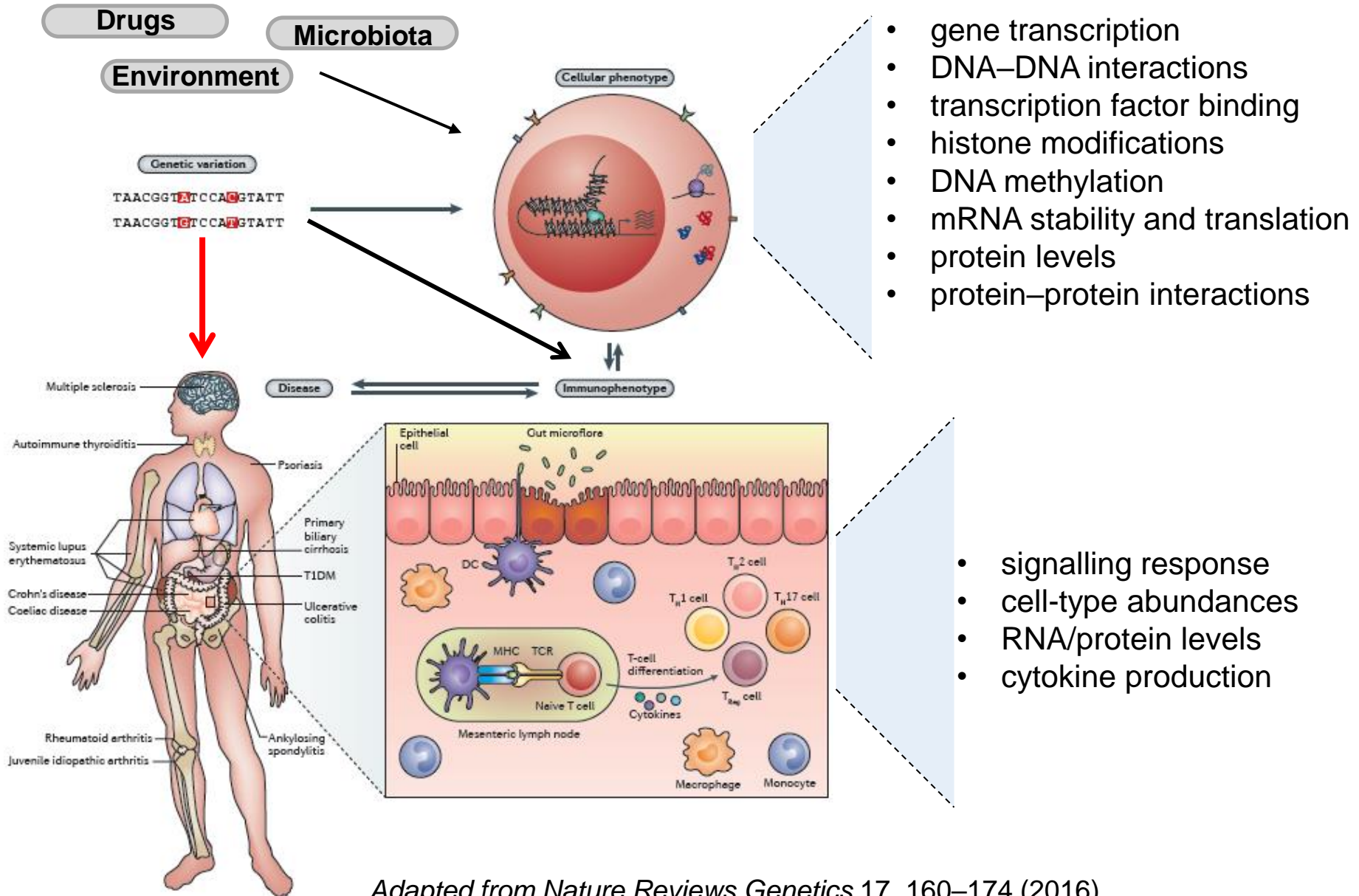
INNATE IMMUNITY (rapid response)



ADAPTIVE IMMUNITY (slow response)



Systems Immunology

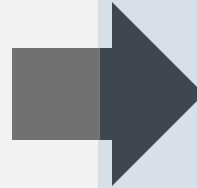


Adapted from *Nature Reviews Genetics* 17, 160–174 (2016)

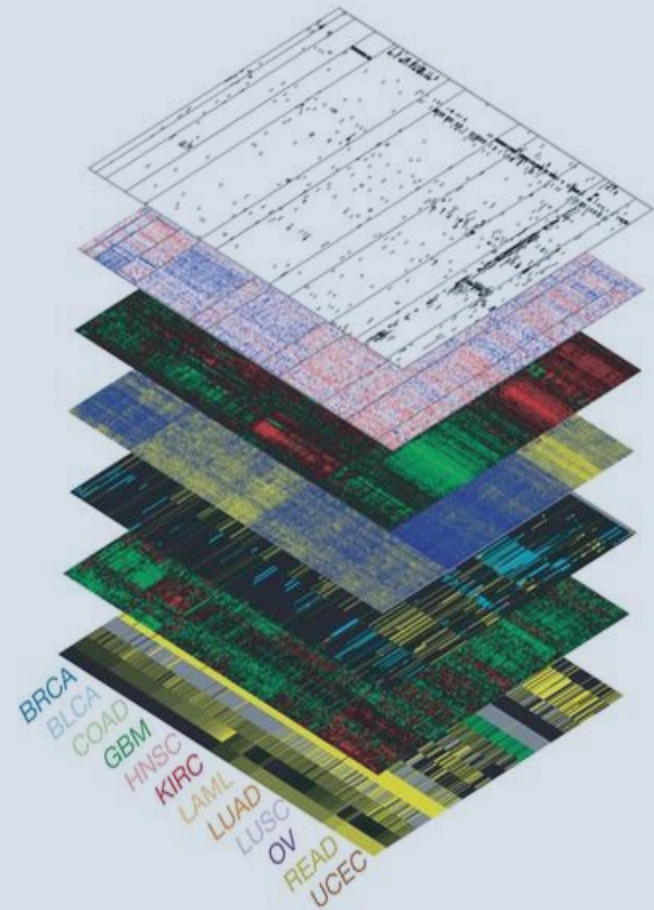
Technologies



Technologies



Data



Technologies

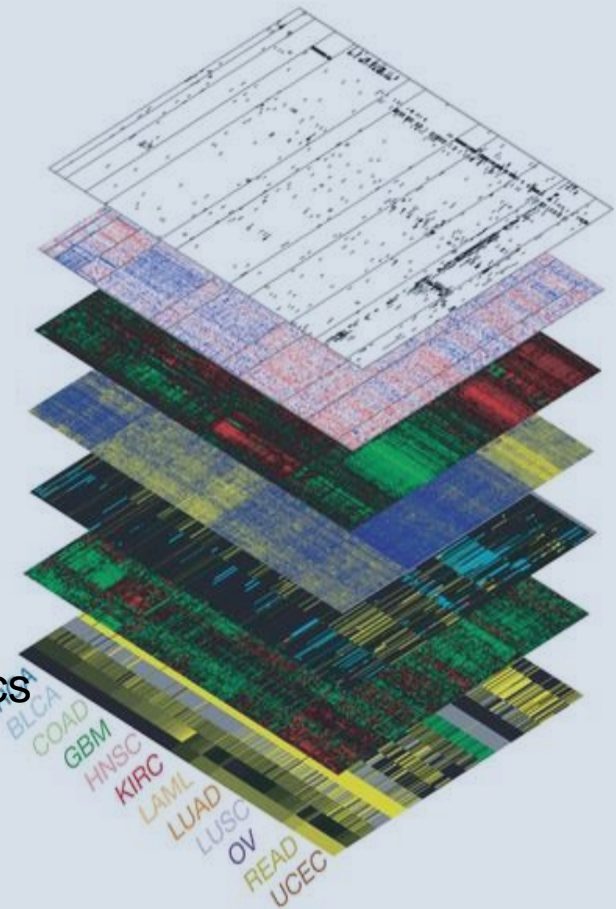


Data

Analysis



Math
Stats,
Bioinformatics
ML/AI

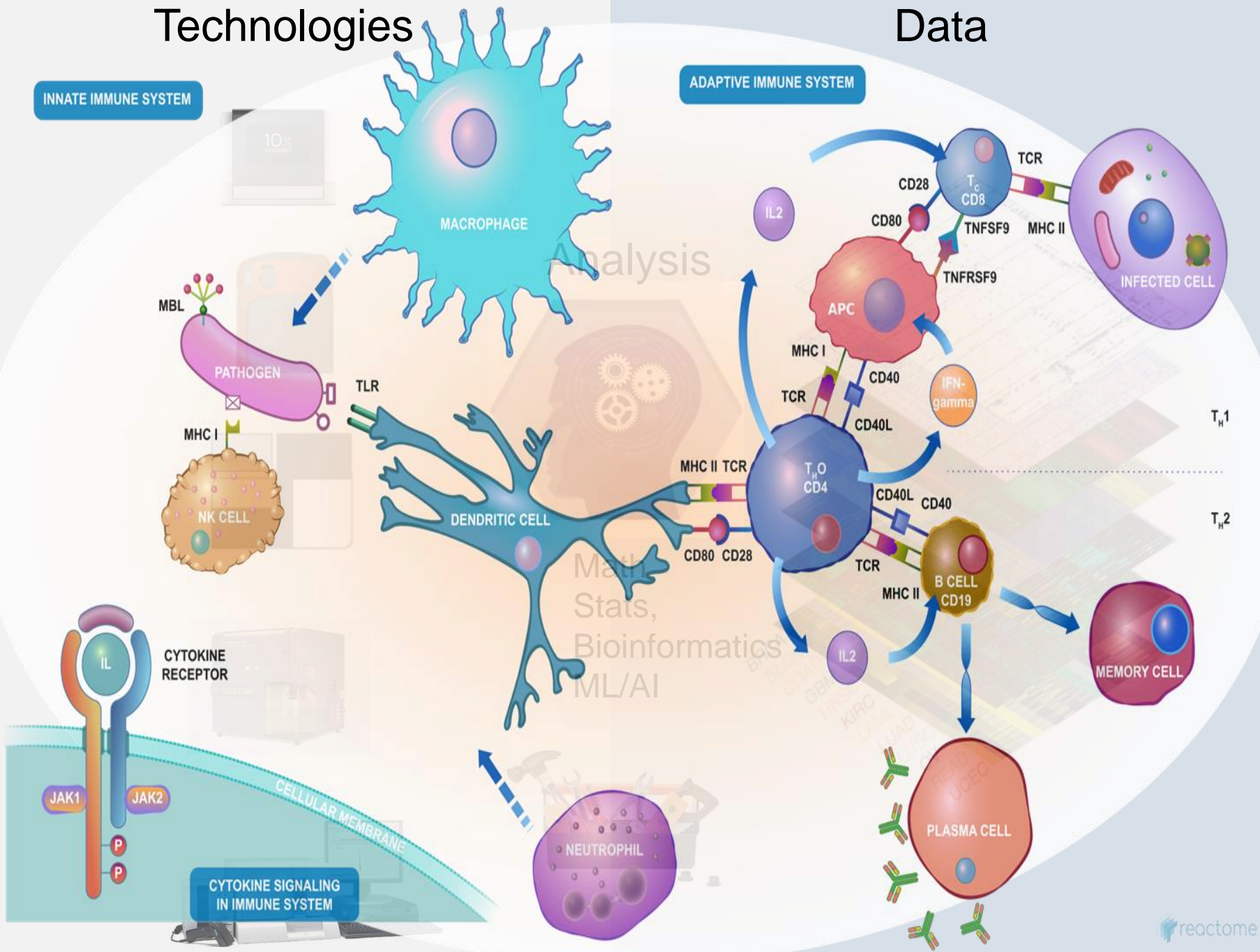


Technologies

Data

INNATE IMMUNE SYSTEM

ADAPTIVE IMMUNE SYSTEM



Immune Cell Profiling: Flow Cytometry and Gene Expression

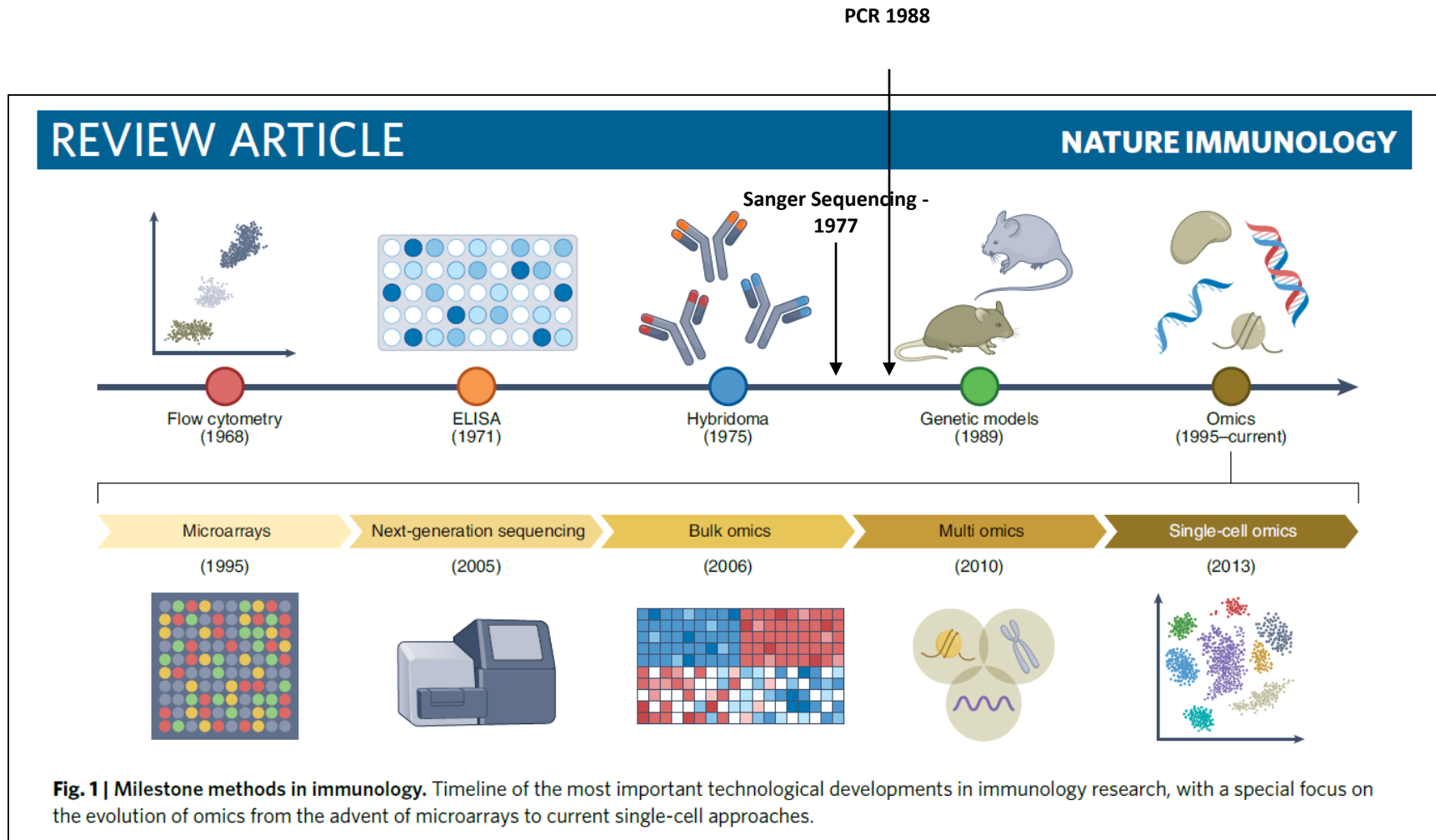
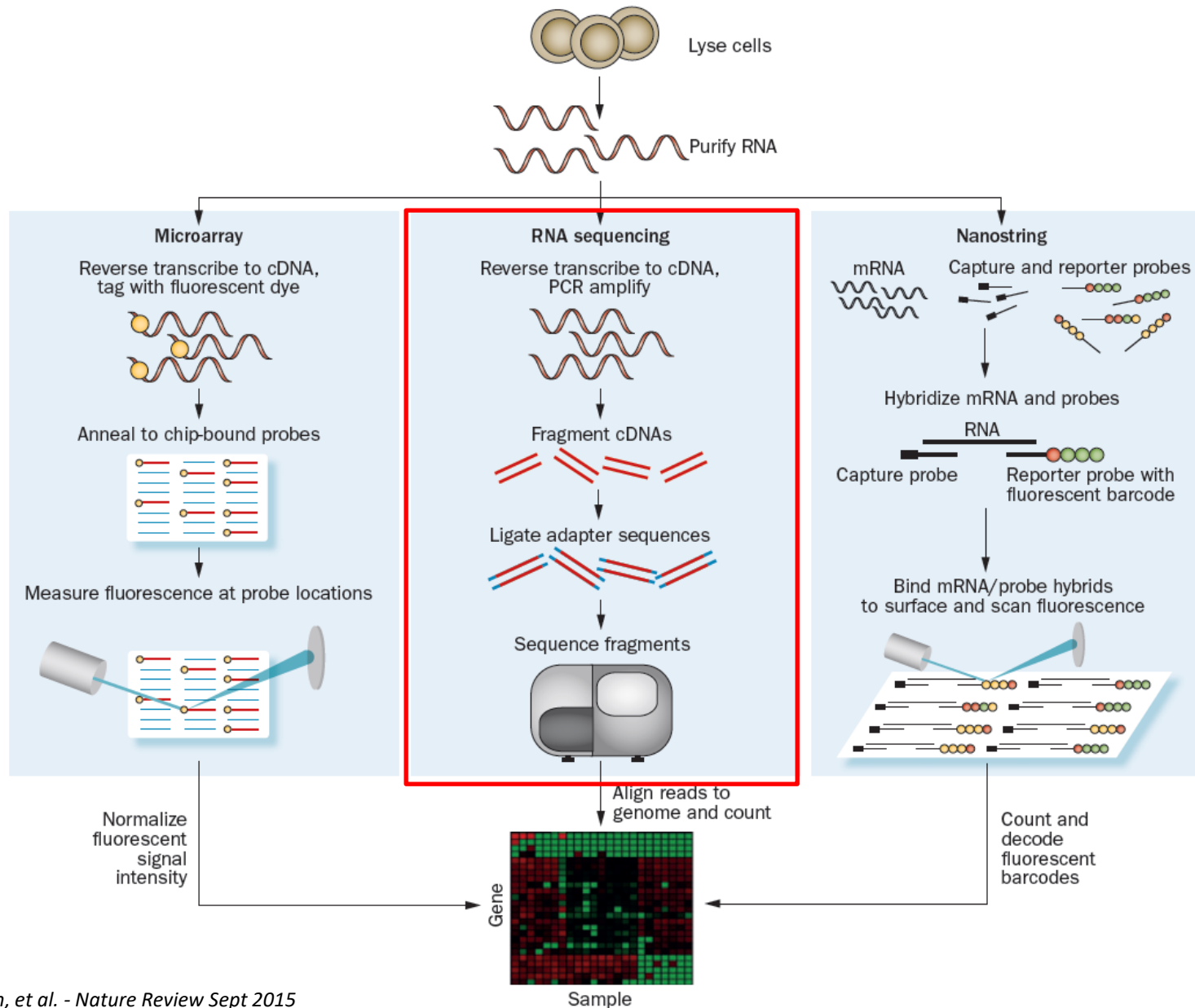


Fig. 1 | Milestone methods in immunology. Timeline of the most important technological developments in immunology research, with a special focus on the evolution of omics from the advent of microarrays to current single-cell approaches.

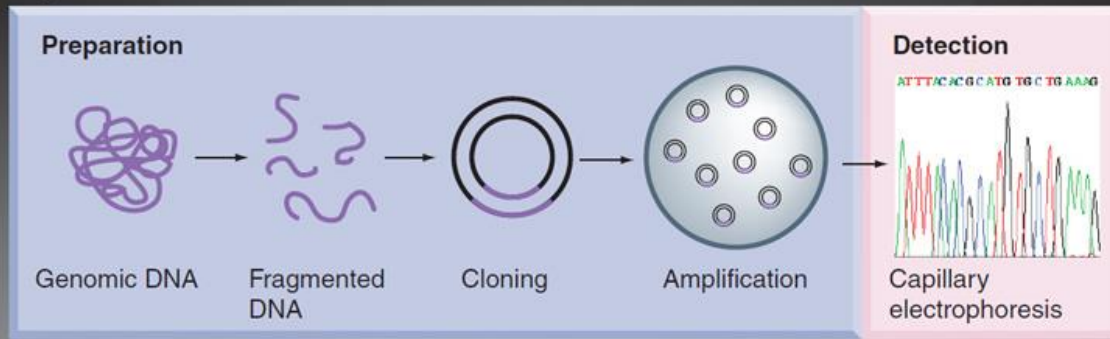
Nature Immunology volume 23, pages1412–1423 (2022)

Gene Expression in "Bulk" Samples

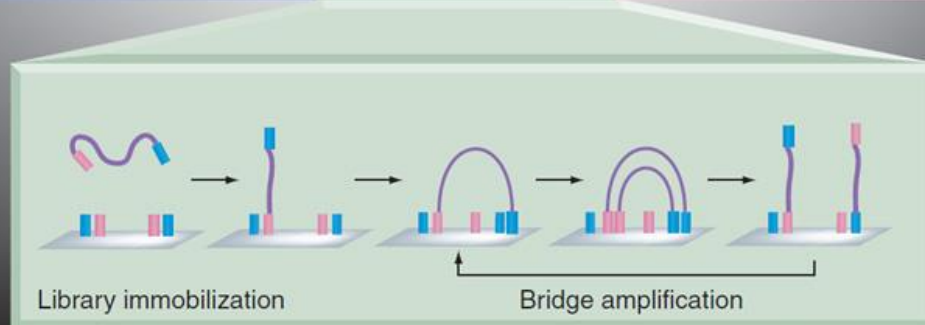
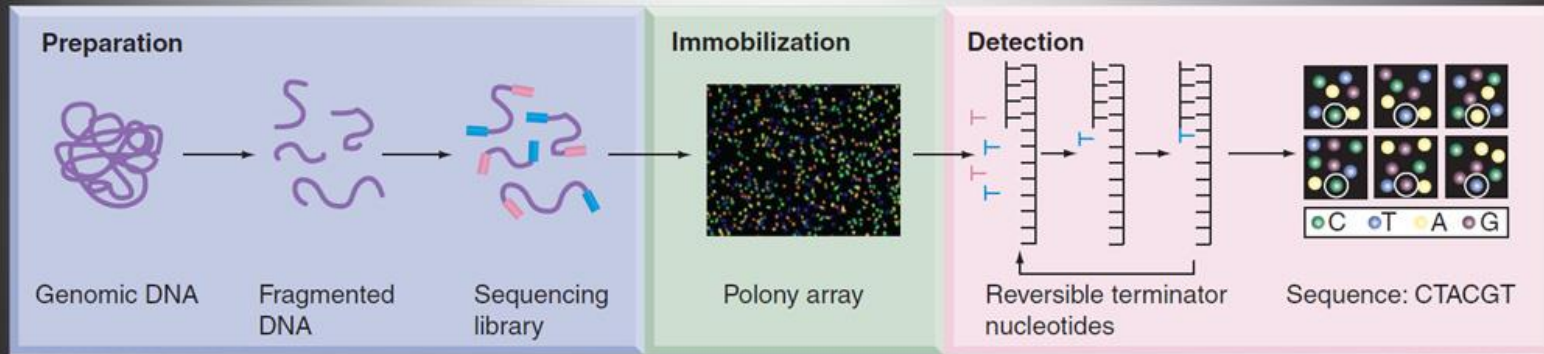


Next Generation Sequencing – The Illumina Platform

Sanger sequencing

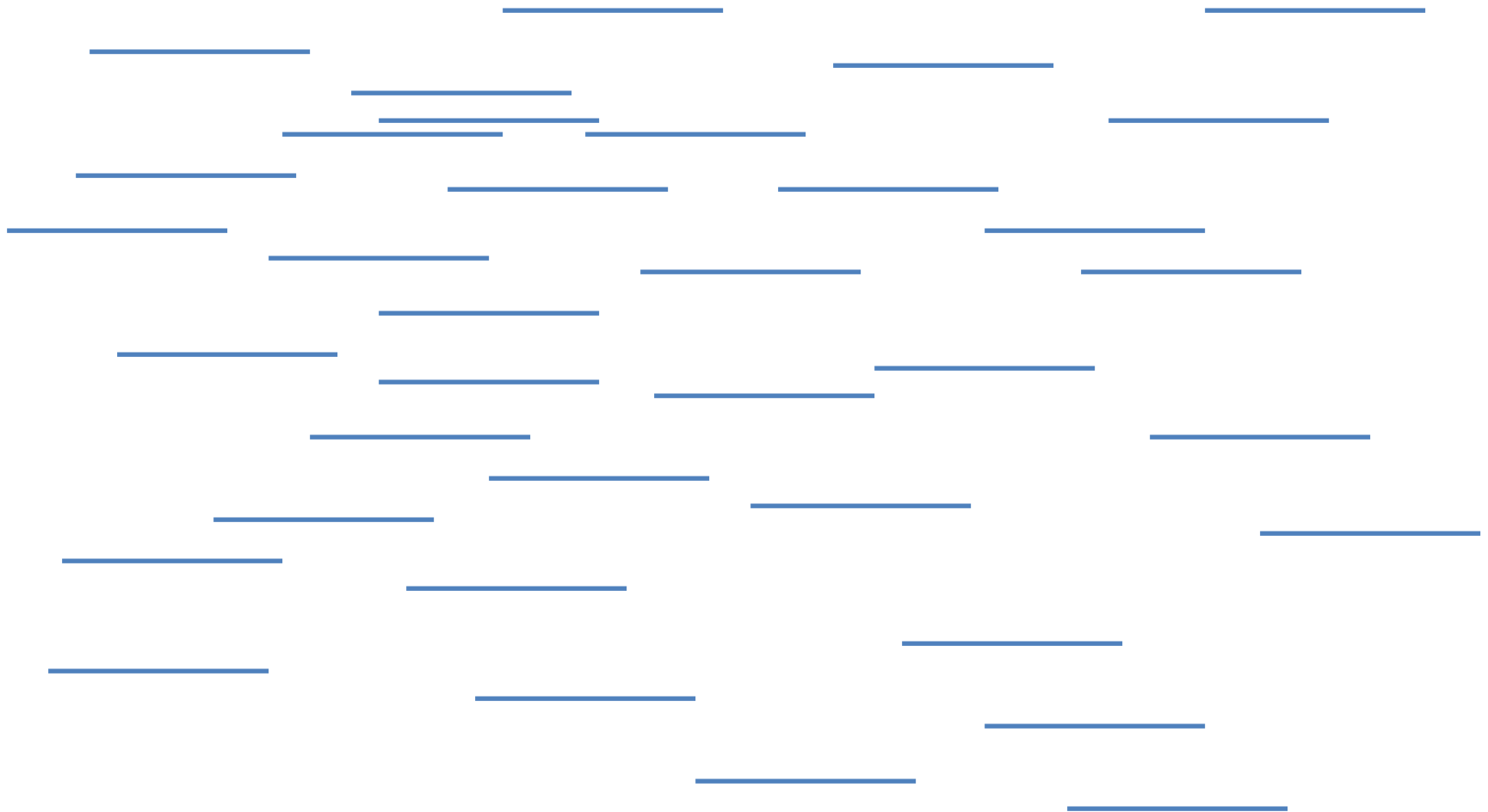


Next-generation sequencing



Lots of sequenced reads...

Illumina NovaSeq 6000: up to 20 billion reads, 3.000Gb data, less than 2 days



What do we do with them ?

Reads

denovo Assembly

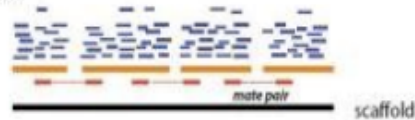
Find overlapping reads



Assemble reads into contigs



Join contigs into scaffolds using mate pairs



Join scaffolds into "finished" sequence

Sequencing of a new organism
Meta-Genomics
Reconstructing cancer genomes

Reference based analysis

Analysis based on a known reference genome

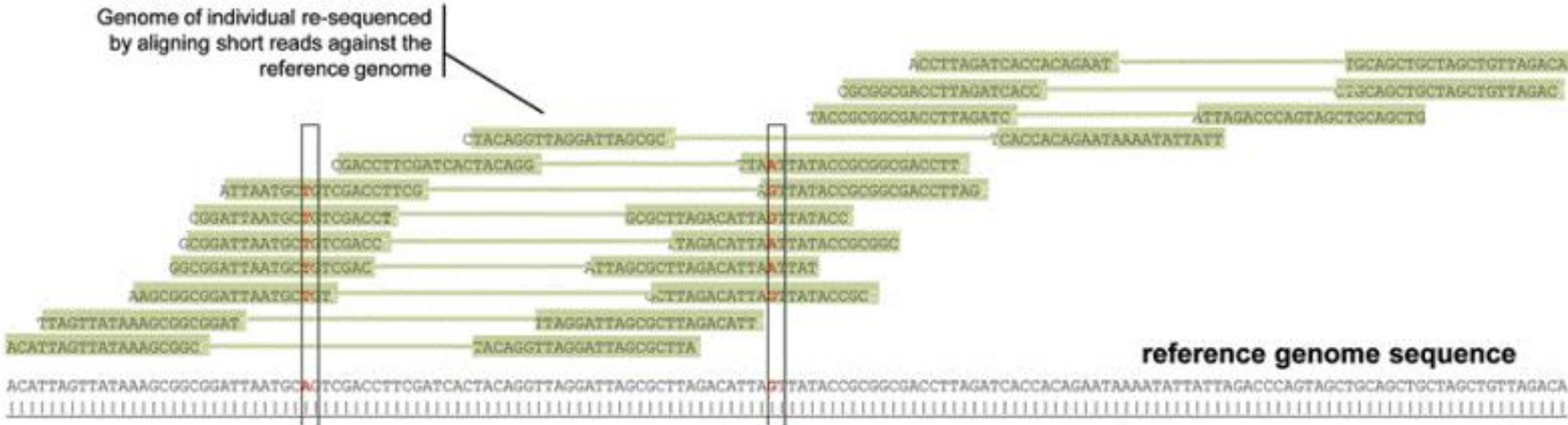
TGACATGCTGTGATGCCCA **CAGTCG** TAGATCGTGGATTCACACAGCTGACAGTA **GACATG** ACA

CAGTCG

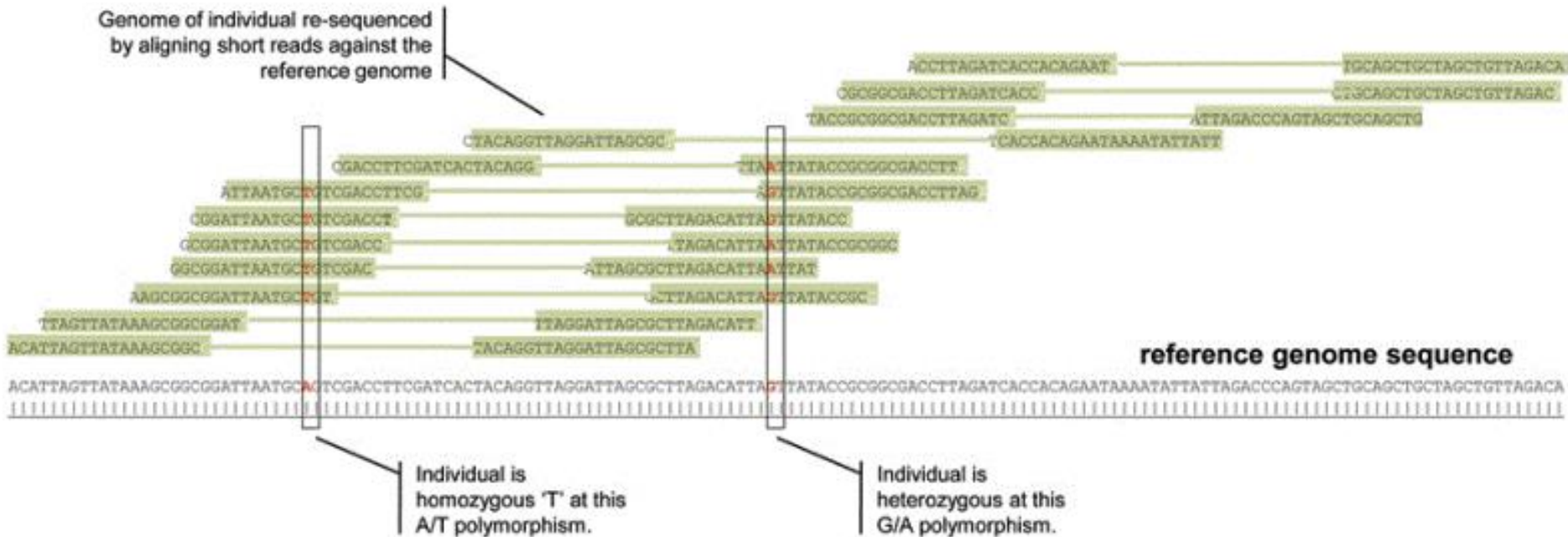
GACATG

Organism specific experiments
Annotating functional elements

Reads Alignment

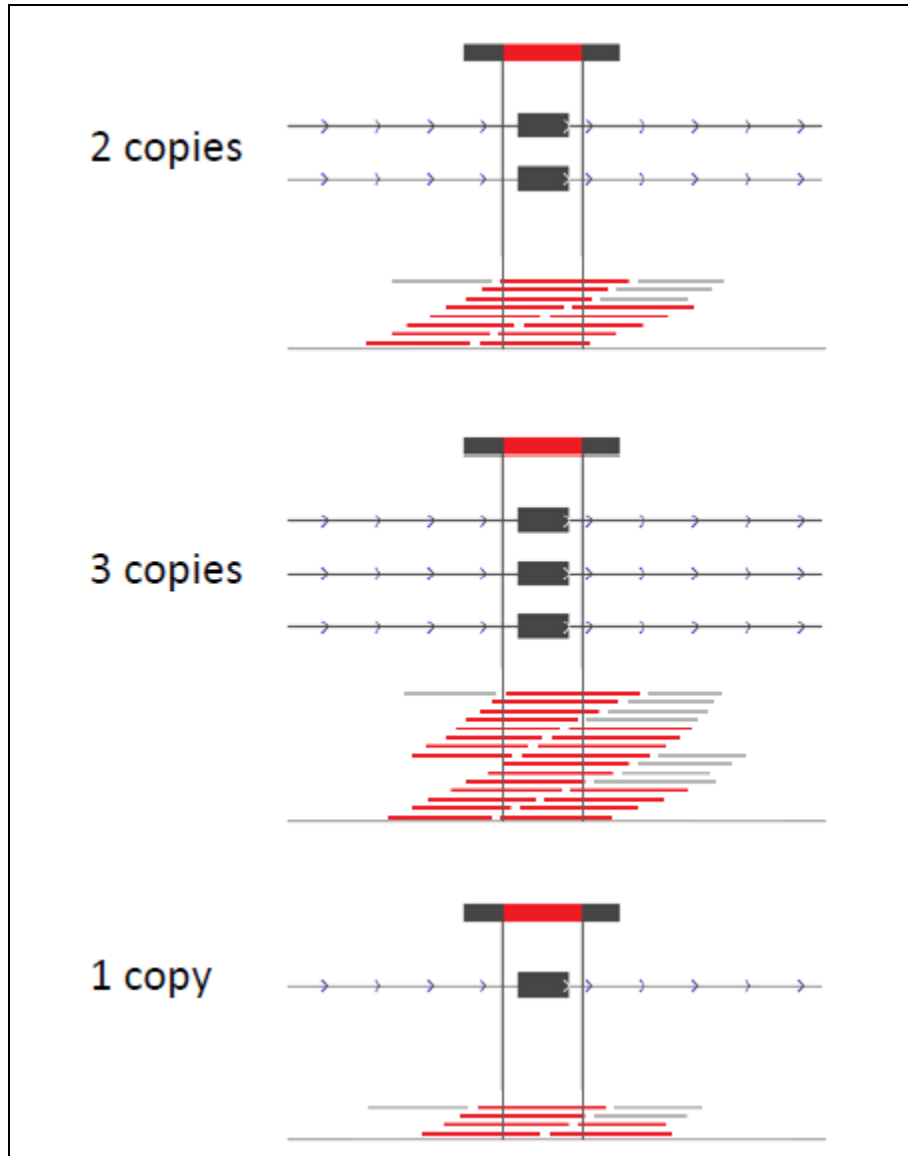


Genome Sequencing: Variant Calling



Differences between aligned reads and reference genome can be identified → variants

Genome Sequencing: Copy Numbers



- The number of aligned reads on a given region is proportional to the number of starting DNA copies of that region
- This information can be used to infer DNA copy number variations (CNVs)

RNA Sequencing

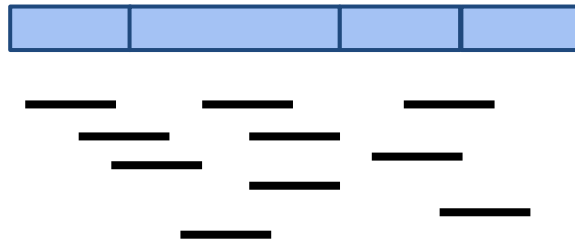
DNA



RNA

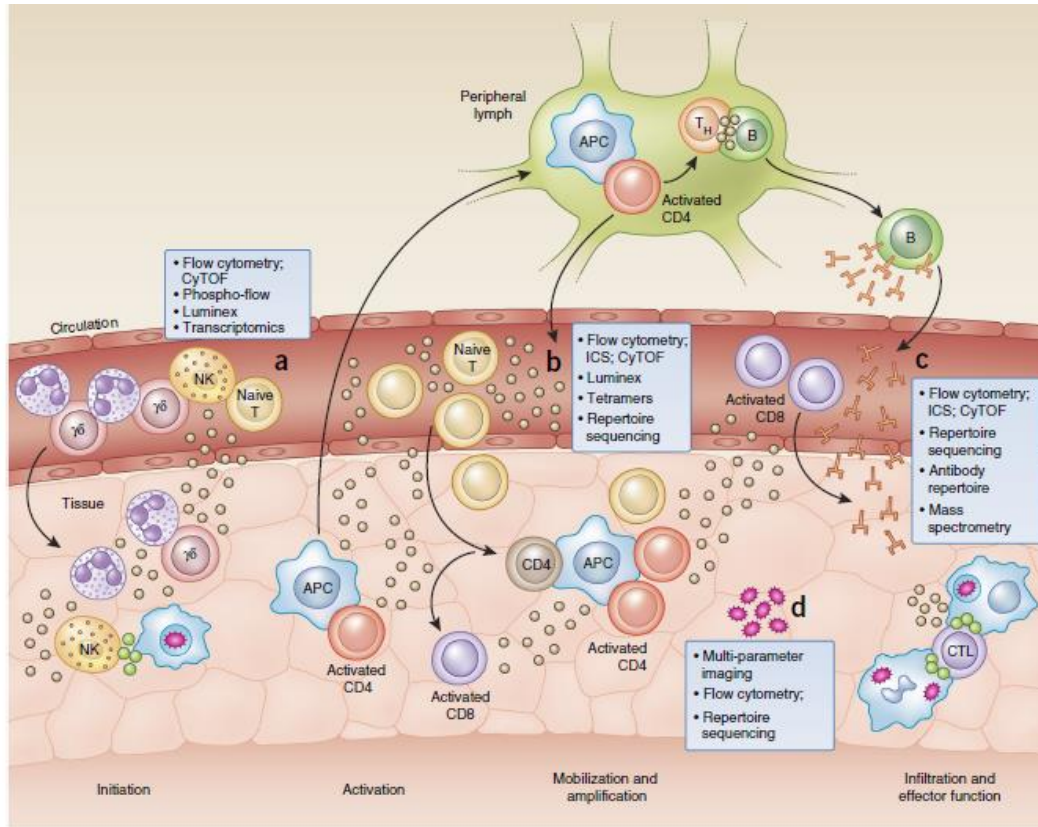


RNA



- The number of aligned reads on a given RNA is proportional to the number of its starting molecules
- This information can be used to infer RNA abundances

Immune Systems Heterogeneity



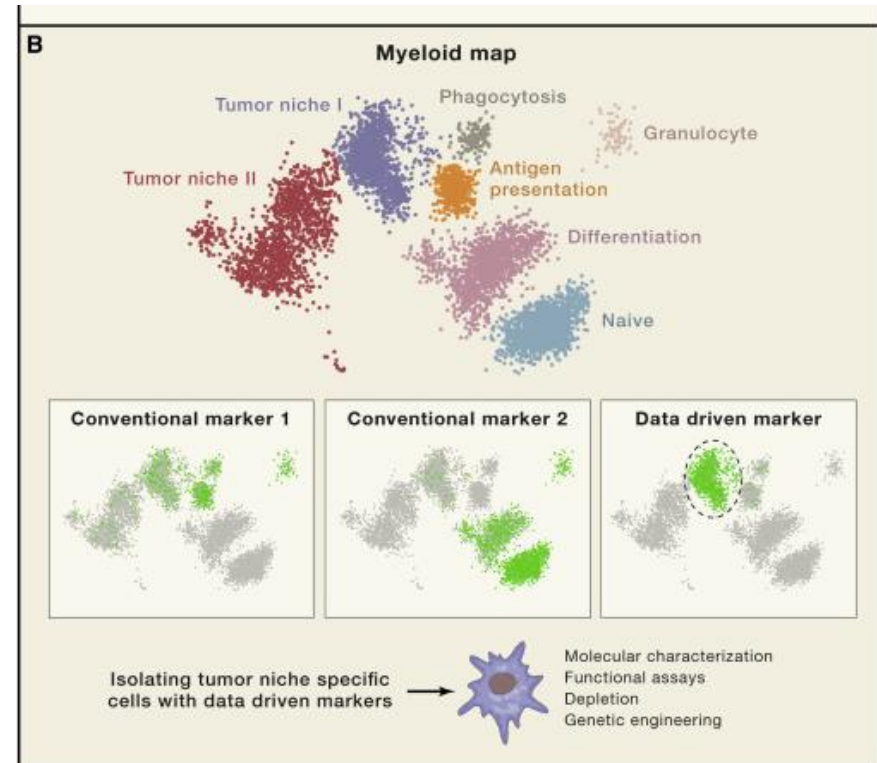
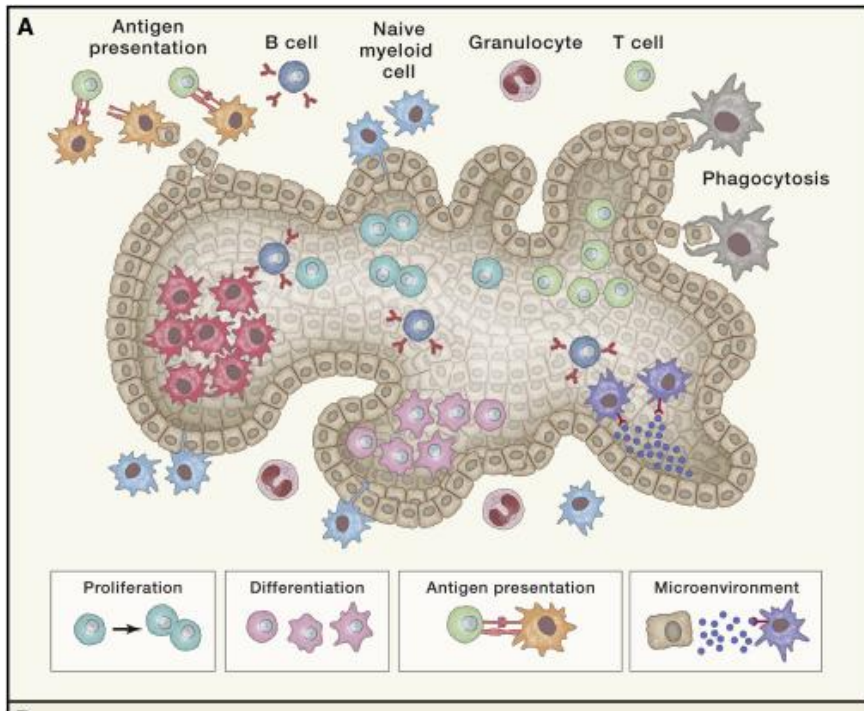
Davis et al. - *Nature Immunology* **18**, 725–732 (2017)



6 JULY 2017 | VOL 547 | NATURE | 27

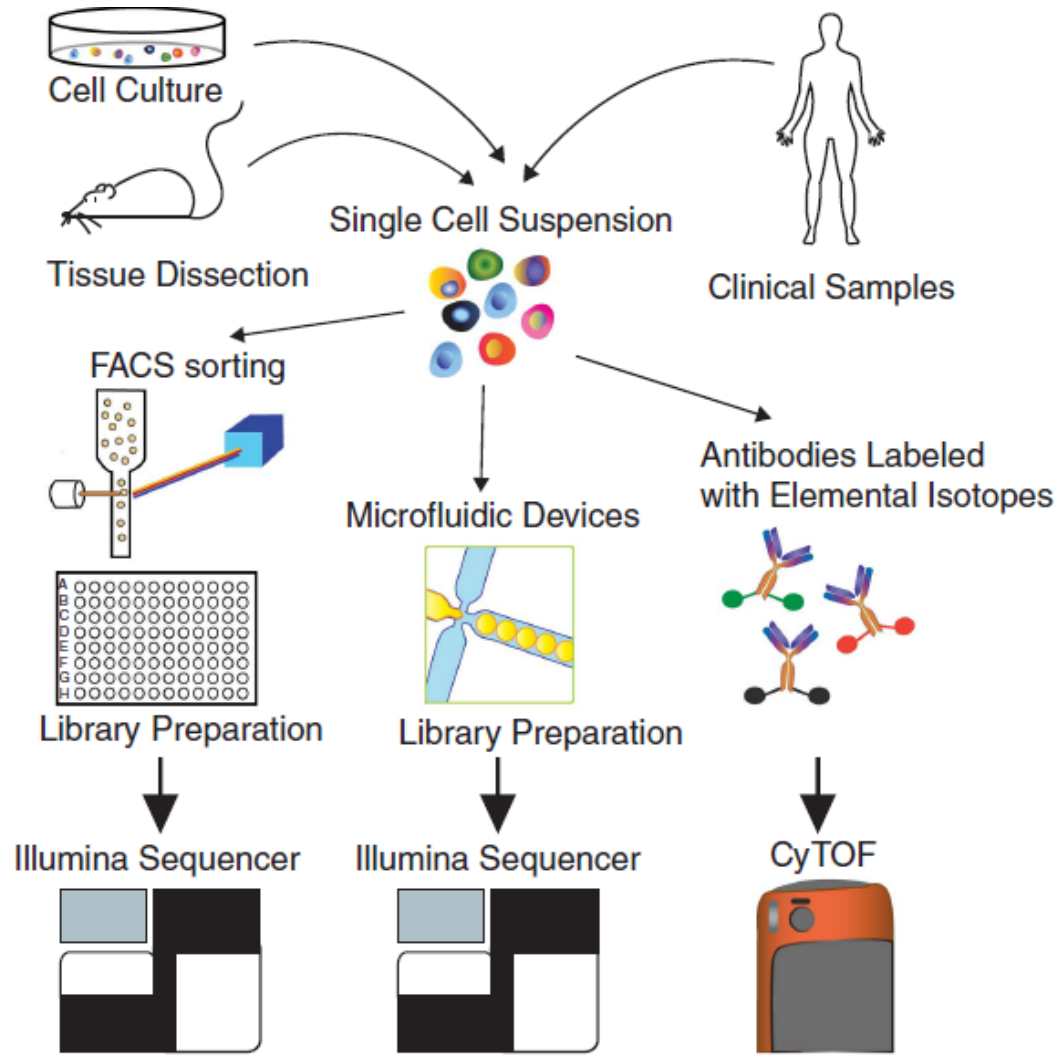
- ❑ Average signals are measured and fails to capture sample heterogeneity
- ❑ Individual cell properties and interplay between different cell subtypes can be captured

Dissecting tissue heterogeneity at single-cell level

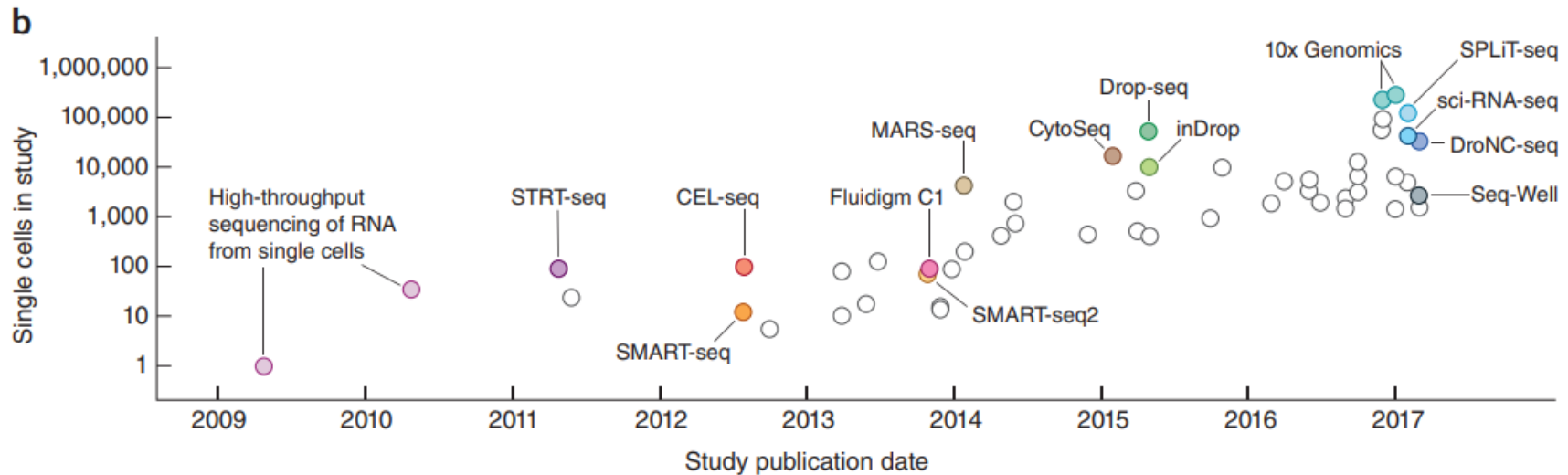
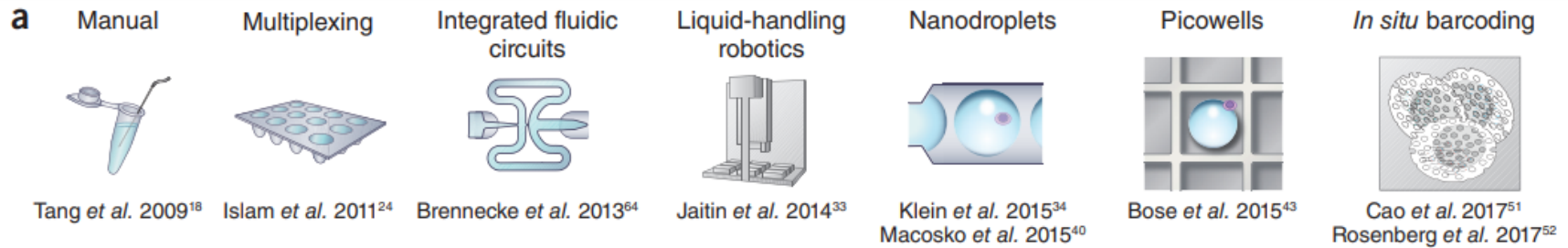


Giladi A, Amit I. Cell. 2018 Jan

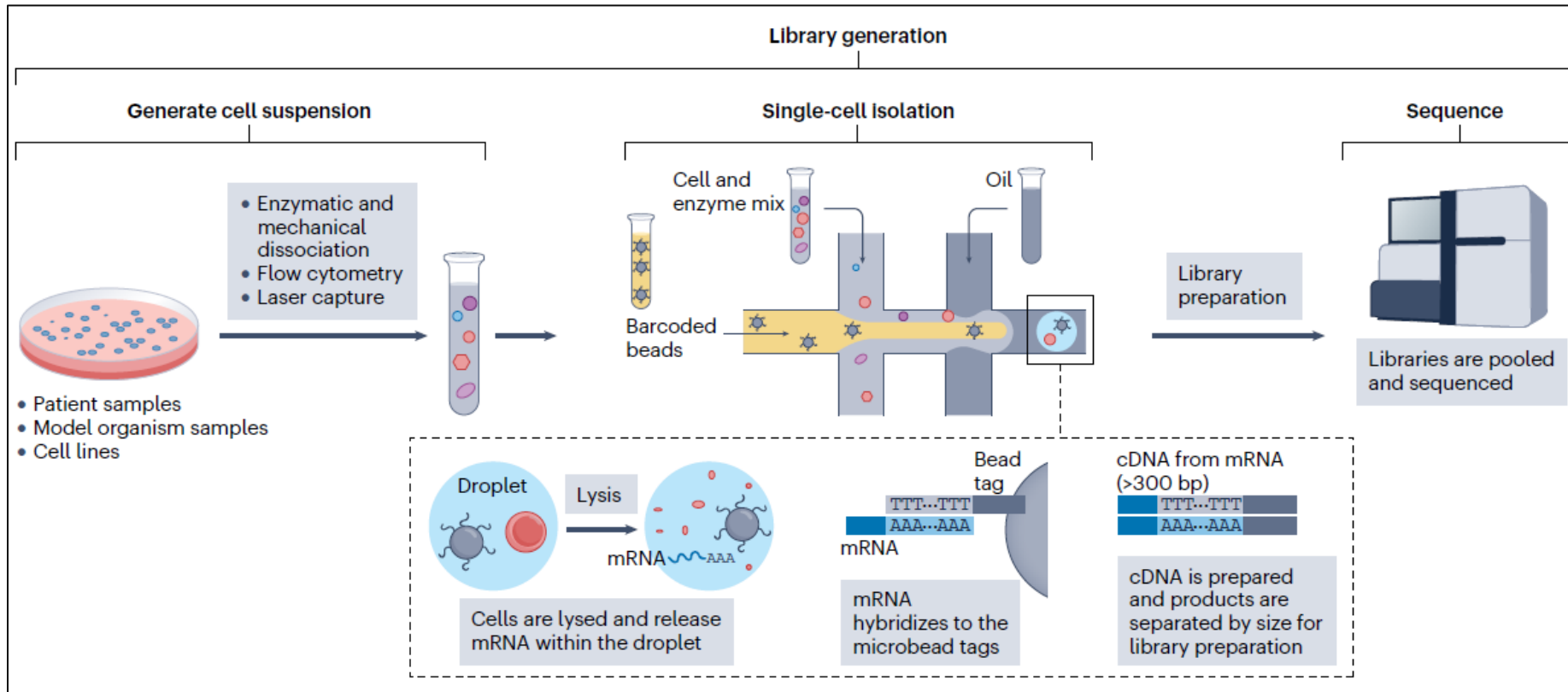
Single-Cell Analysis



Development of Single-Cell Technologies

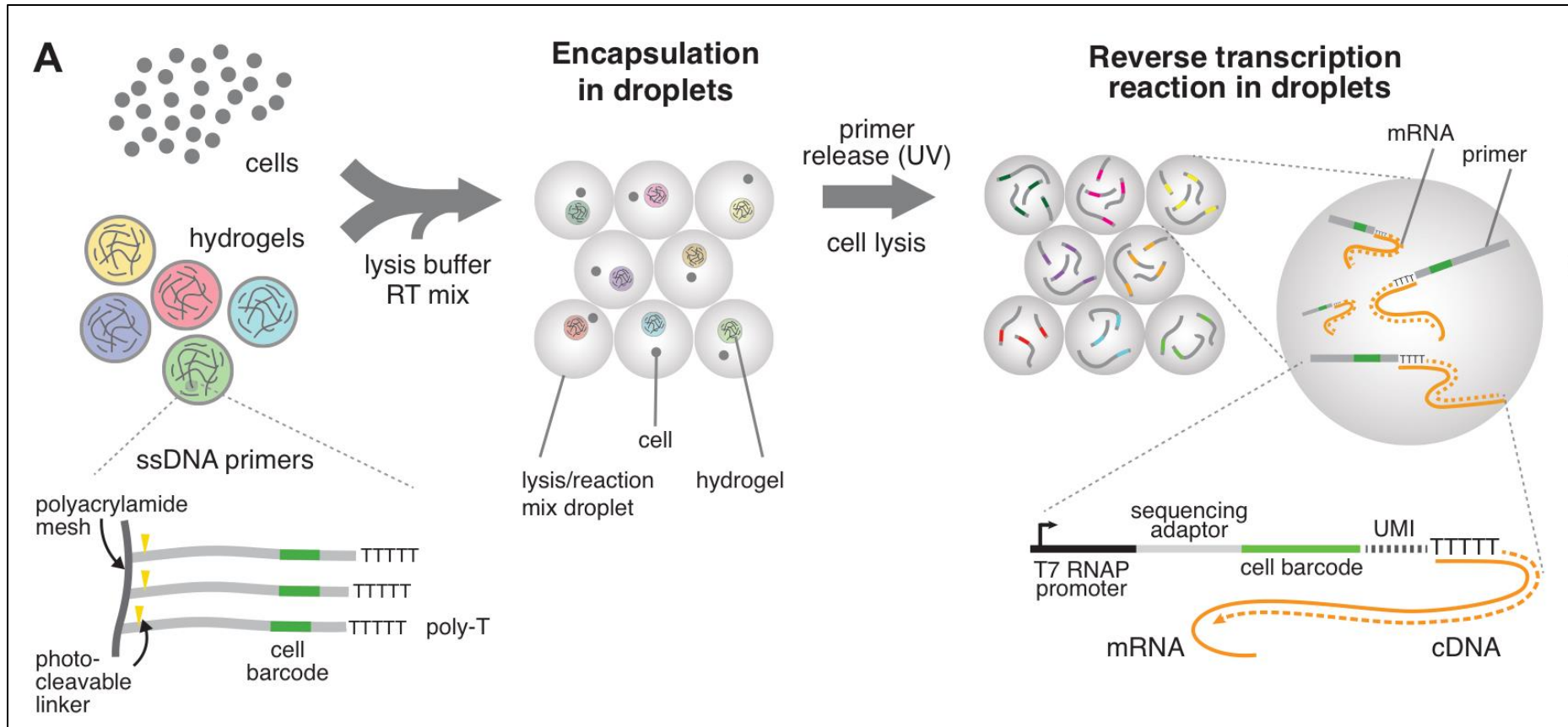


Basic workflow in 10x Chromium



Nature Reviews Drug Discovery volume 22, pages496–520 (2023)

InDrop: Barcoding process

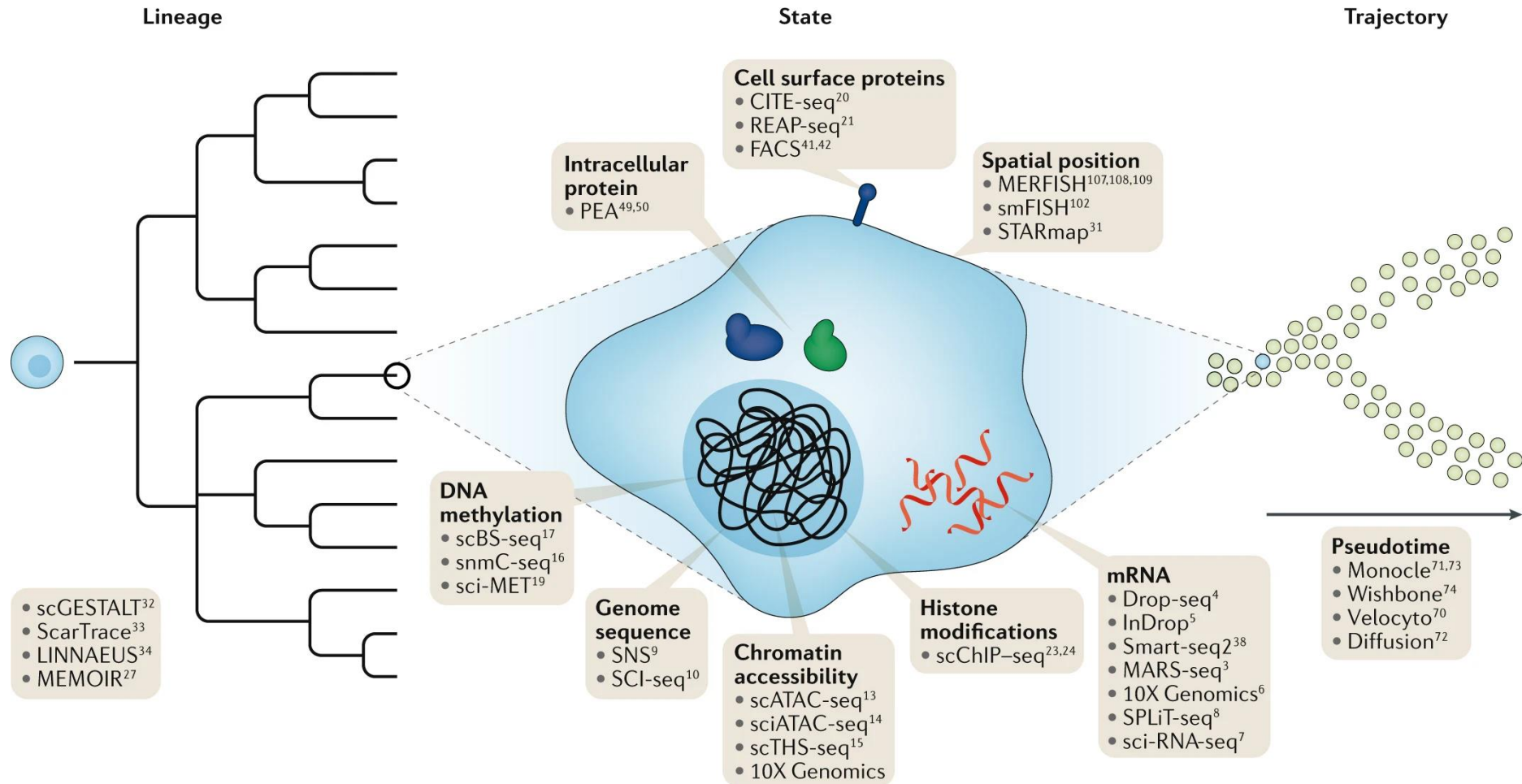


Single-Cell Sequencing Methods

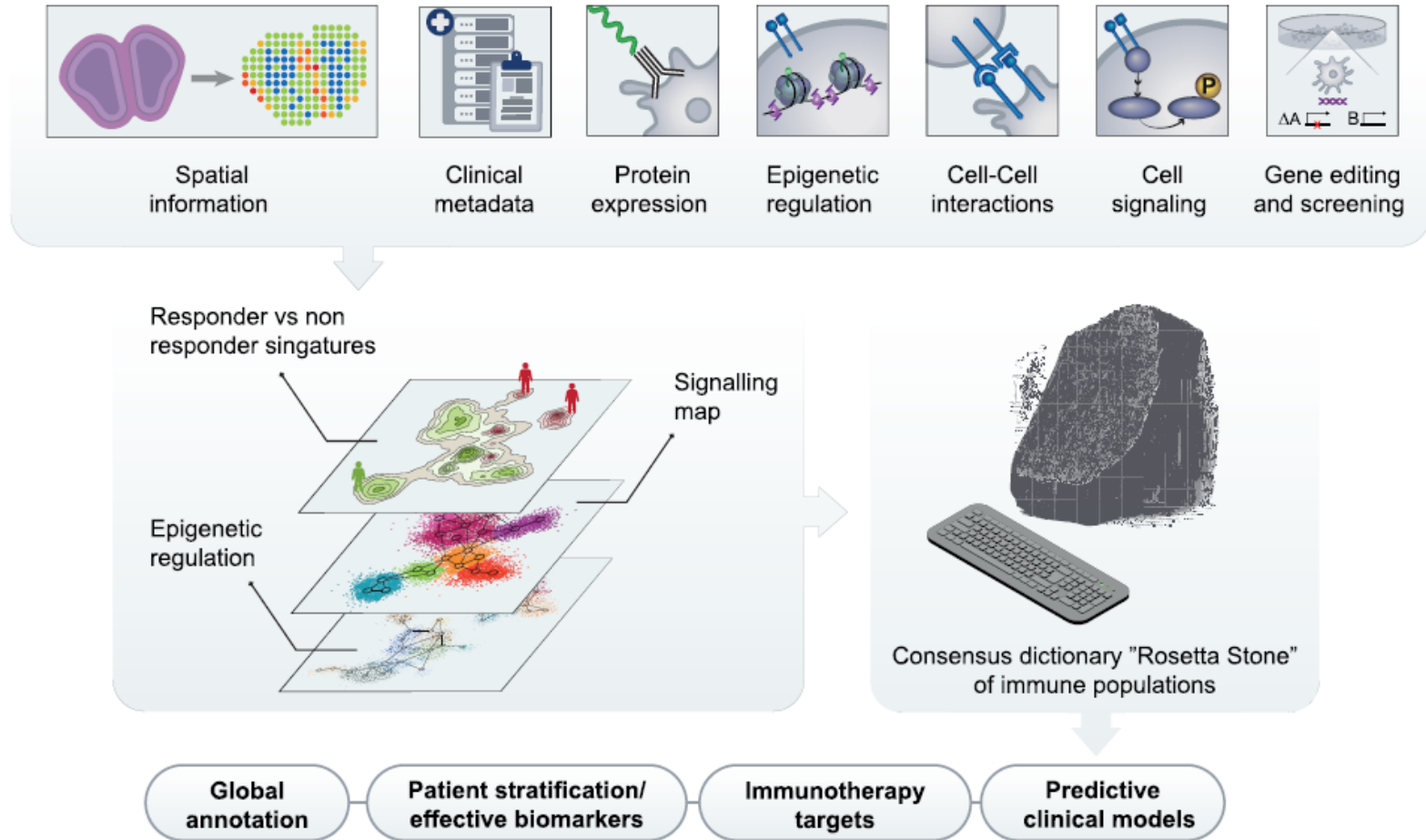
TABLE 1 | Summary of single-cell RNA sequencing methods.

Method	Fluidigm C1 system (SMART-seq)	Fluidigm C1 system (mRNA Seq HT)	SMART-seq2	10X Genomics Chromium system	MARS-seq
cDNA coverage	Full-length	3' counting	Full-length	5'/3' counting	3' counting
UMI	No	No	No	Yes	Yes
Amplification technology	Template switching-based PCR	Template switching-based PCR	Template switching-based PCR	Template switching-based PCR	<i>in vitro</i> transcription
Multiplexing of samples	No	Yes	No	Yes	Yes
Single cell isolation	Fluidigm C1 machine	Fluidigm C1 machine	FACS	10X Genomics Chromium single cell controller	FACS
Cell size limitations	Homogenous size of 5–10, 10–17, or 17–25 μM	Homogenous size of 5–10, 10–17, or 17–25 μM	Independent of cell size	Independent of cell size	Independent of cell size
Required cell numbers per run	≥ 10,000	≥ 10,000	No limitation	≥ 20,000	No limitation
Visual quality control check	Microscope examination	Microscope examination	No	No	No
Long term storage	No, must process immediately	No, must process immediately	Yes	No, must process immediately	Yes
Throughput	Limited by number of machines	Limited by number of machines	Limited by operator efficiency	Up to 8 samples per chip	Process is automated
Cost	++++	+++	++++	+	++
Sample Preparation Scenario 1 (~5000 single cell)	Targeted cell No: 4992 cells 26 rounds of 2 runs (2 C1 machines; concurrent) ~26 weeks	Targeted cell No: 4800 cells 3 rounds of 2 runs (2 C1 machines; concurrent) ~3 weeks	Targeted cell No: 4992 cells 26 rounds of 2 96-well plates ~26 weeks	Targeted cell No: 5000 cells 1 run ~2–3 days	Targeted cell No: 4992 cells 13 runs of 1 384-well plate ~7 weeks
Sample Preparation Scenario 2 (~96 single cell)	Targeted cell No: 96 cells 1 run (1 C1 machine) ~1 week	Targeted cell No: Minimum 800 cell 1 run (1 C1 machine) ~1 week	Targeted cell No: 96 cells 1 run of 96-well plates ~1 week	Targeted cell No: Minimum 500 cells 1 run ~2–3 days	Targeted cell No: 96 cells 1 run of 384-well plate ~2–3 days

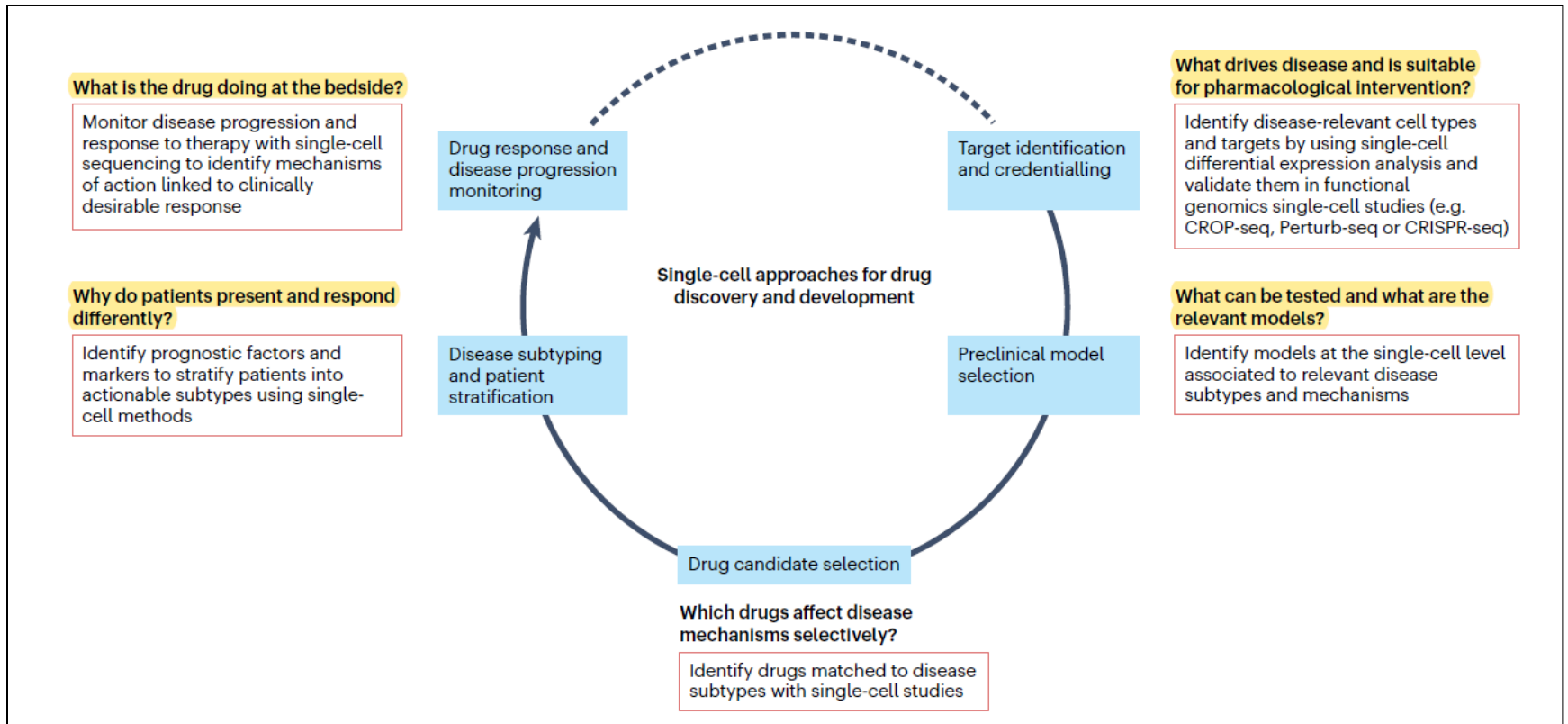
Single-cell multi-omics



Systems Biology Through Single-Cell Multi-omics



Applications T Drug Discovery



Applications To Drug Discovery

a Standard high-throughput chemical screens

- 100k-1M compounds
- Typically one compound dose tested on one condition



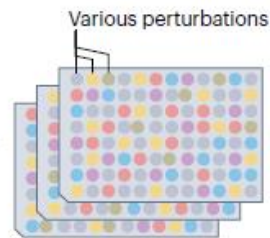
List of most active compounds

Dose-response and other studies

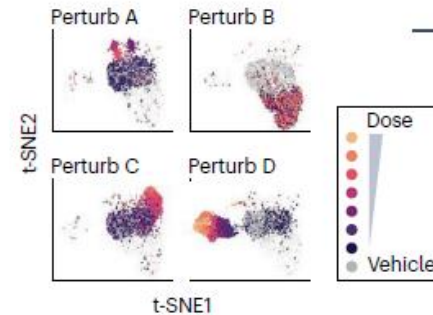
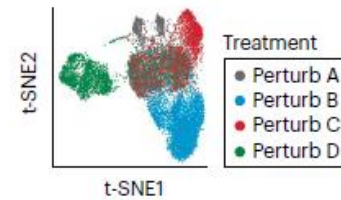
Hits suitable for drug discovery

b Single-cell high-throughput chemical screens

- 100-1,000 drugs
- Typically 5-10 drug doses tested on several cell lines (-100k-1M single cells in all)

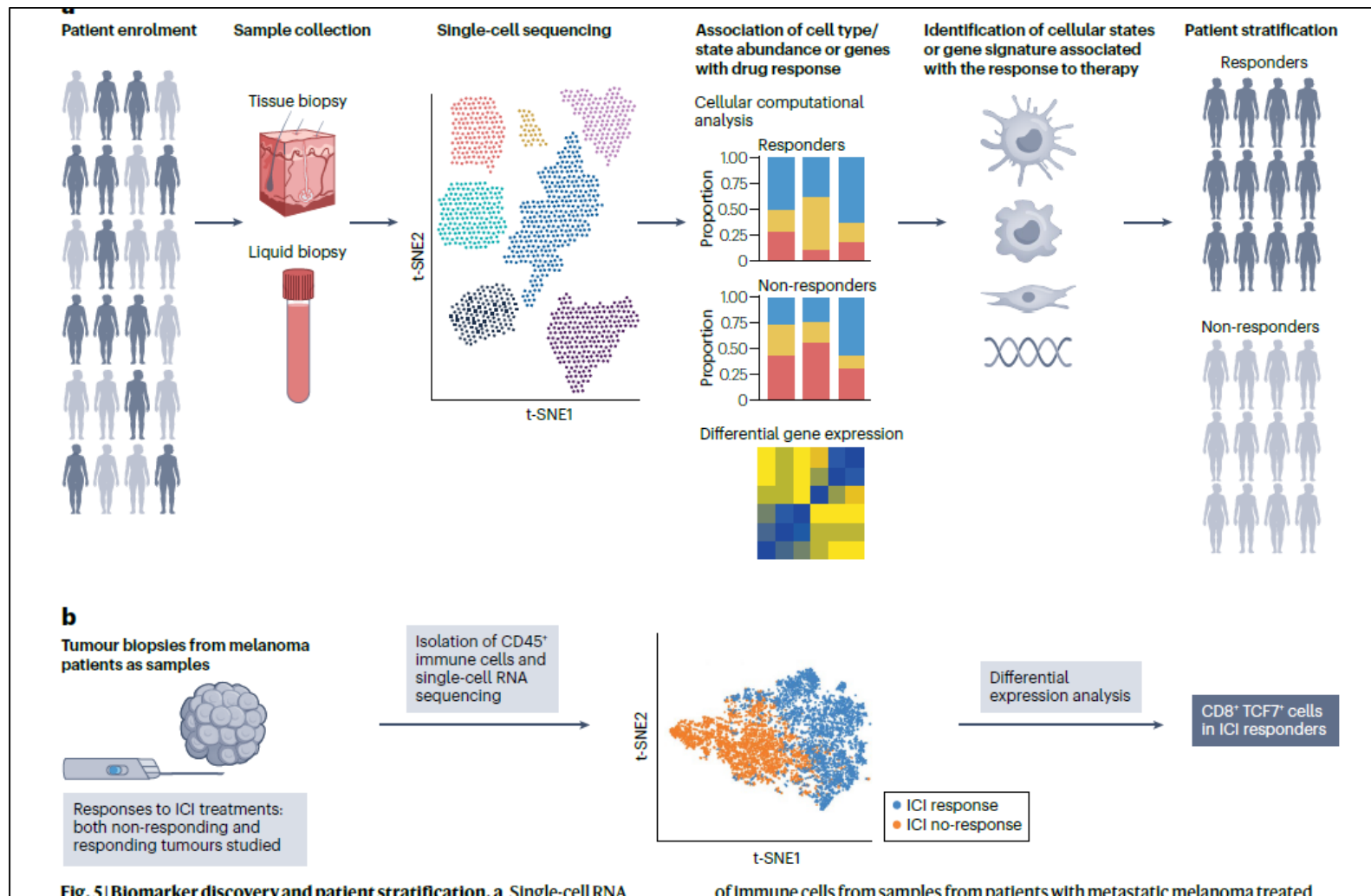


Pooling
scRNA-seq



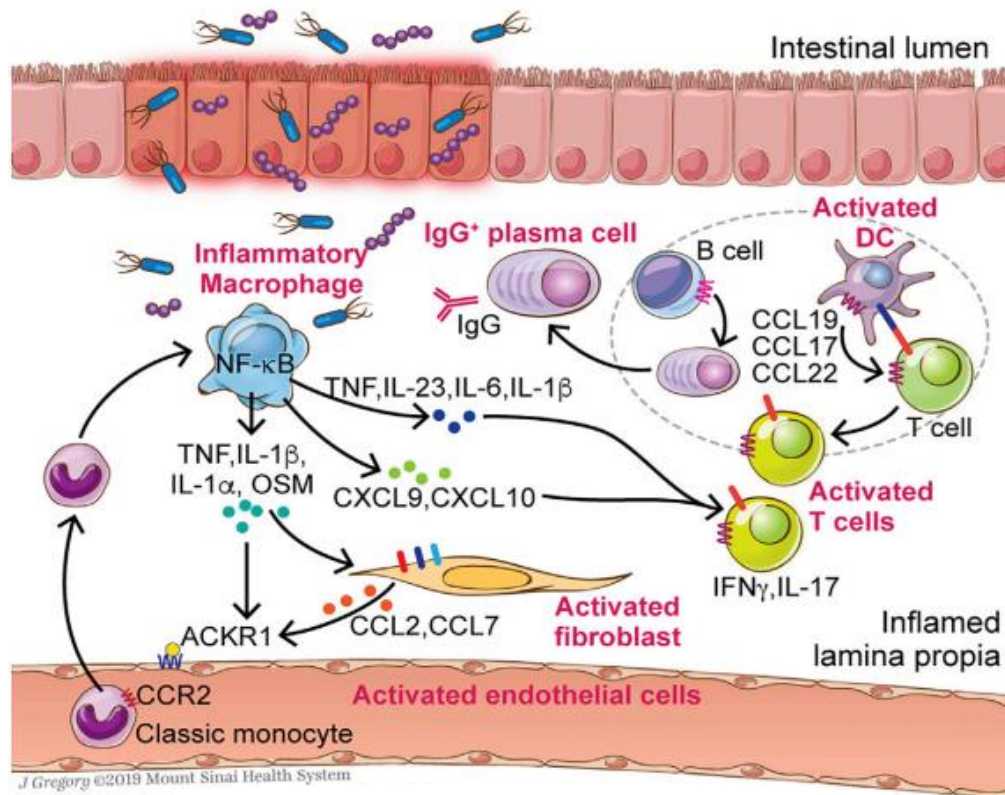
Hints on drug's MoA

Single-Cell for Biomarker Discovery and Patient Stratification



Single-Cell-Based Biomarker of response in anti-TNF therapy (CD)

GIMATS^{high} Module in CD inflamed ileums associates with resistance to anti-TNF responder CD patients

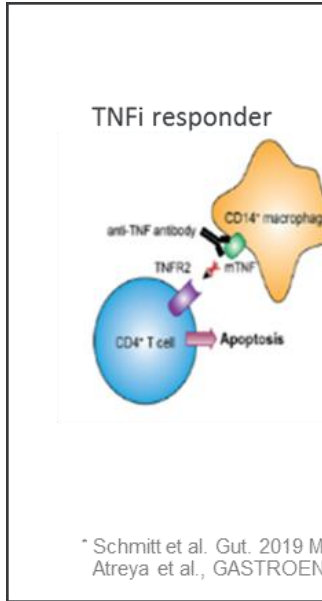


□ GIMATS=IgG plasma cells, inflammatory mononuclear phagocytes, activated T cells, and stromal cells

□ Existence of two qualitatively distinct subsets of disease, with distinct responses to anti-TNF therapy.

3
2

Understanding dual targeting: TNF/IL23 in Chron's Disease



Results of Novel Clinical Study Show Adults with Moderately to Severely Active Ulcerative Colitis Achieved Higher Rates of Clinical Response, Clinical Remission, and Endoscopic Improvement at 12 Weeks with Guselkumab and Golimumab Combination Therapy Versus Either Monotherapy Alone

The VEGA Phase 2a proof-of-concept study shows 83.1 percent of patients who received combination therapy achieved the primary endpoint of clinical response and 36.6 percent of patients achieved clinical remission at week 12

The VEGA study represents a first-of-its-kind biologic combination assessment of an interleukin (IL)-23p19 subunit antagonist with a tumor necrosis factor-alpha (TNF α) antagonist in ulcerative colitis



of single-cell
in Phase 2a
py

erative Colitis

ical Study

on therapeutics for patients with autoimmune
will apply its proprietary single-cell
2a clinical trial evaluating the efficacy and

of patients with inflammatory bowel
or how we can apply our integrated platform
data that will fuel Celsius' novel target and

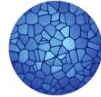
collaboration. Celsius retains the ability to
conomic information, and will further

idly integrate and interrogate the large
fic officer of Celsius. "The longitudinal patient

Working Hypo
responders

non-

Human Cell Atlas



HUMAN
CELL
ATLAS

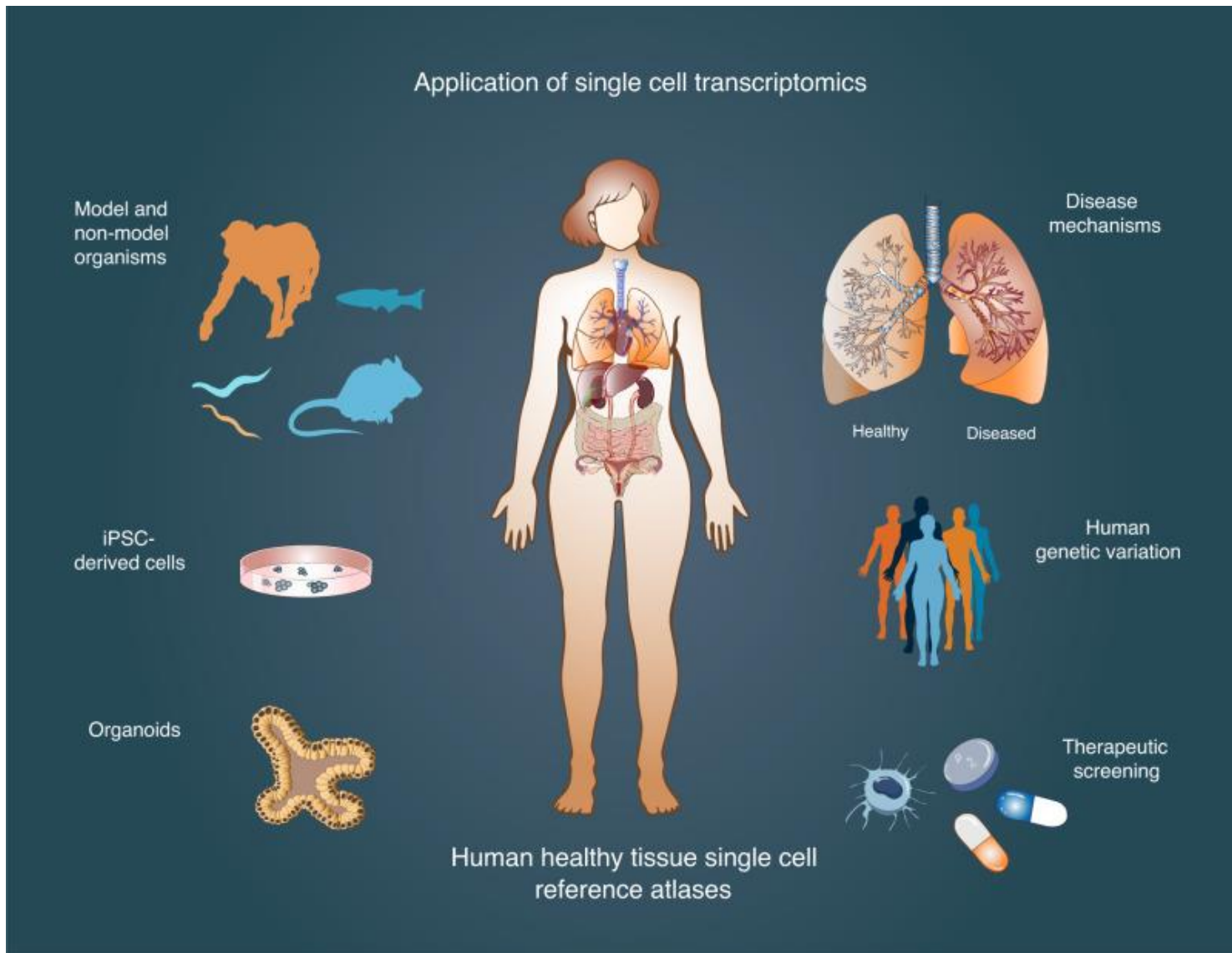
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MISSION

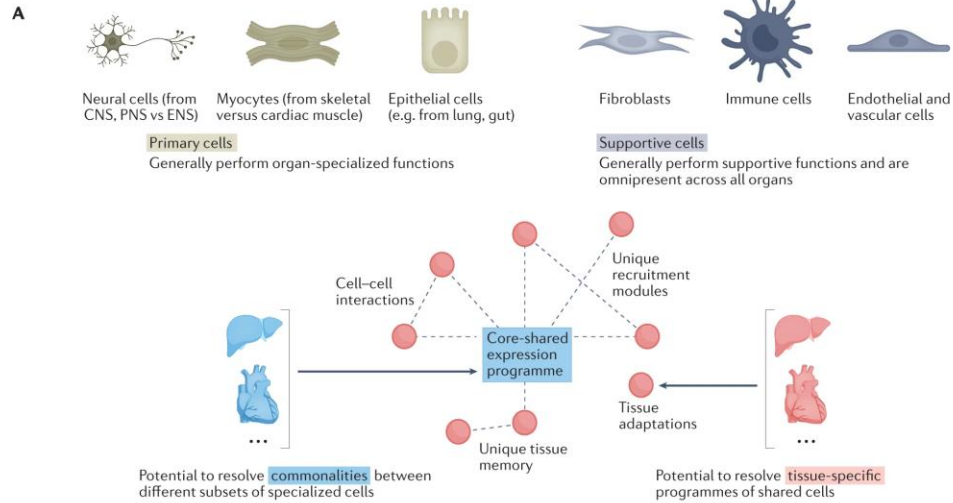
To create comprehensive reference maps of all human cells—the fundamental units of life—as a basis for both understanding human health and diagnosing, monitoring, and treating disease.

<https://www.humancellatlas.org/>

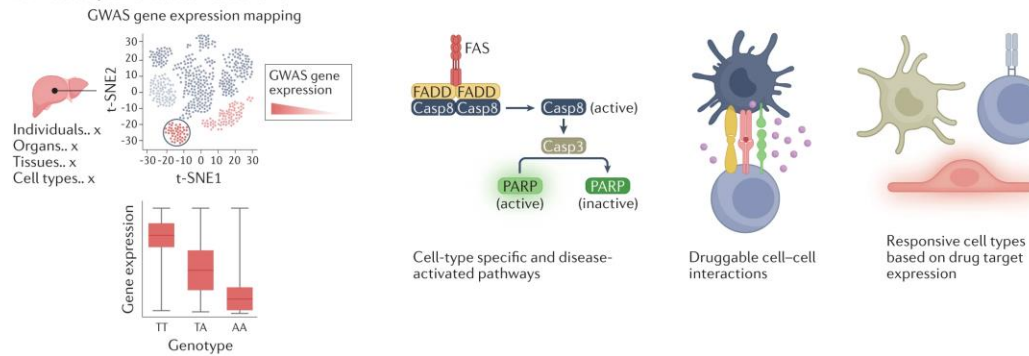
Single-Cell Atlas



Single-Cell Cross-Tissue Comparisons



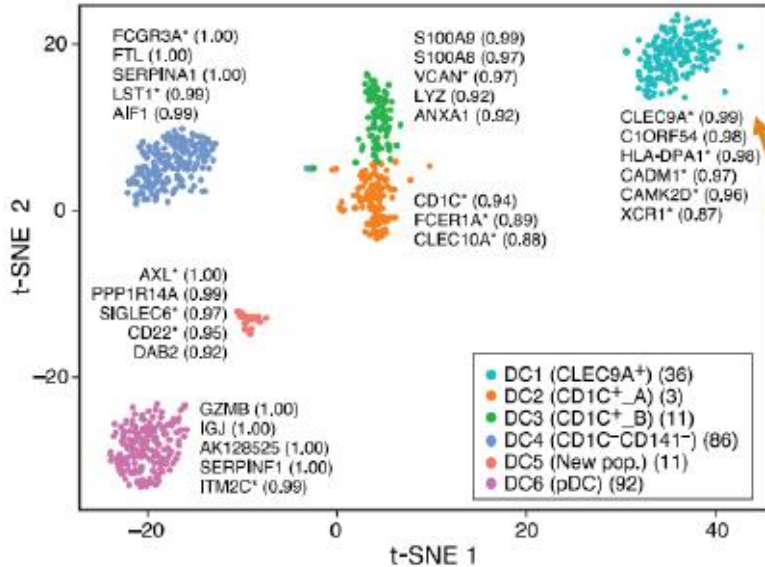
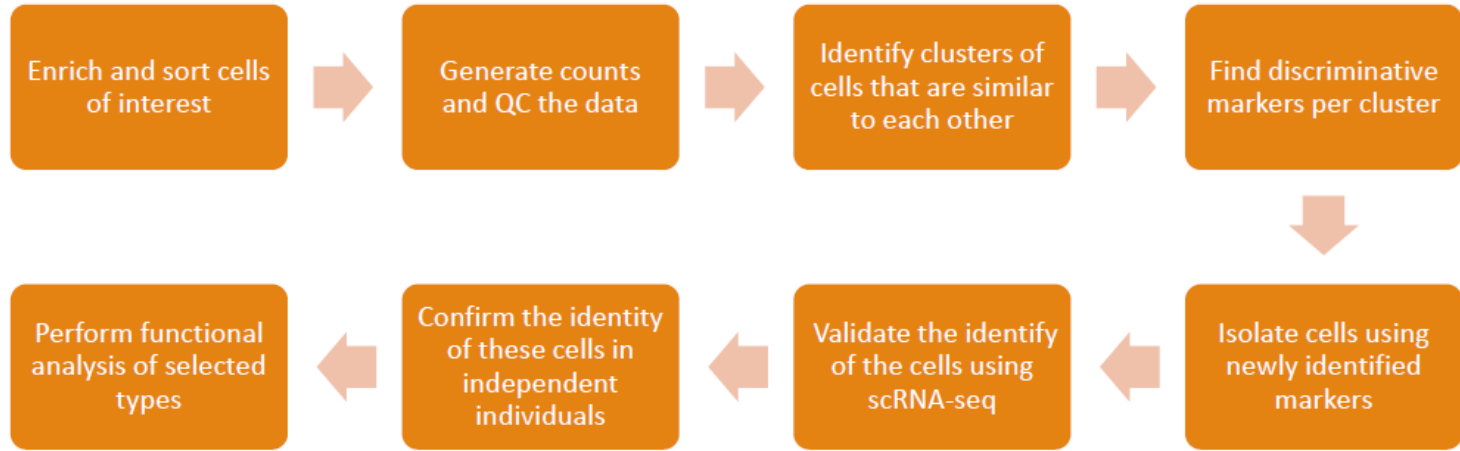
Ba Tissue-specific disease mechanisms



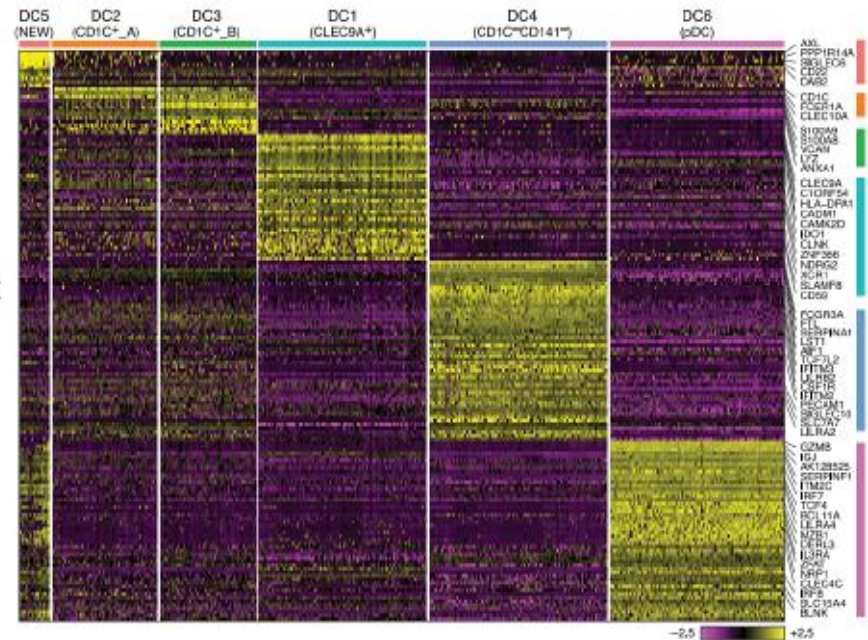
Bb Therapeutic avenues



Immune System Heterogeneity: Reclassification of DCs and monocytes by scRNA-Seq

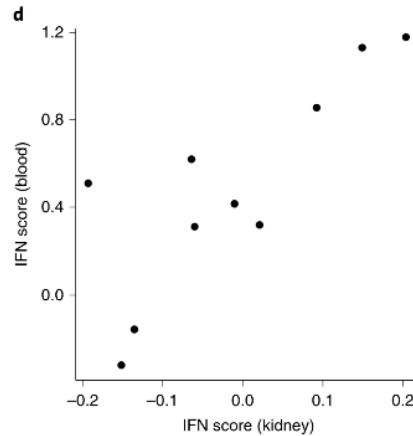
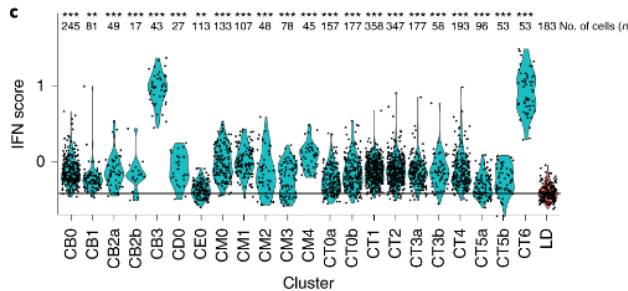
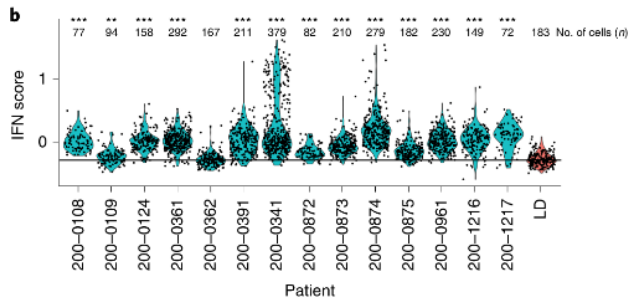
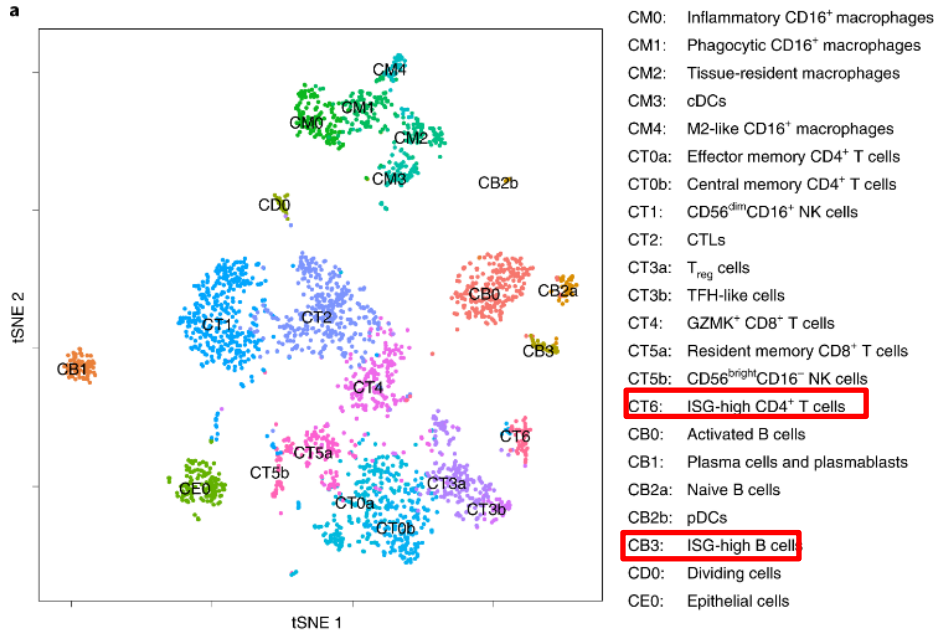


AUC



2.400 HLA-DR⁺lineage⁻ cells

Immune cell landscape in kidneys of patients with lupus nephritis

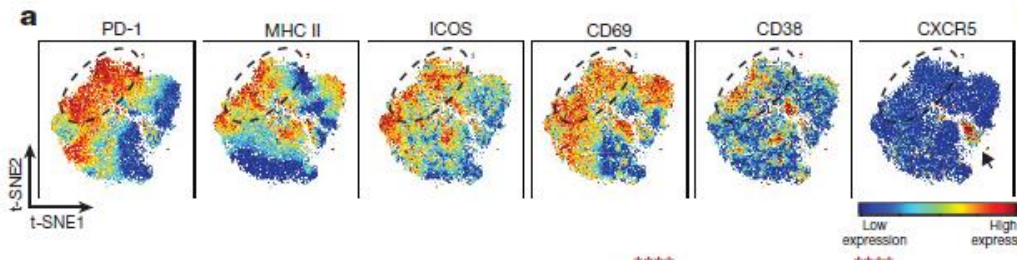


- First single-cell dissection of LN
- 24 patients, 10 ctrls
- 21 subsets of leukocytes, including clusters of myeloid cells, T-cells, NK, B-cells
- CXCR4, CX3CR1 broadly expressed
- Use of urine liquid biopsies and kidney samples



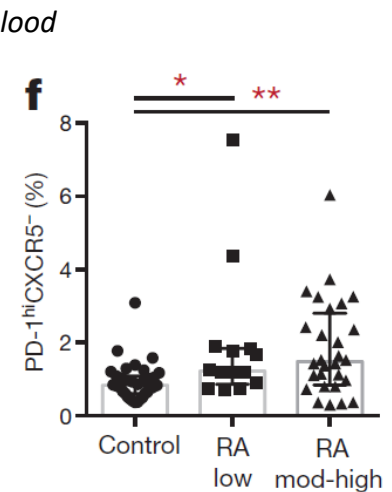
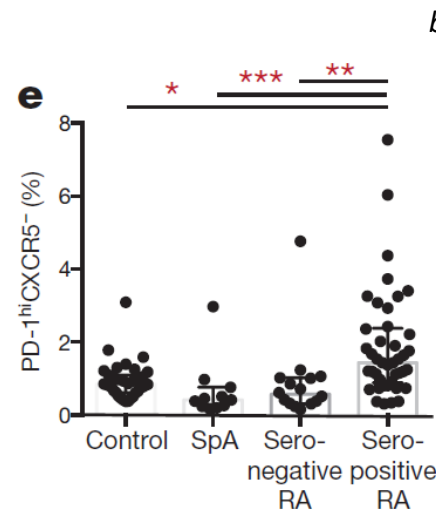
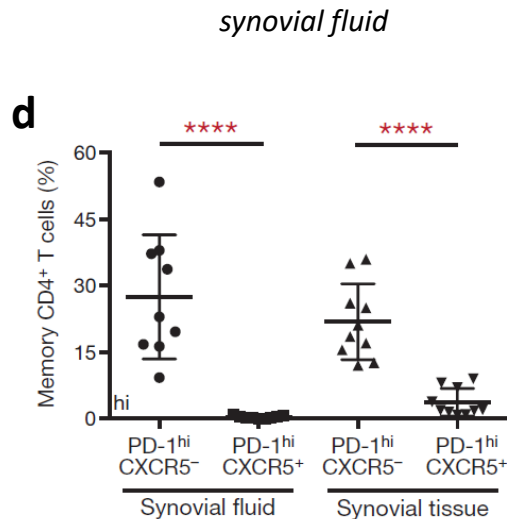
Novel pathological cell types: expanded peripheral Th subsets in RA

PD-1^{hi} CXCR5⁻ CD4⁺



Pathologically expanded peripheral T helper cell subset drives B cells in RA

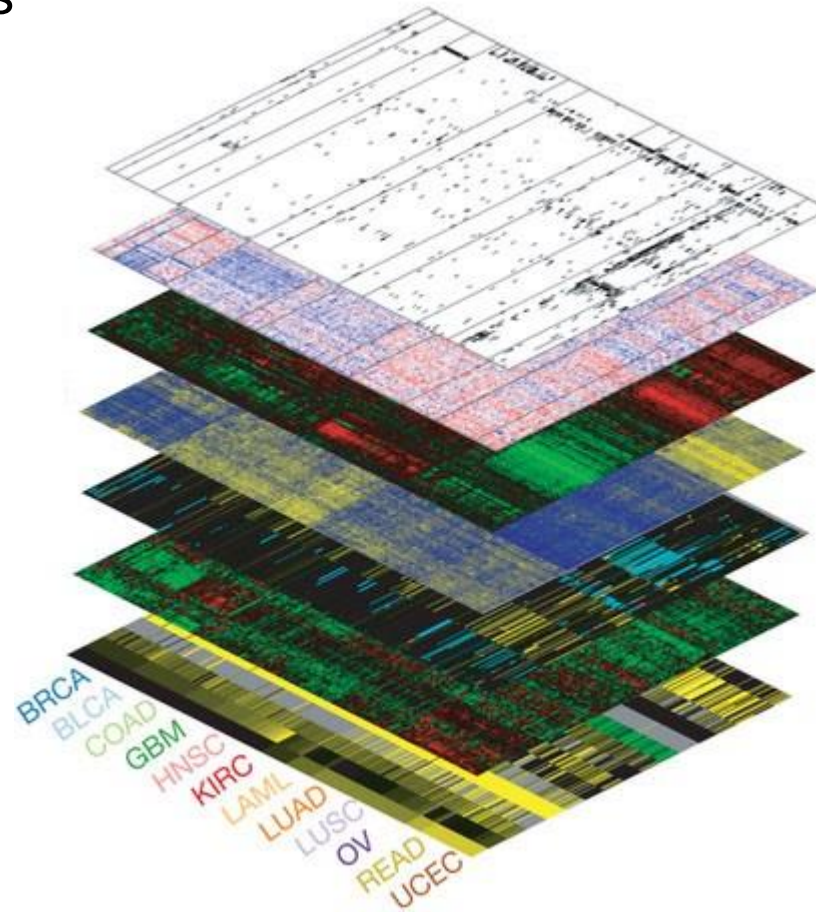
Synovial PD-1^{hi} CXCR5⁻ CD4⁺ T cells express factors associated with B-cell help (ICOS, IL21, MAF).



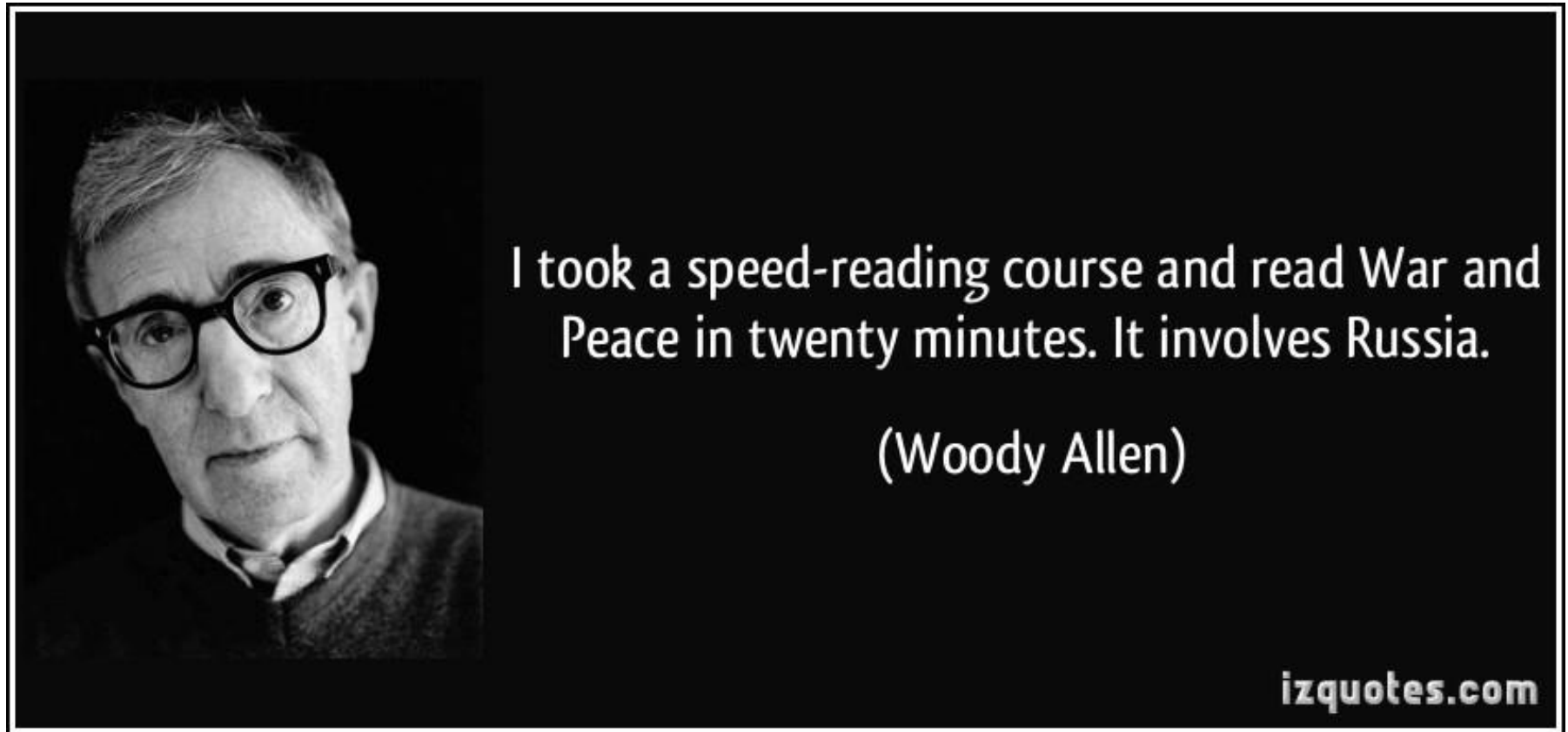
DATA AGGREGATION AND VISUALIZATION

Data Dimensionality

- Gene/protein expression
- Methylation
- Epigenetics markers
- Genetics SNPs
- Matabolomics
-



Data aggregation is any process in which information is gathered and expressed in a summary form, for purposes such as statistical analysis

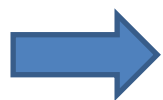
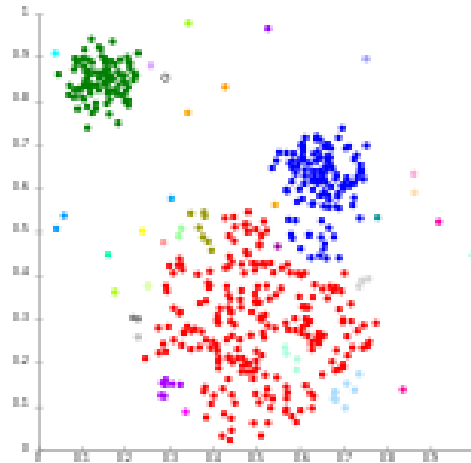


- Clustering and Geometrical Representation of Data
- Dimensions Reduction
- Pathways and Gene Sets

Clustering

Finding a partition such that:

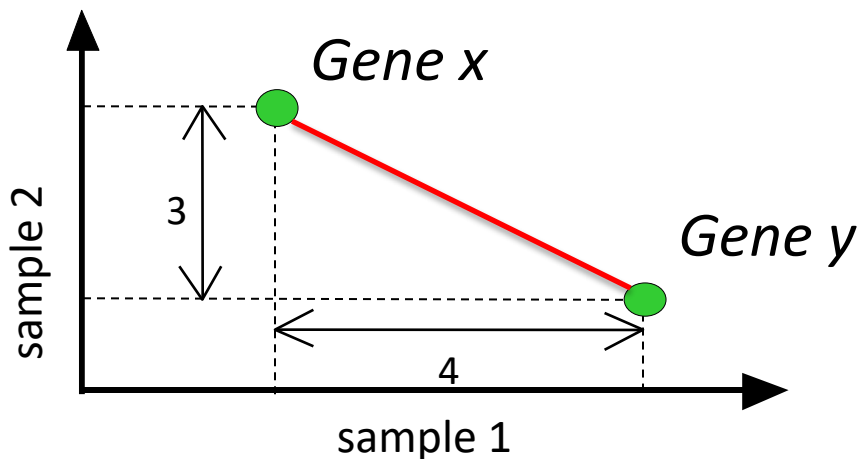
- Distance between objects within the same cluster is **minimised**
- Distance between objects from different clusters is **maximised**



Requires defining a **similarity measure**

Geometrical Distances as measures of similarity

	Sample 1	Sample 2
Gene X	2	3
Gene Y	5	1

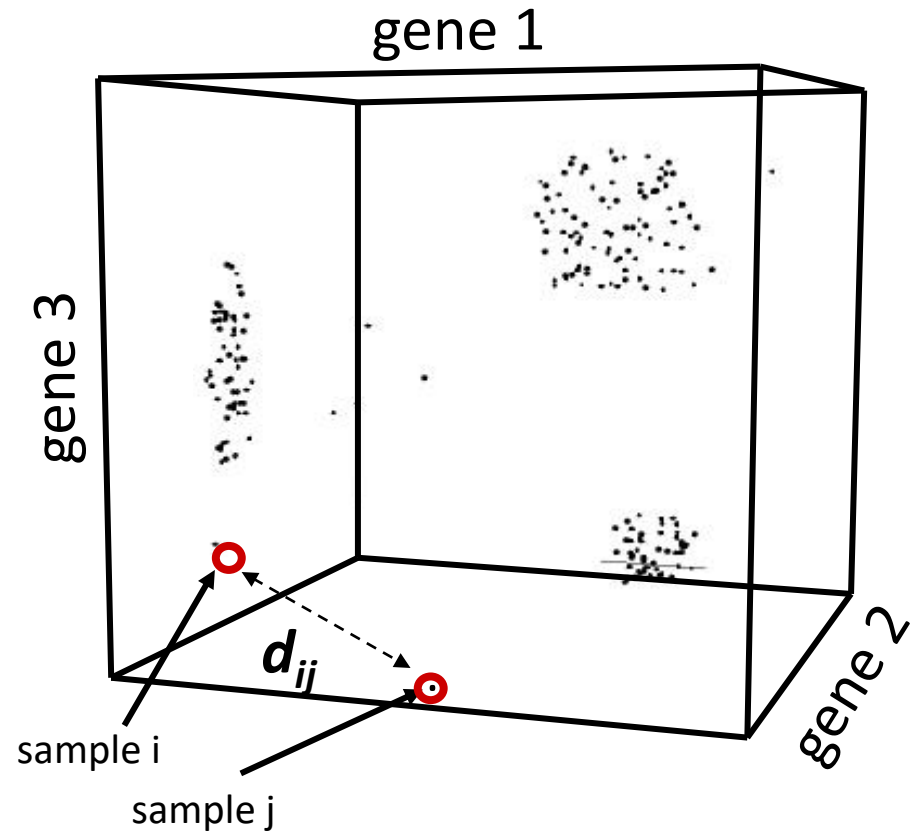


- Euclidean distance : $\sqrt{4^2 + 3^2} = 5$
- Manhattan distance : $4 + 3 = 7$
- "sup" distance : $\max\{4, 3\} = 4$

- Similarity among genes/samples is expressed as a mathematical distance
- Genes/samples close in the “expression space” have similar expression profiles

Geometrical Distances as measures of similarity

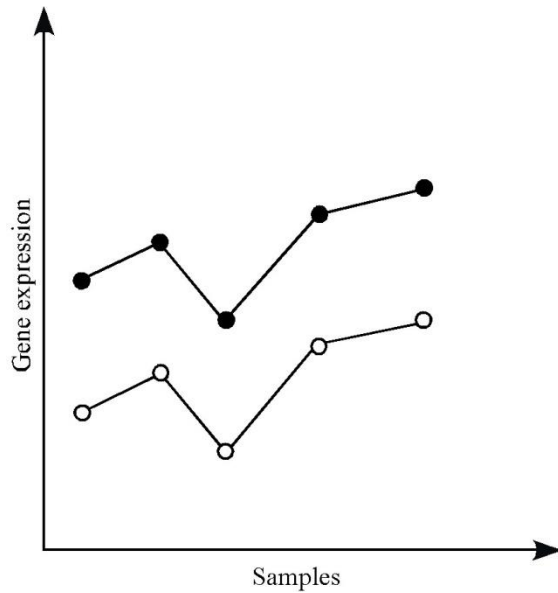
	Sample ₁	Sample ₂
Gene ₁	2	3
Gene ₂	5	1
Gene ₃	7	4



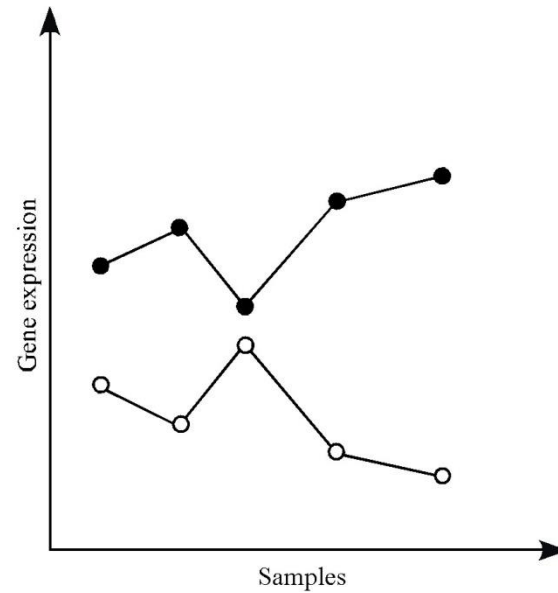
□ N genes = N dimensions

□ each sample can be represented as a point in the N-dimensional space

Similarity based on correlation



positive correlation



negative correlation

$$\sigma(x, y) = E [(x - E[x])(y - E[y])],$$

- correlation distance: $\frac{\text{cov}(a, b)}{\text{std}(a) \cdot \text{std}(b)}$

Unsupervised Methods: Hierarchical Clustering

At the beginning, each object (gene) is a cluster. In each of the subsequent steps, the two *closest* clusters are merged into one cluster until there is only one cluster left.

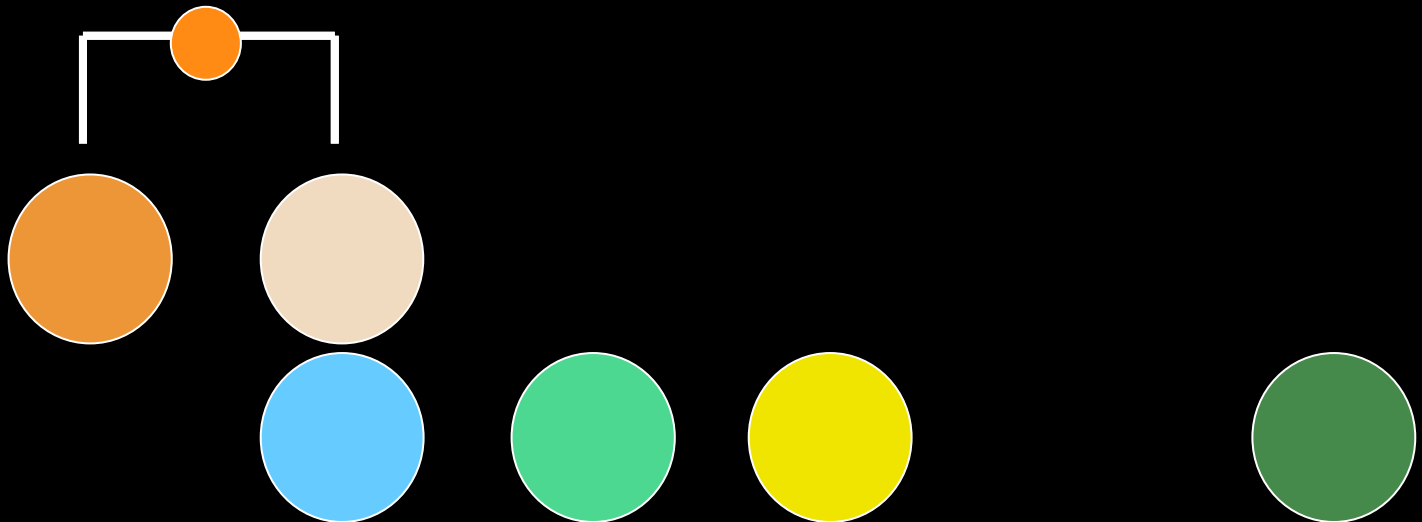
Hierarchical Clustering

Similarity distance: color



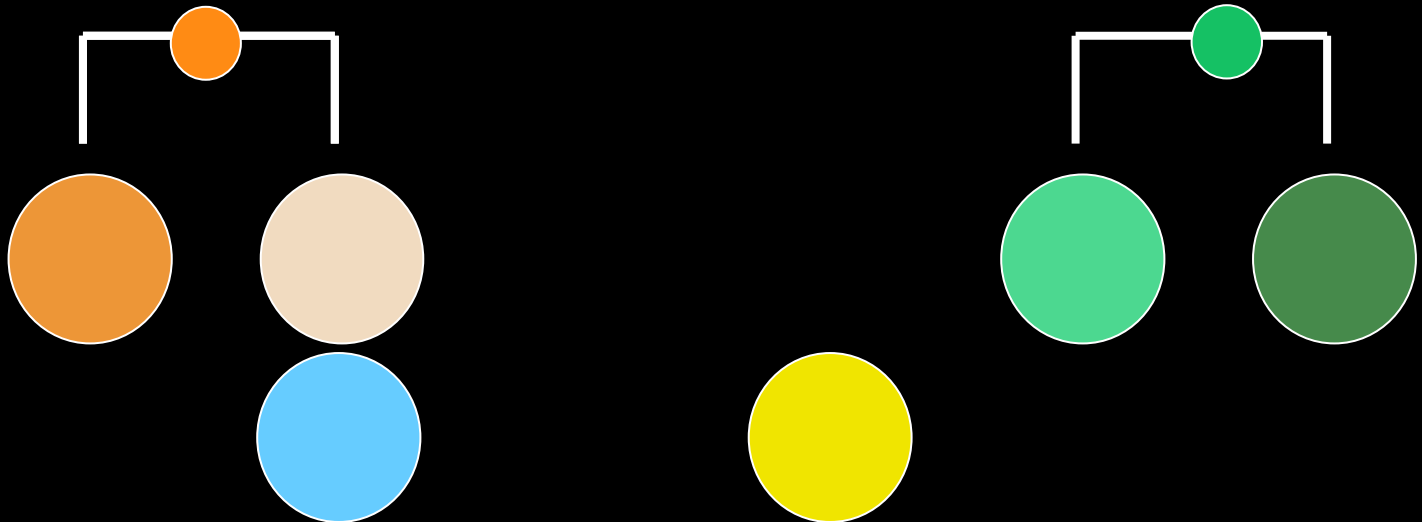
Hierarchical Clustering

Similarity distance: color



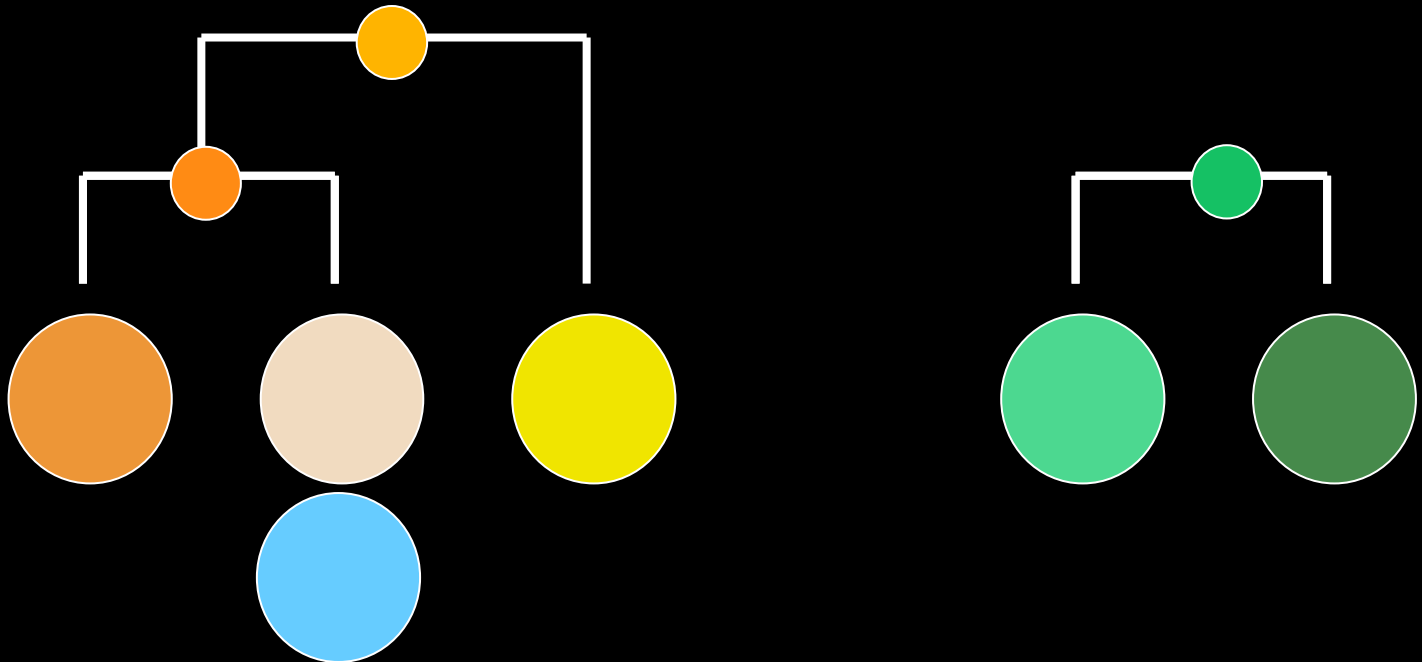
Hierarchical Clustering

Similarity distance: color



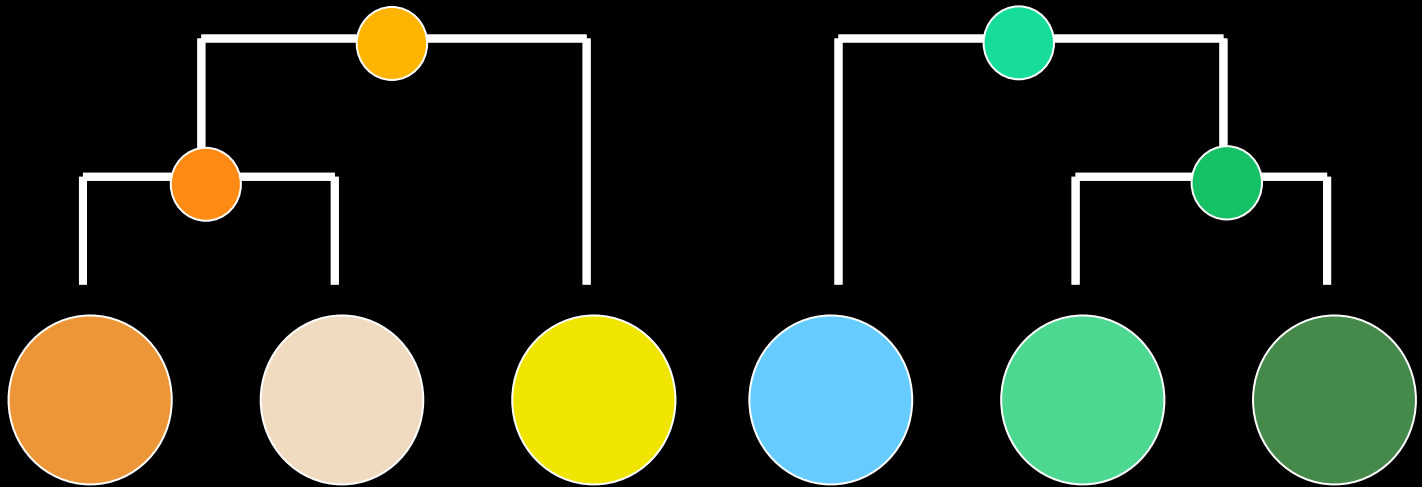
Hierarchical Clustering

Similarity distance: color



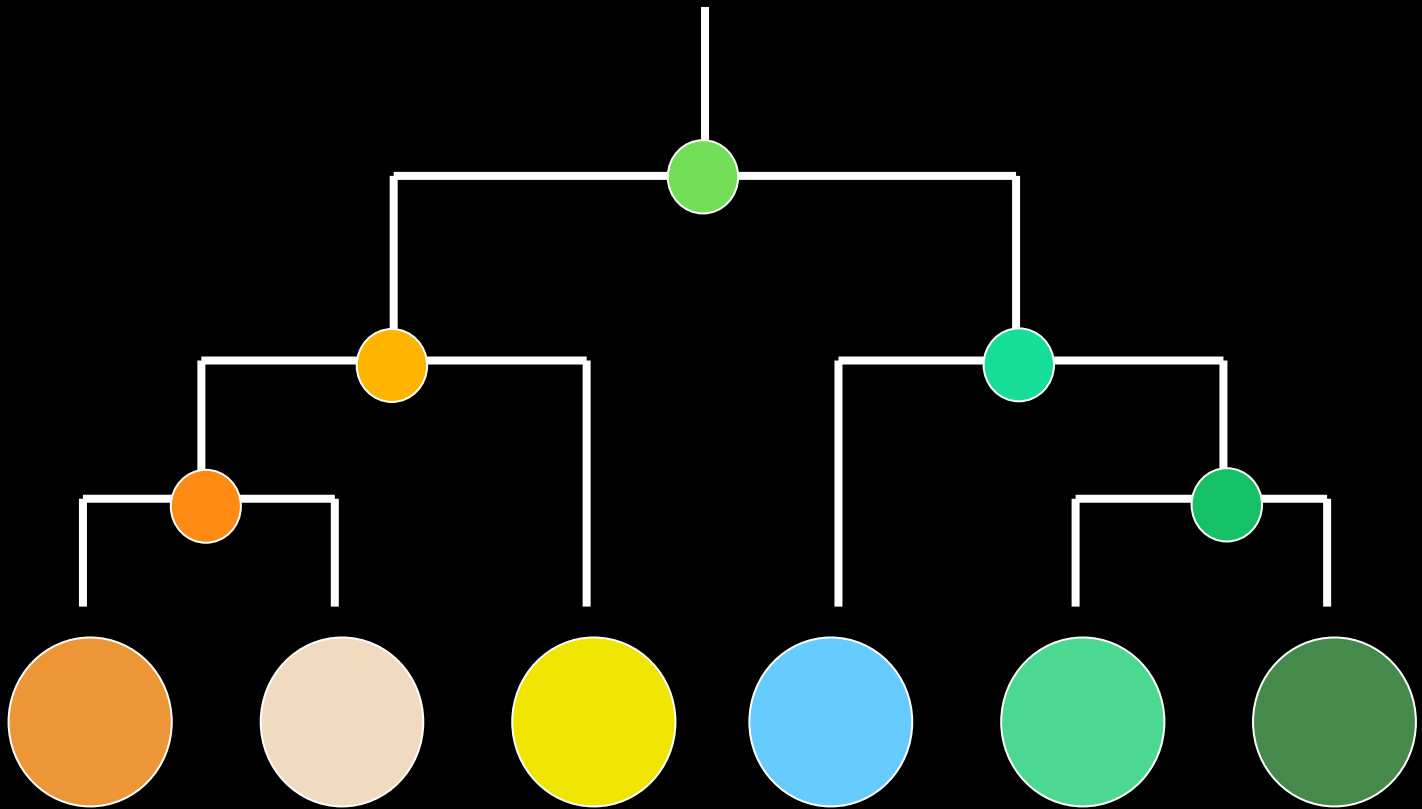
Hierarchical Clustering

Similarity distance: color



Hierarchical Clustering

Similarity distance: color



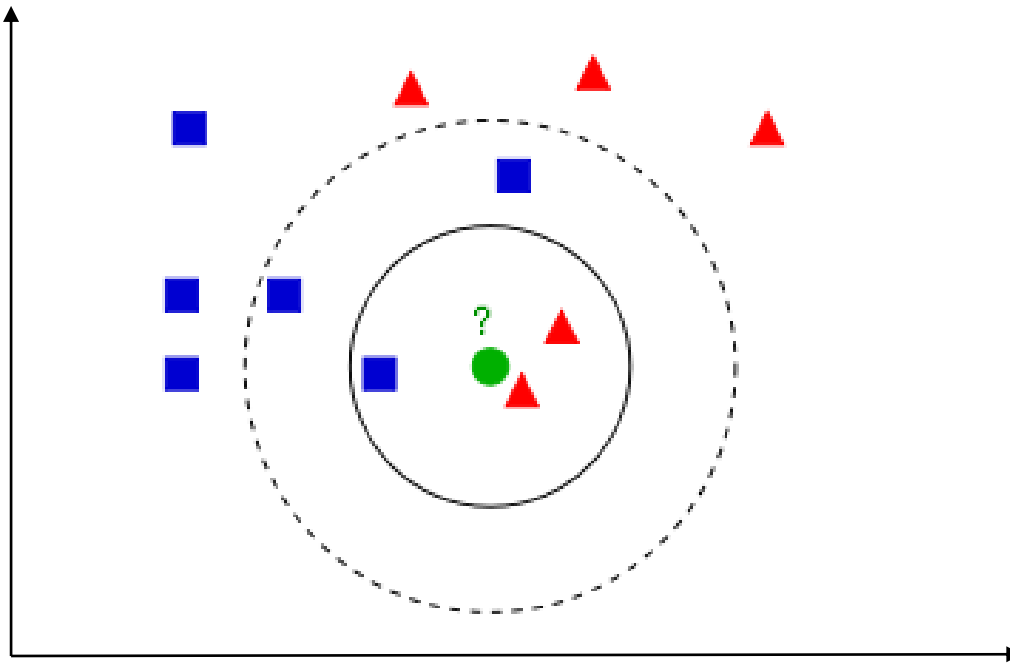
Supervised Clustering (Classification)

- use pre-existing biological information (e.g. tumor type, immune cell type, responders/non-responders etc.)
- Are used to infer which class an unknown sample belongs to
- Machine learning methods: **k-nearest neighbors**, SVM, Random forests, Bayesian networks, Deep Learning

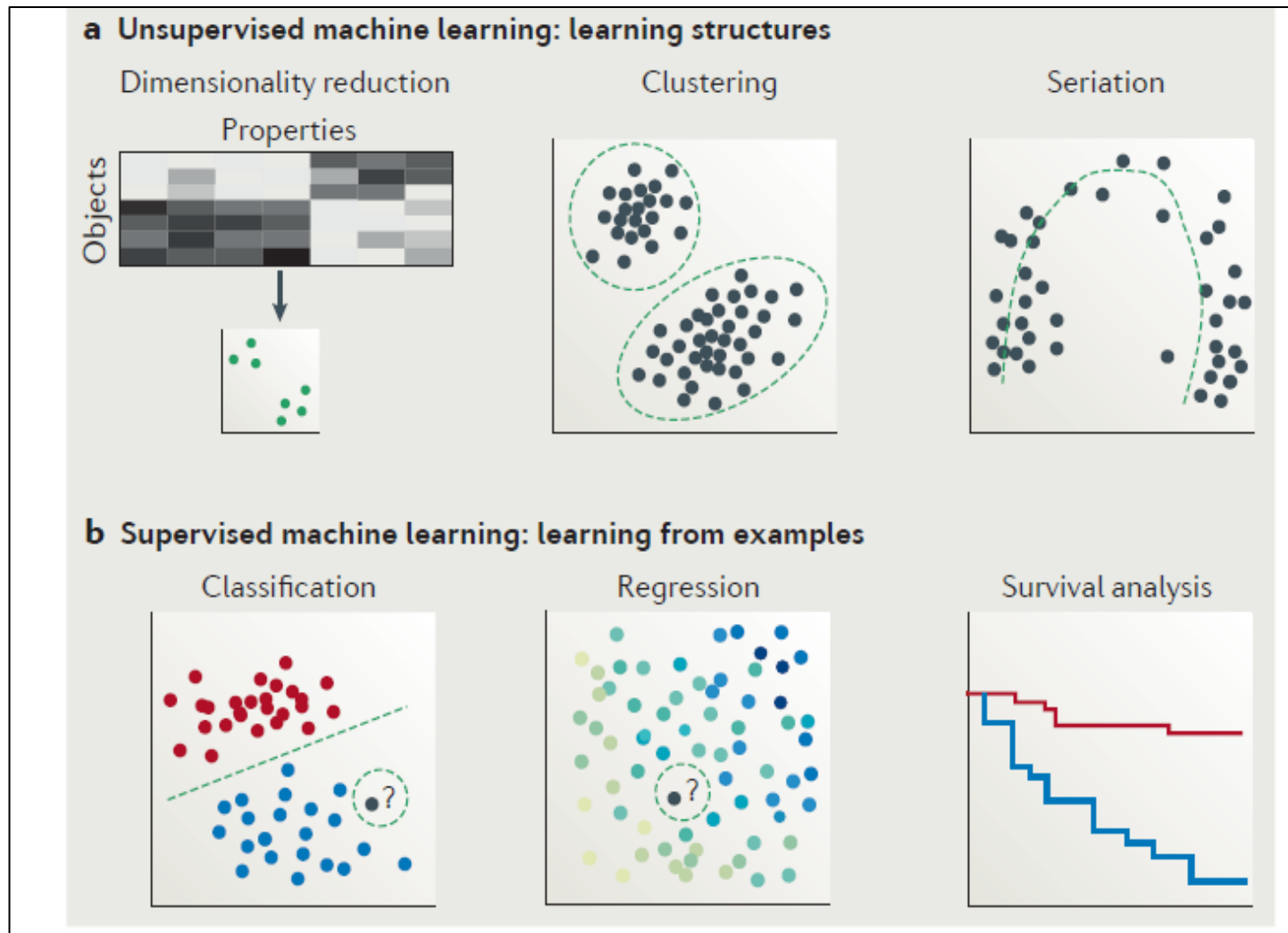
k-nearest neighbors classification (k-NN)

An object is classified by a majority vote of its neighbours, with the object being assigned to the class most common among its k nearest neighbours (k is a positive integer, typically small).

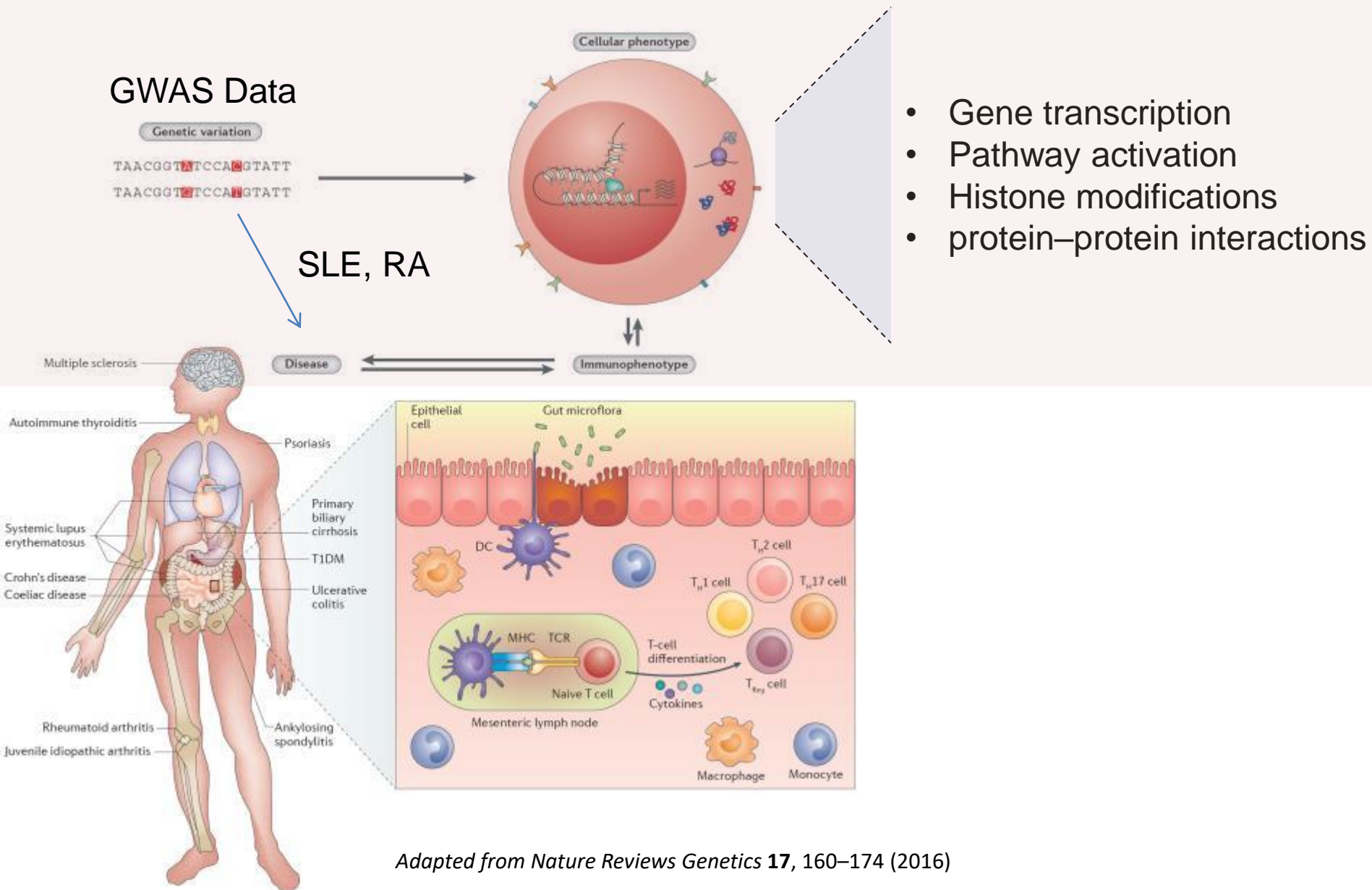
If $k = 1$, then the object is simply assigned to the class of that single nearest neighbour.



Supervised and Unsupervised Machine Learning



Exploring connections between Genetics, Immune Phenotypes and Clinical Phenotypes



Adapted from *Nature Reviews Genetics* **17**, 160–174 (2016)

Q & A

FOCIS JUNE
ANNUAL MEETING 20-23
BOSTON - 2023



Deep Dive into Case Studies:

From Systems Immunology to Novel Therapeutic Insights

Emanuele de Rinaldis

*Workshop in Systems Immunology
June 19thth, 2023*



Leveraging on integration of orthogonal data sets to identify genes of therapeutic interest in RA (R. Plenge's)

LETTER

doi:10.1038/nature12873

Genetics of rheumatoid arthritis contributes to biology and drug discovery

[Nature](#). 2014 Feb 20;506(7488):376-8

Understanding SLE biology and stratifying patients using blood bulk gene expression data (V. Pascual's)

Article

Cell

Personalized Immunomonitoring Uncovers Molecular Networks that Stratify Lupus Patients

[Cell](#). 2016 Apr 21;165(3):551-6

Genetics and Drug Discovery in RA – Study Workflow

LETTER

doi:10.1038/nature12873

Genetics of rheumatoid arthritis contributes to biology and drug discovery

Identification of SNPs associated to RA

From SNPs to causal genes through fine-mapping

Characterization of results

Data integration and genes prioritization

Assessment of the workflow using validated targets

- Novel loci associated to RA
- New hints on disease biology
- Novel candidate targets
- Repositioning of existing drug targets

Toolbox

- **Genome Wide Association Studies (GWAS) and Meta-Analysis**
 - Genome variability and SNPs
 - Logistic Regression
 - Linkage Disequilibrium
 - Imputation
 - Manhattan Plots
- **Multiple Testing**
- **Network Analysis**
- **Fine-mapping and data integration**
 - Epigenetics data
 - Transcriptional data → eQTLs
- **Statistical enrichment**




```
graph LR; A[Identification of SNPs associated to RA] --> B[From SNPs to causal genes through fine-mapping]; B --> C[Characterization of results]; C --> D[Data integration and genes prioritization]; D --> E[Assessment of the workflow using validated targets];
```

Identification of SNPs associated to RA

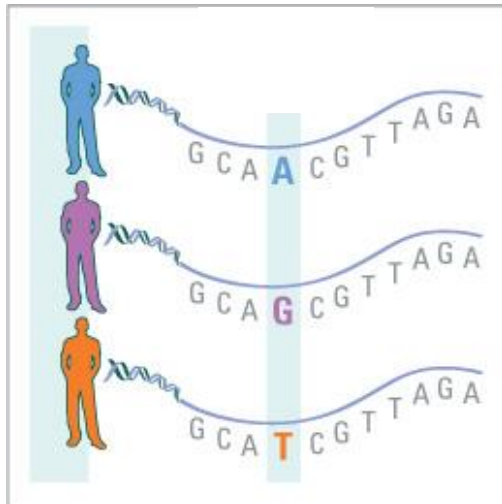
From SNPs to causal genes through fine-mapping

Characterization of results

Data integration and genes prioritization

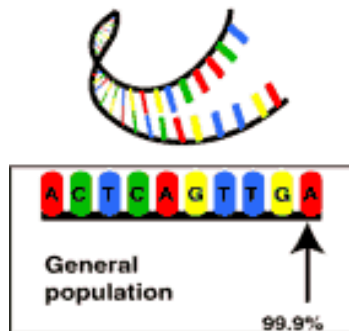
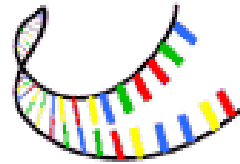
Assessment of the workflow using validated targets

Genetic Variability



Polymorphism

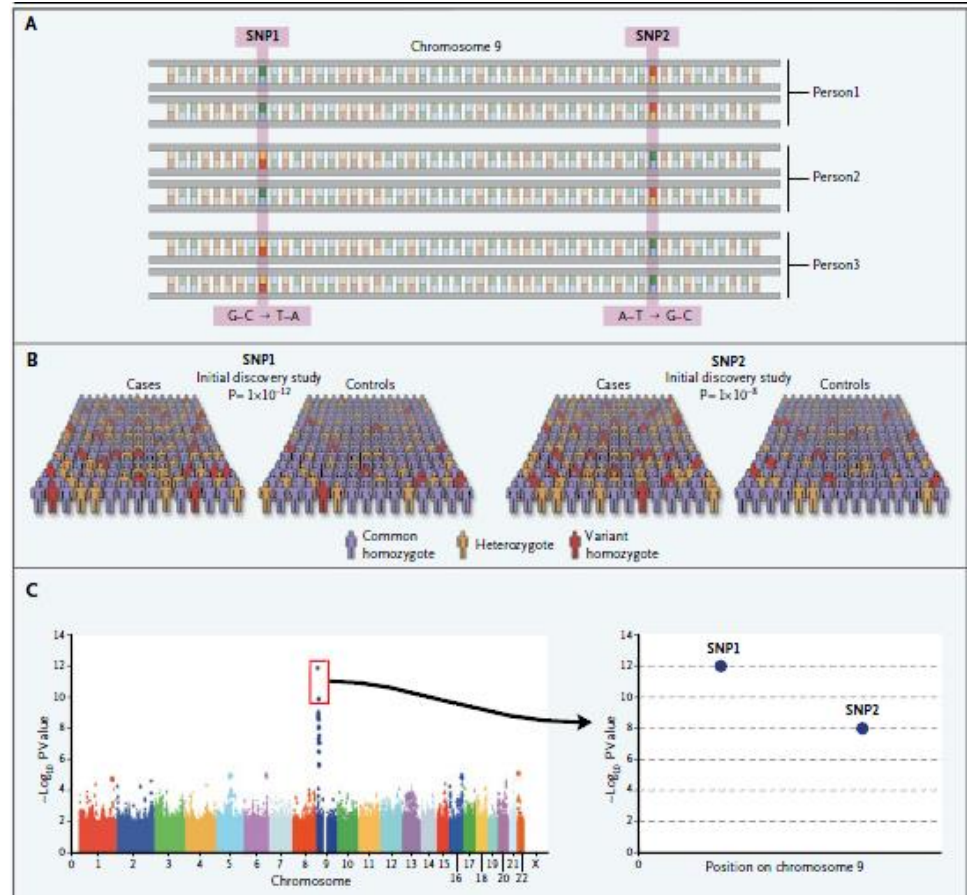
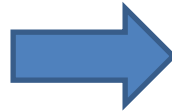
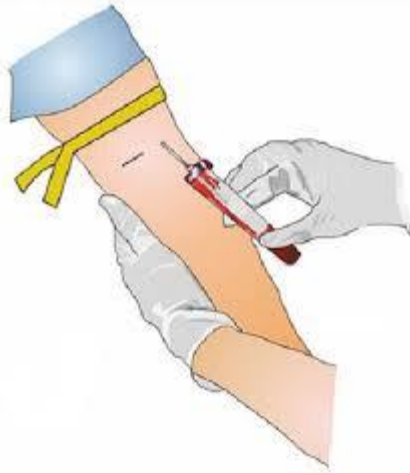
"Poly" *many* "morpho" *form*



SNP: common (>1%) variant of one **Single Nucleotide**

Mutation: typically a rare variant, associated with a disease

Genome-Wide Association Studies (GWAS)



N Engl J Med. 2010 Nov 18;363(21):2076-7.

Extraction of germline DNA (e.g. blood)

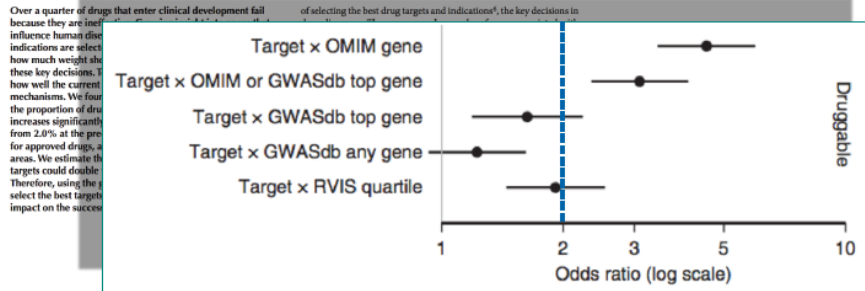
Genome-wide association study: An approach used in genetics research to look for associations between many - typically hundreds of thousands - specific genetic variations - most commonly single-nucleotide polymorphisms - and particular diseases

Why Pharma Is Investing in Genetics

The support of human genetic evidence for approved drug indications

Matthew R Nelson¹, Hannah Tipney², Jeffery L Painter¹, Judong Shen¹, Paola Nicoletti¹, Yufeng Shen^{3,4}, Aris Floratos^{3,4}, Pak Chung Sham^{3,4}, Mulin Jun Li^{6,7}, Junwen Wang^{6,7}, Lon R Cardon³, John C Whittaker² & Philippe Sansseau²

~2-fold increase in success for genetic targets



RESEARCH ARTICLE

Are drug targets with genetic support twice as likely to be approved? Revised estimates of the impact of genetic support for drug mechanisms on the probability of drug approval

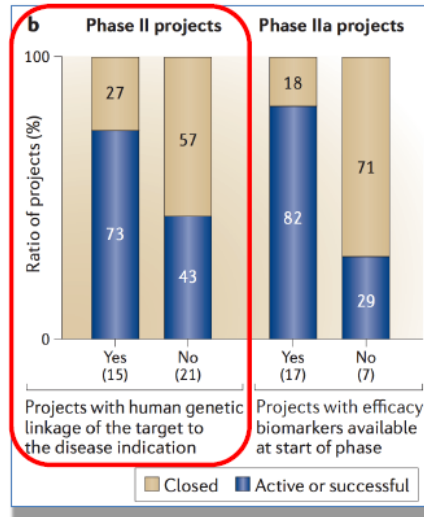
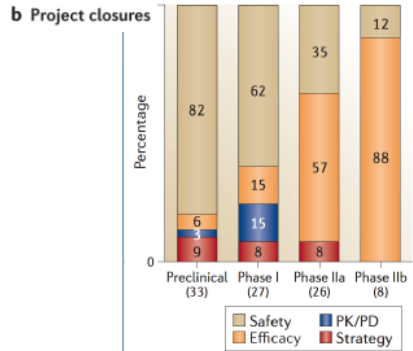
Emily A. King¹*, J. Wade Davis, Jacob F. Degner

Department of Computational Genomics, AbbVie, North Chicago, Illinois, United States of America

* emily.king@abbvie.com

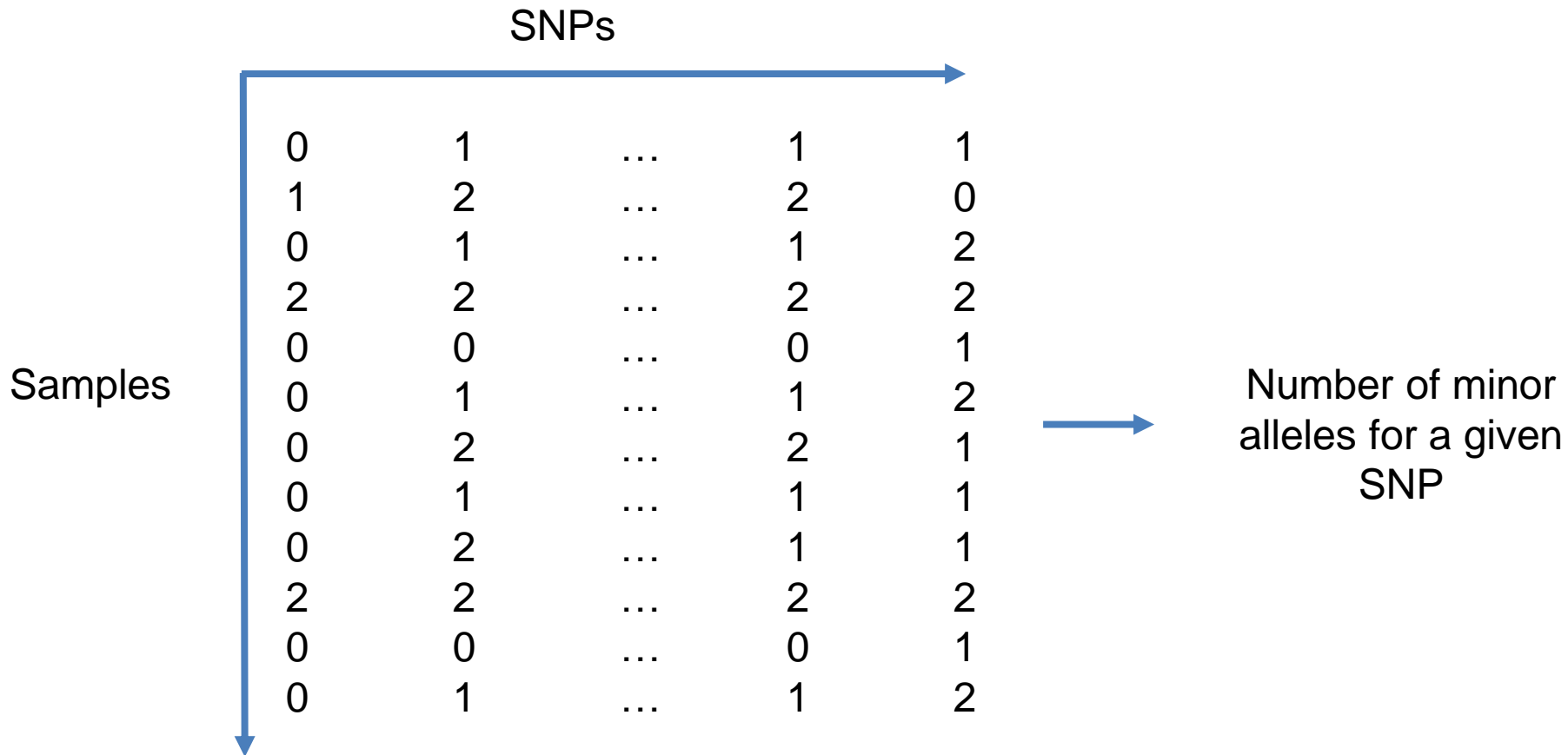
Lessons learned from the fate of AstraZeneca's drug pipeline: a five-dimensional framework

David Cook, Deary Brown, Robert Alexander, Ruth March, Paul Morgan, Gemma Satterthwaite and Menelas N. Pangalos



- ❑ When causality is clear (Mendelian traits or coding variants) fold increase for success greater than 2 folds
- ❑ Limited contribution to GWAS genetic evidences not in OMIM → undetermined function

Genetic Dataset



Single test for association



Do people carrying a certain genotype have an increased probability of having the disease?

SNP	S_IBD1	S_IBD2	S_IBD3	S_Cont1	S_Cont2	S_Cont3
rsxxxxx	0	0	1	0	1	0
rsxxxx0	0	0	0	0	0	0
rs...	1	0	0	0	2	1
rs...	0	0	0	1	0	1
rs...	0	0	1	1	1	1
PC1	..					
PC2			..			
PC

$$\text{Log}\left(\frac{p}{1-p}\right) = \beta_0 + \beta_1 \text{snp} + \beta_2 \text{pc1} + \beta_3 \text{pc2} + \beta_4 \text{pc3} + \dots$$

The tool used was “plink”, What we get back is:
the value of B1, and
a p-value for B1 being different than zero.

Other variable of interest like ethnicity, age, sex can be added to the model

Logistic regression



Test all SNPs for
association independently



Correct for multiple
testing

Instead of using 0.05 as a threshold for significance divide it by the total number of independent tests (5×10^{-8} for genome-wide studies)

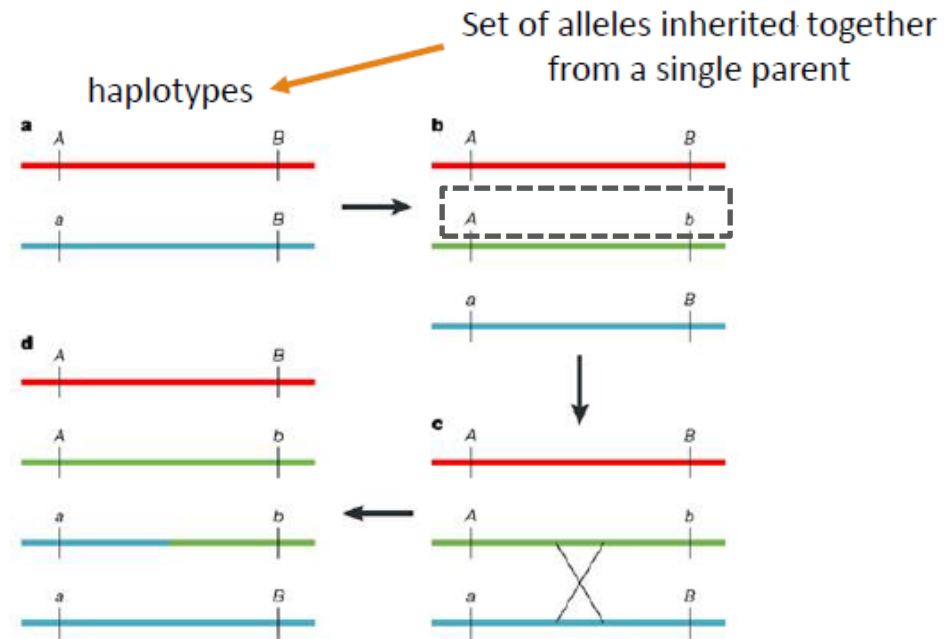
Linkage Disequilibrium



Non-random association of alleles at different loci
(genetic positions)



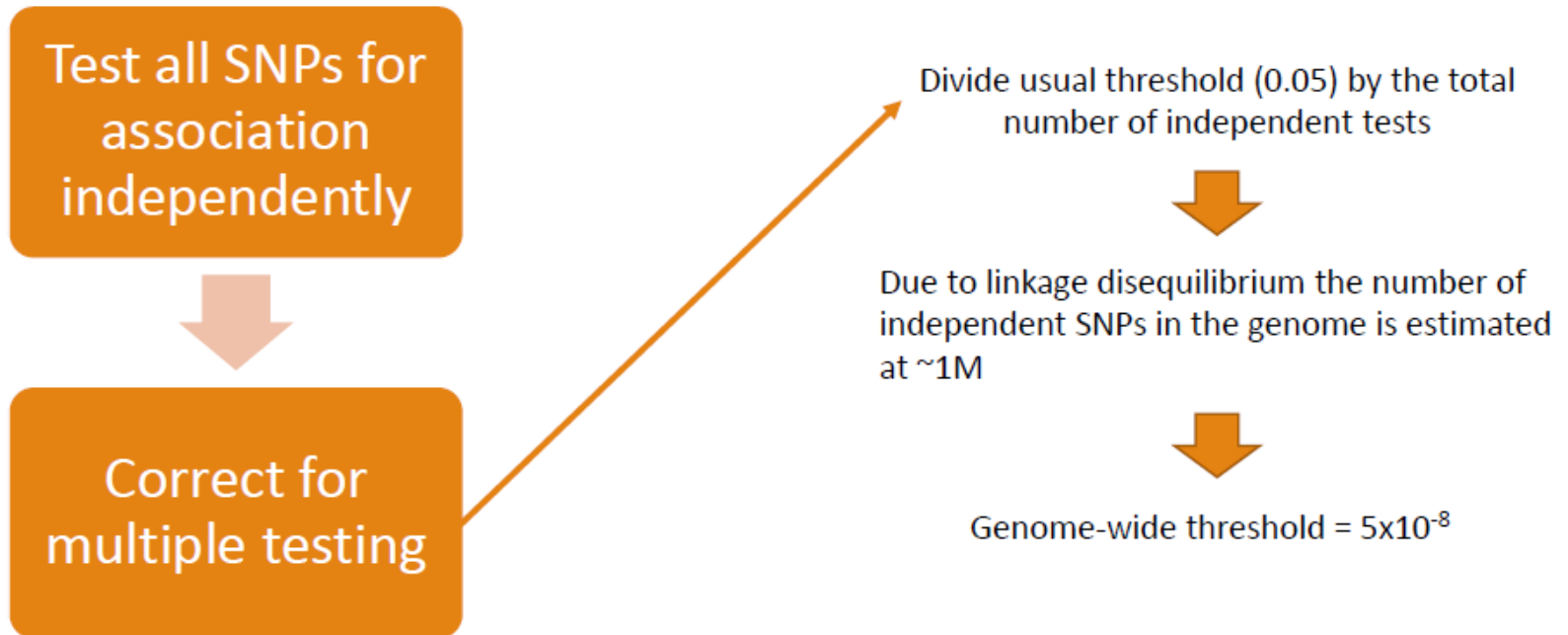
Genotypes are not independent but correlated



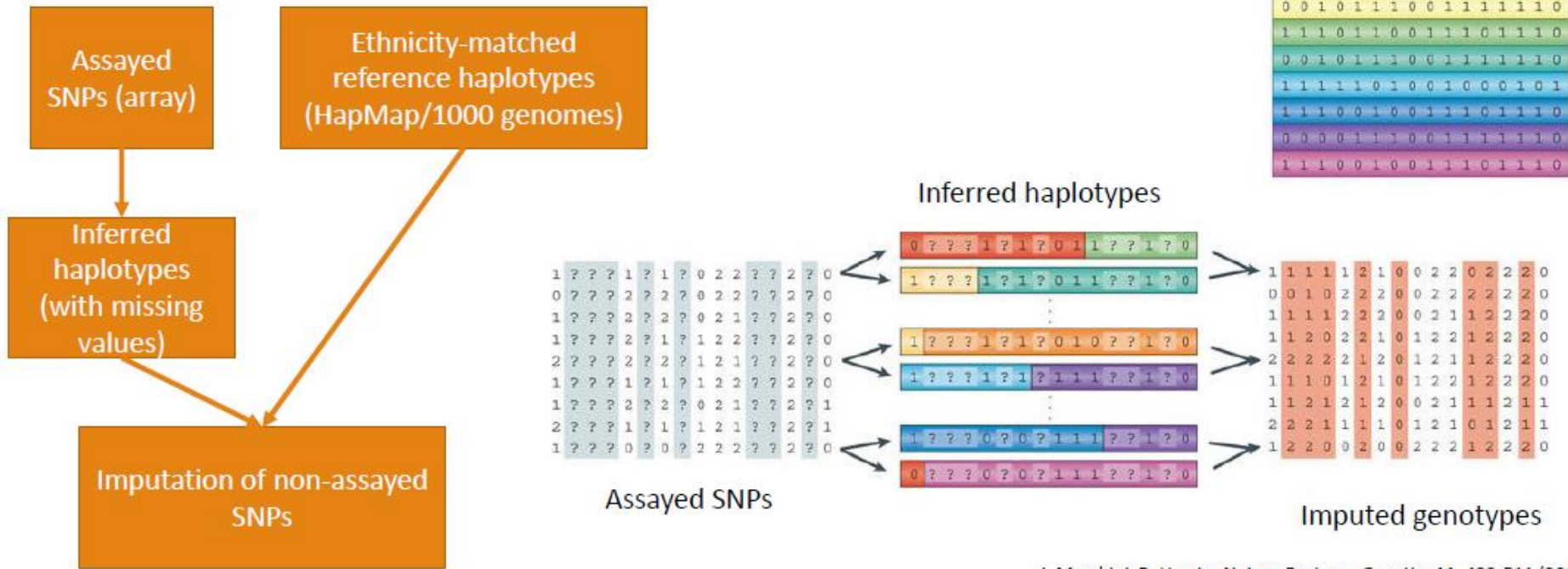
Nature Reviews | Genetics

K. G. Ardlie, L. Kruglyak, M. Seielstad, *Nature reviews Genetics* 3, 299-309 (2002)

Multiple tests in the same cohort

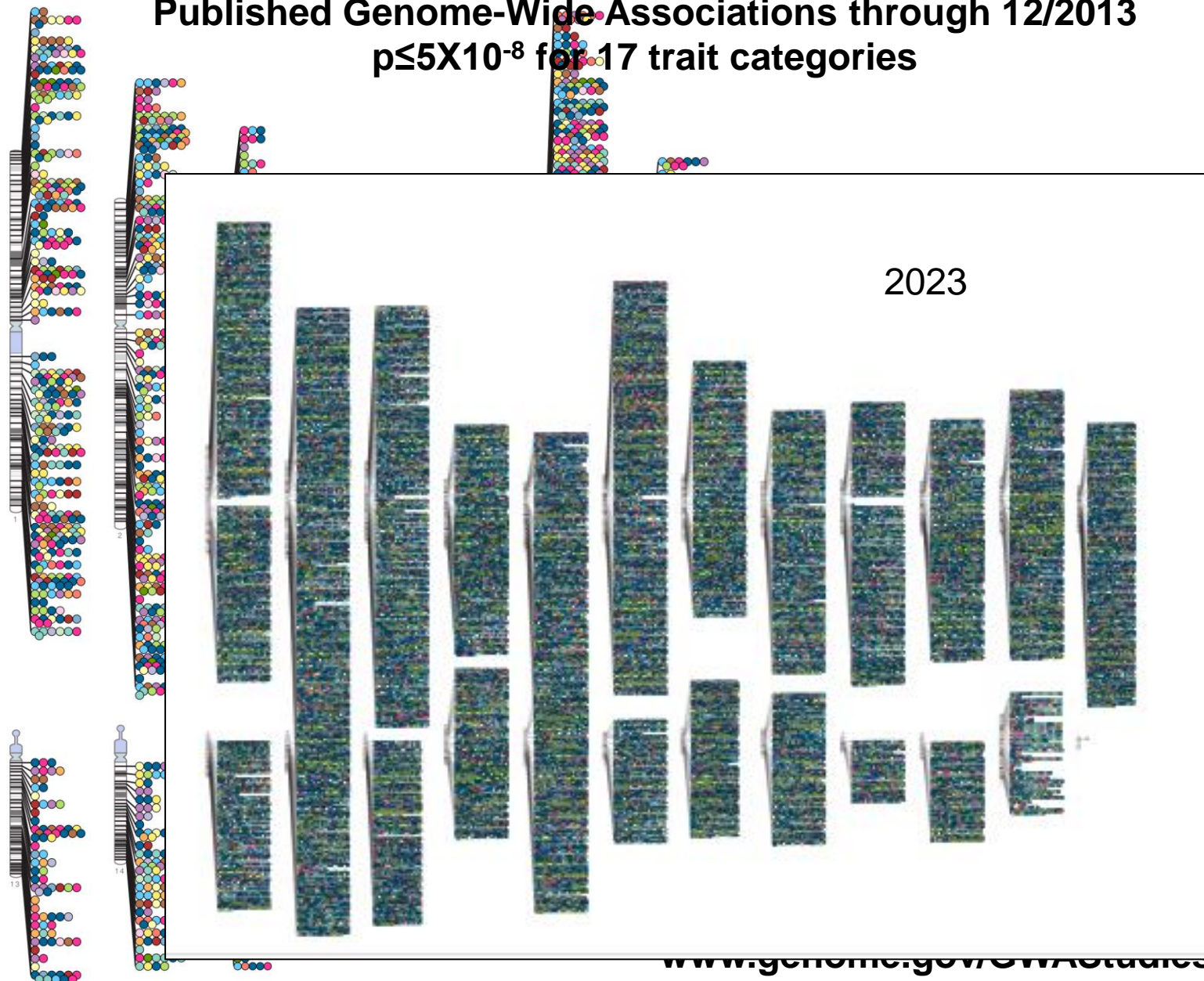


Imputation



J. Marchini, B. Howie, *Nature Reviews. Genetics* 11, 499-511 (2010)

Published Genome-Wide Associations through 12/2013 $p \leq 5 \times 10^{-8}$ for 17 trait categories



- Digestive system disease
- Cardiovascular disease
- Metabolic disease
- Immune system disease
- Nervous system disease
- Liver enzyme measurement
- Lipid or lipoprotein measurement
- Inflammatory marker measurement
- Hematological measurement
- Body measurement
- Cardiovascular measurement
- Other measurement
- Response to drug
- Biological process
- Cancer
- Other disease
- Other trait

www.genome.gov/GWAStudies
www.ebi.ac.uk/fgpt/gwas/

Study Design

a

Stage 1 : Trans-ethnic GWAS meta-analysis

19,234 RA cases and 61,565 controls
(EUR : 14,361 RA cases and 43,923 controls)
(ASN : 4,873 RA cases and 17,642 controls)

57 loci (**17 novel**) $p_{val} < 10^{-8}$



146 loci with $P < 5.0 \times 10^{-6}$ in
trans-ethnic/EUR/ASN study

Stage 2 : *In silico* replication study

3,708 RA cases and 5,535 controls
(EUR : 2,780 RA cases and 4,700 controls)
(ASN : 928 RA cases and 835 controls)



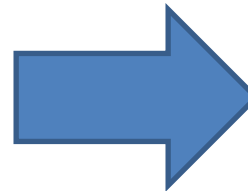
20 loci with the highest statistical power
for EUR and ASN separately (in total 32 SNPs)

Stage 3 : *De novo* replication study

6,938 RA cases and 6,658 controls
(EUR : 995 RA cases and 1,101 controls)
(ASN : 5,943 RA cases and 5,557 controls)



Combining 1-3: **42 novel loci** with $P < 5 \times 10^{-8}$



**100 Total RA risk loci (58
known + 42 novel),
including 377 genes**

Identification of SNPs associated to RA

From SNPs to causal genes through fine-mapping

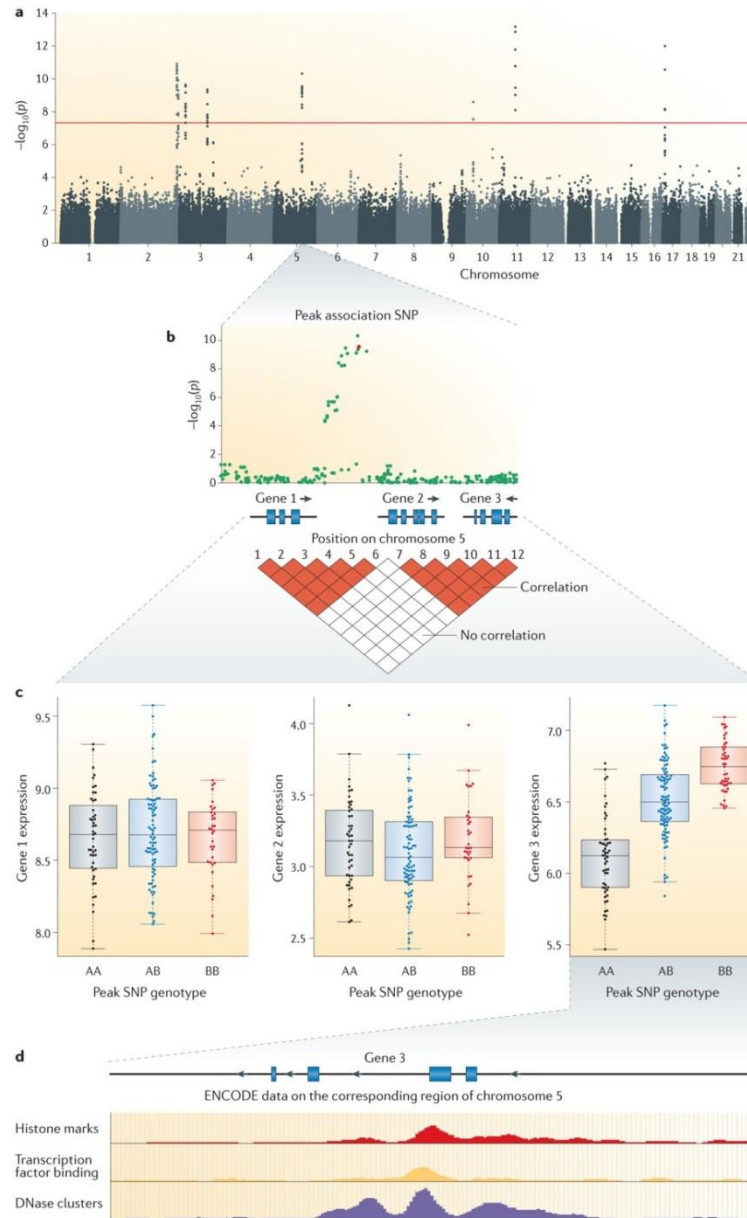
Characterization of results

Data integration and in-silico genes prioritization

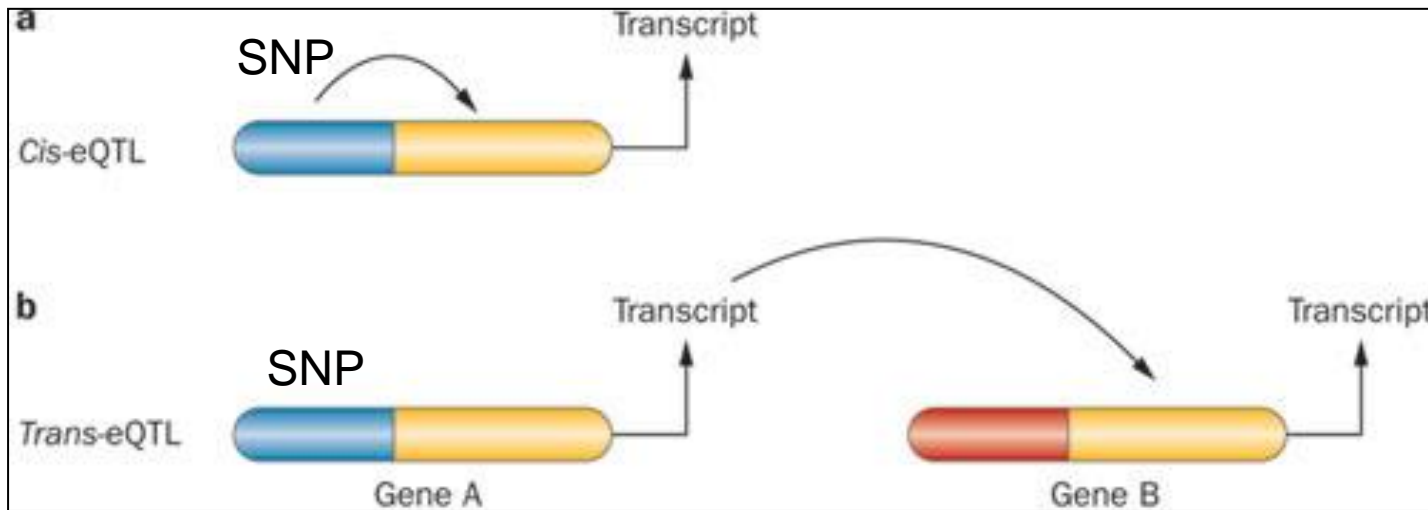
Assessment of the workflow using validated targets

Fine-Mapping: overlaying different information to identify causal genes

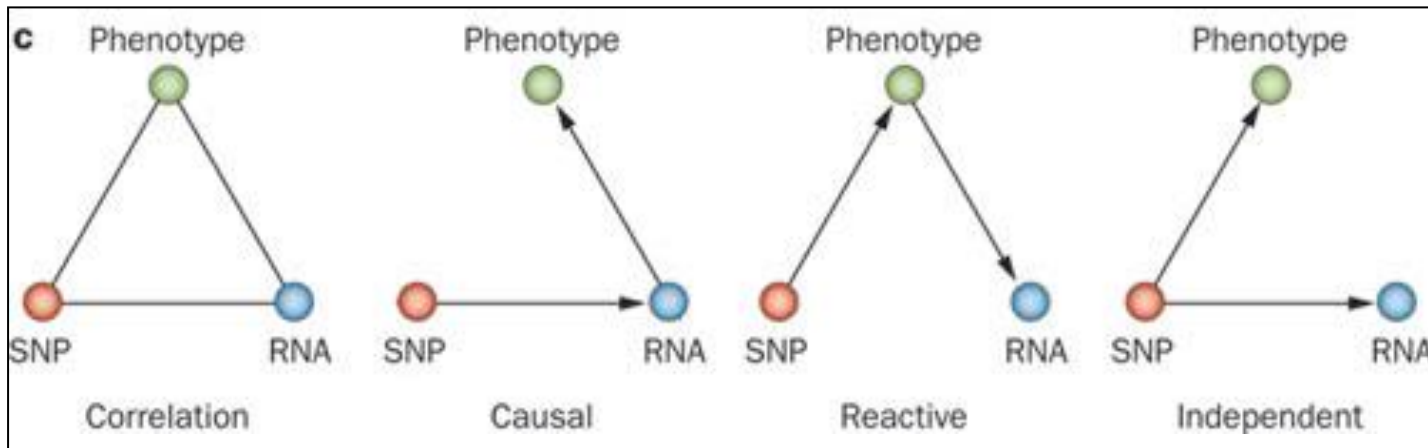
M. Civelek & A. J. Lusis – *Nat. Rev. Genetics*
(2013)



Combining DNA and RNA information – eQTLs and Causal Networks

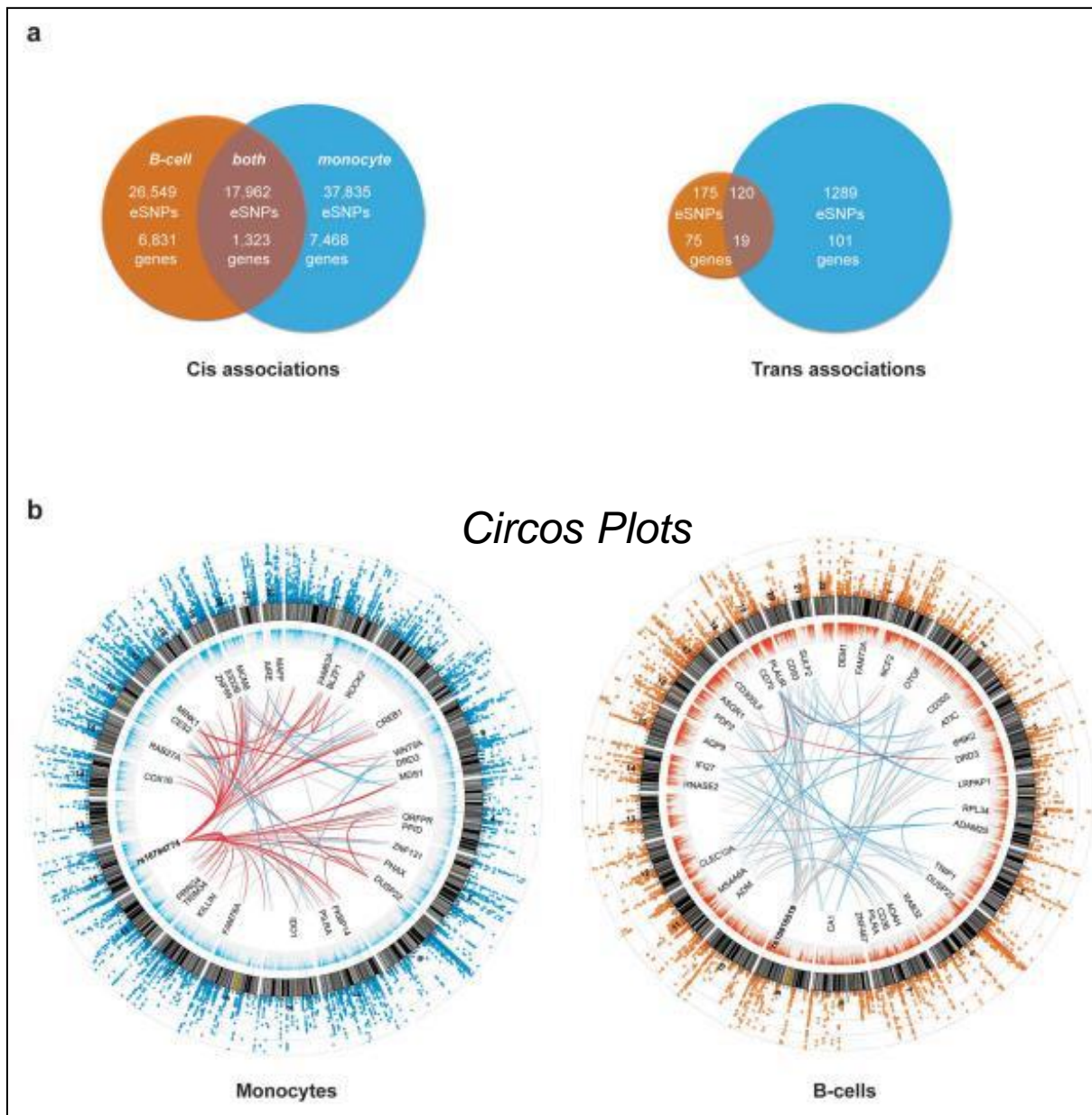


- DNA/RNA combined analysis can highlight eSNPs
- eSNPs exert an effect on genes' transcription



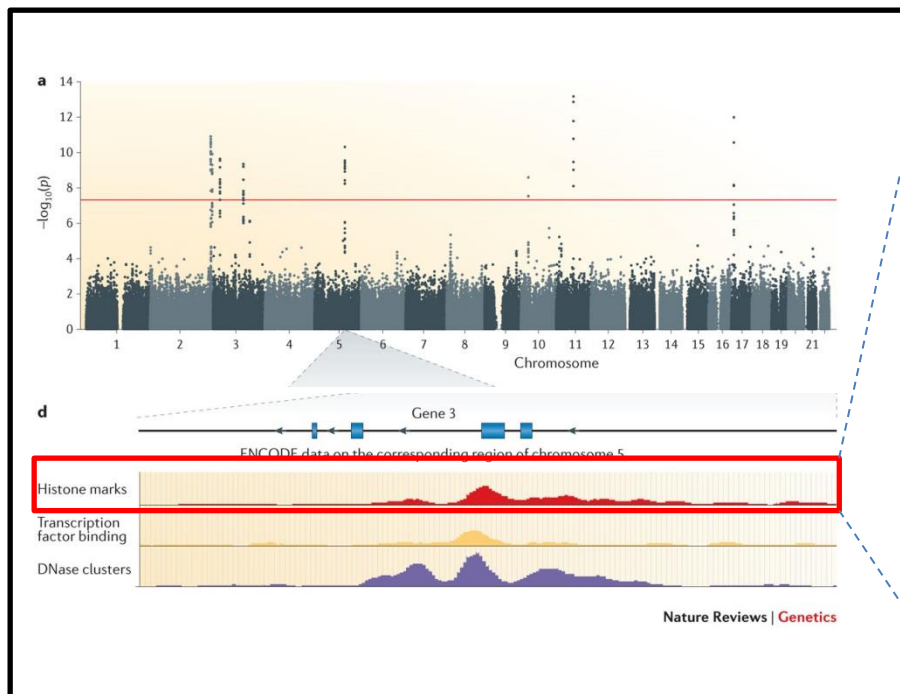
Using the SNP as a 'causal anchor', causal relationships between the three can be modelled: Causal Networks

Cell type-specific eQTLs in B-cell and monocytes



Assessment of enrichment of 100 non-MHC RNA risk loci in epigenetic chromatin marks

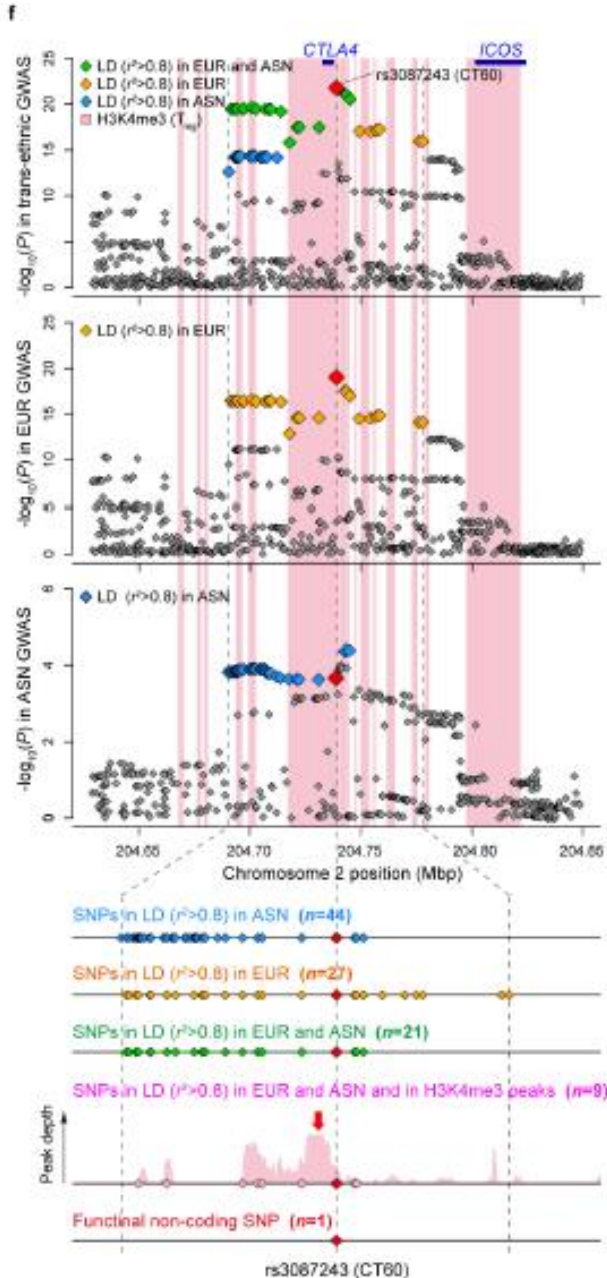
100 RA risk loci



d

Cell types	<i>P</i> for H3K4me3 enrichment
T _{reg} primary cells	$\leq 1.0 \times 10^{-5}$
CD4 ⁺ memory primary cells	3.0×10^{-9}
CD4 ⁺ naive primary cells	0.0041
CD8 ⁺ memory primary cells	0.0065
Smooth muscle, rectal Mucosa, colon	0.034
CD8 ⁺ naive primary cells	0.12
Mucosa, stomach	0.13
CD34 ⁺ primary cells	0.18
CD34 ⁺ cultured cells	0.19
Mobilized CD34 ⁺ primary cells	0.19
CD19 ⁺ primary cells	0.24
CD3 ⁺ primary cells	0.30
Mucosa, duodenum	0.40
Muscle satellite cultured cells	0.46
Cingulate gyrus (brain)	0.53
Skeletal muscle	0.77
Mucosa, rectal	0.77
Smooth muscle, colon	0.79
Mesenchymal stem cells (adipose)	0.81
Adipose nuclei	0.84
Smooth muscle, duodenum	0.85
Mid frontal lobe (brain)	0.86
Hippocampus middle (brain)	0.91
Mesenchymal stem cells (bone marrow)	0.91
Pancreatic islets	0.93
Inferior temporal lobe (brain)	0.93
Substantia nigra (brain)	0.93
Adult kidney	0.94
Adult liver	0.95
Mesenchymal stem cells (adipocyte)	0.98
Mesenchymal stem cells (chondrocytes)	0.99
Anterior caudate (brain)	0.99
Smooth muscle, stomach	0.99

Example: Fine Mapping of CTLA4



Regional (trans-ethnic, European, Asian) SNP associations of the CTLA4 locus in stage 1 GWAS meta-analysis

1. functional non-coding variant of CT60 (rs3087243) showed the most significant association with RA.
2. Trans-ethnic fine mapping of candidate causal variants decreased the number of candidate variants from 44 (LD in Asians) and 27 (LD in Europeans) to 21 (LD in both populations).
3. **Selected the 9 candidate variants included in Treg H3K4me3 peaks, including CT60 (close to H3K4me3 summit)**

Identification of SNPs associated to RA

From SNPs to causal genes through fine-mapping

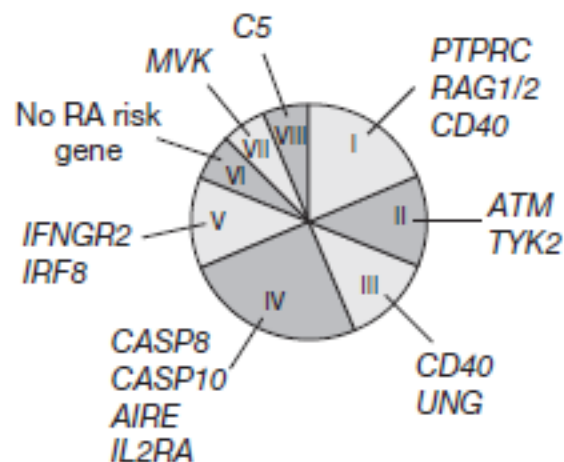
Characterization of results

Data integration and genes prioritization

Assessment of the workflow using validated targets

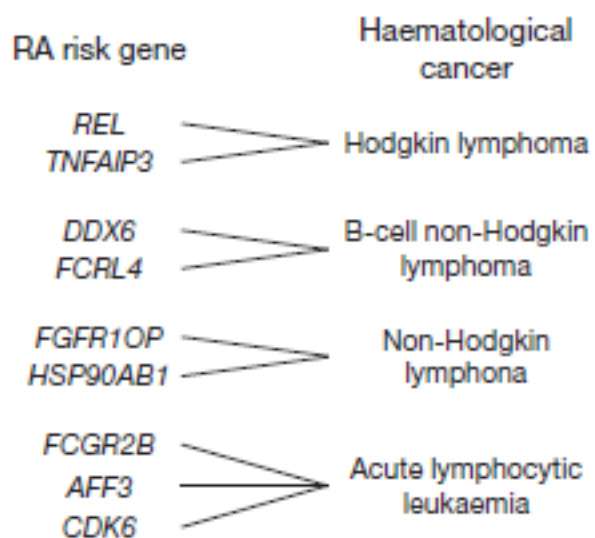
Characterization of Results (100 loci, 377 genes)

a PID categories and RA risk genes

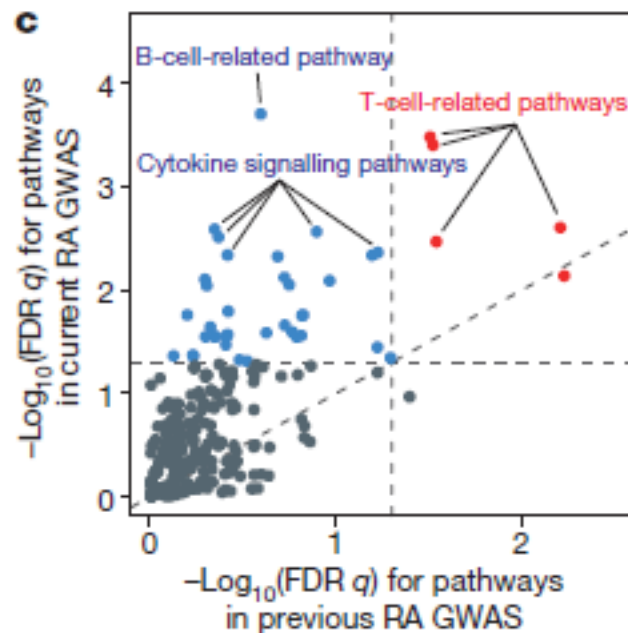


- I : Combined immunodeficiencies
- II : Well-defined syndromes
- III : Primary antibody deficiencies
- IV : Immune dysregulation
- V : Phagocyte defects
- VI : Innate immunity
- VII : Autoinflammatory
- VIII : Complement deficiencies

b



c



Identification of SNPs associated to RA

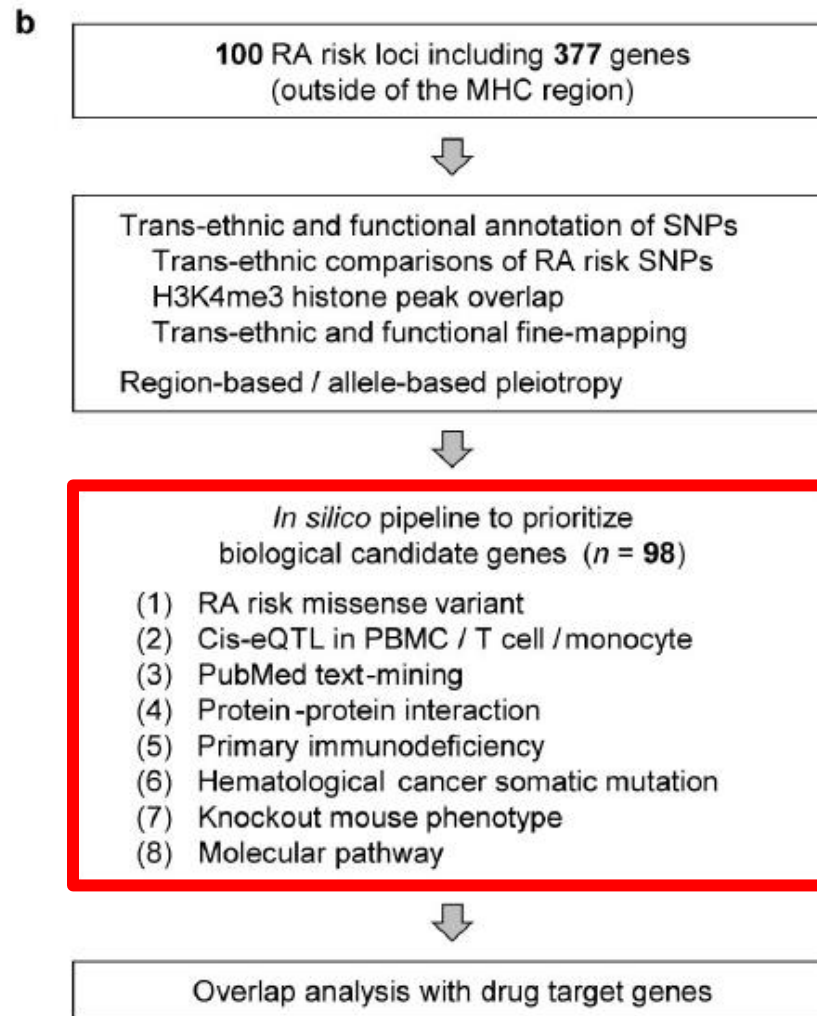
From SNPs to causal genes through fine-mapping

Characterization of results

Data integration and genes prioritization

Assessment of the workflow using validated targets

In-silico genes prioritization



[*Nature*. 2014 Feb 20;506\(7488\):376-81](#)

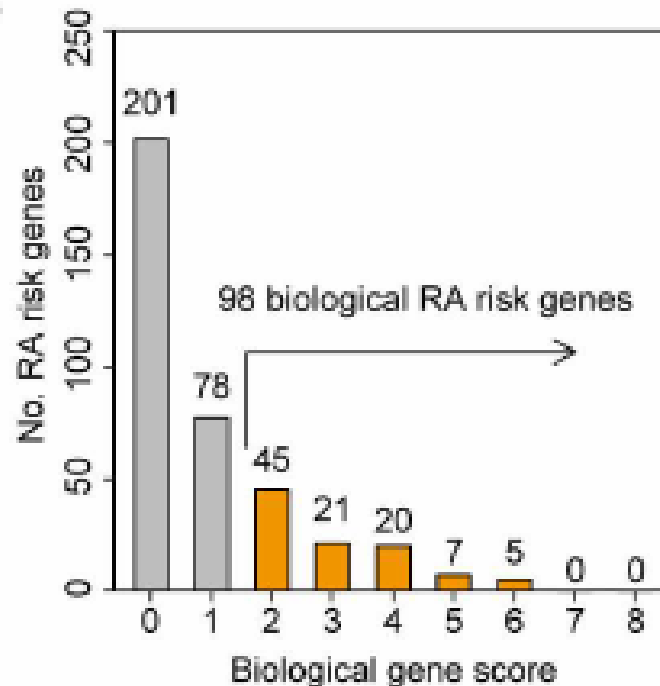
Prioritization of biological candidate genes from RA risk loci

a

Biological RA risk gene prioritization criteria

- (1) RA risk missense variant ($n = 19$)
- (2) Cis-eQTL ($n = 51$)
- (3) PubMed text-mining ($n = 90$)
- (4) Protein-protein interaction ($n = 63$)
- (5) Primary immunodeficiency ($n = 15$)
- (6) Hematological cancer ($n = 17$)
- (7) Knockout mouse phenotype ($n = 86$)
- (8) Molecular pathway ($n = 35$)

b



Identification of SNPs associated to RA

From SNPs to causal genes through fine-mapping

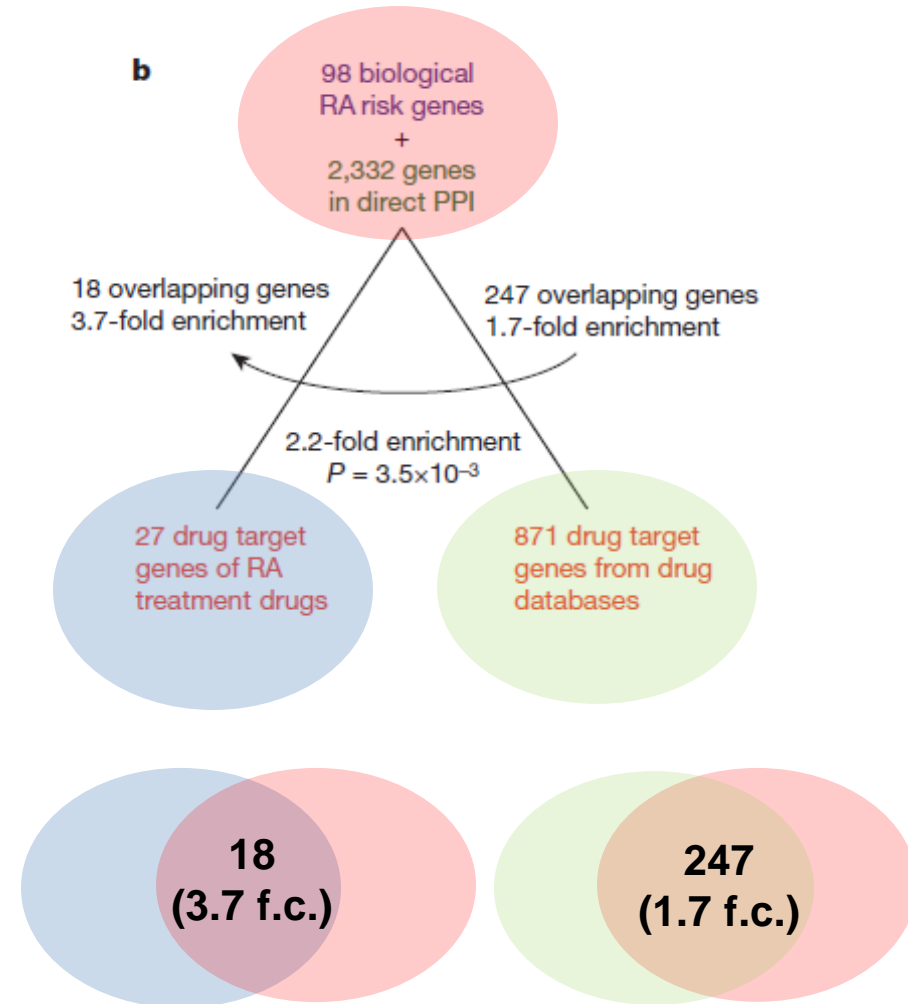
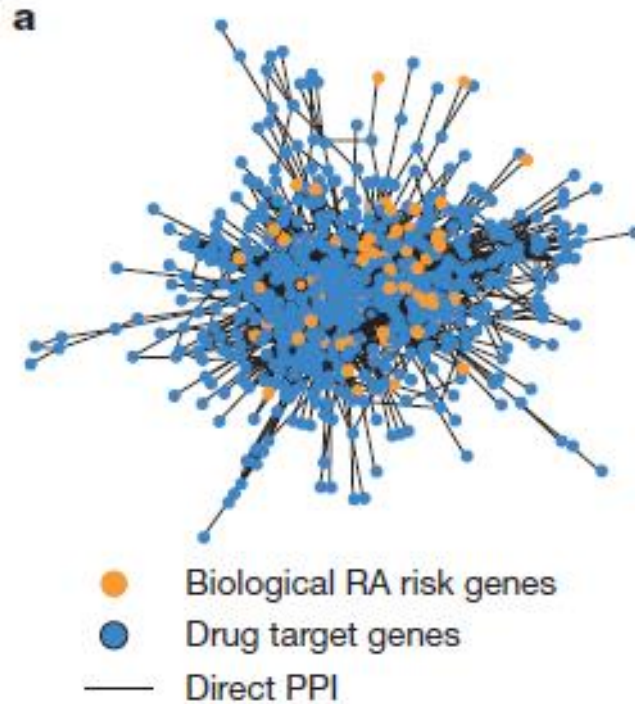
Characterization of results

Data integration and genes prioritization

Assessment of the workflow using validated targets

Assessment of the workflow using validated targets

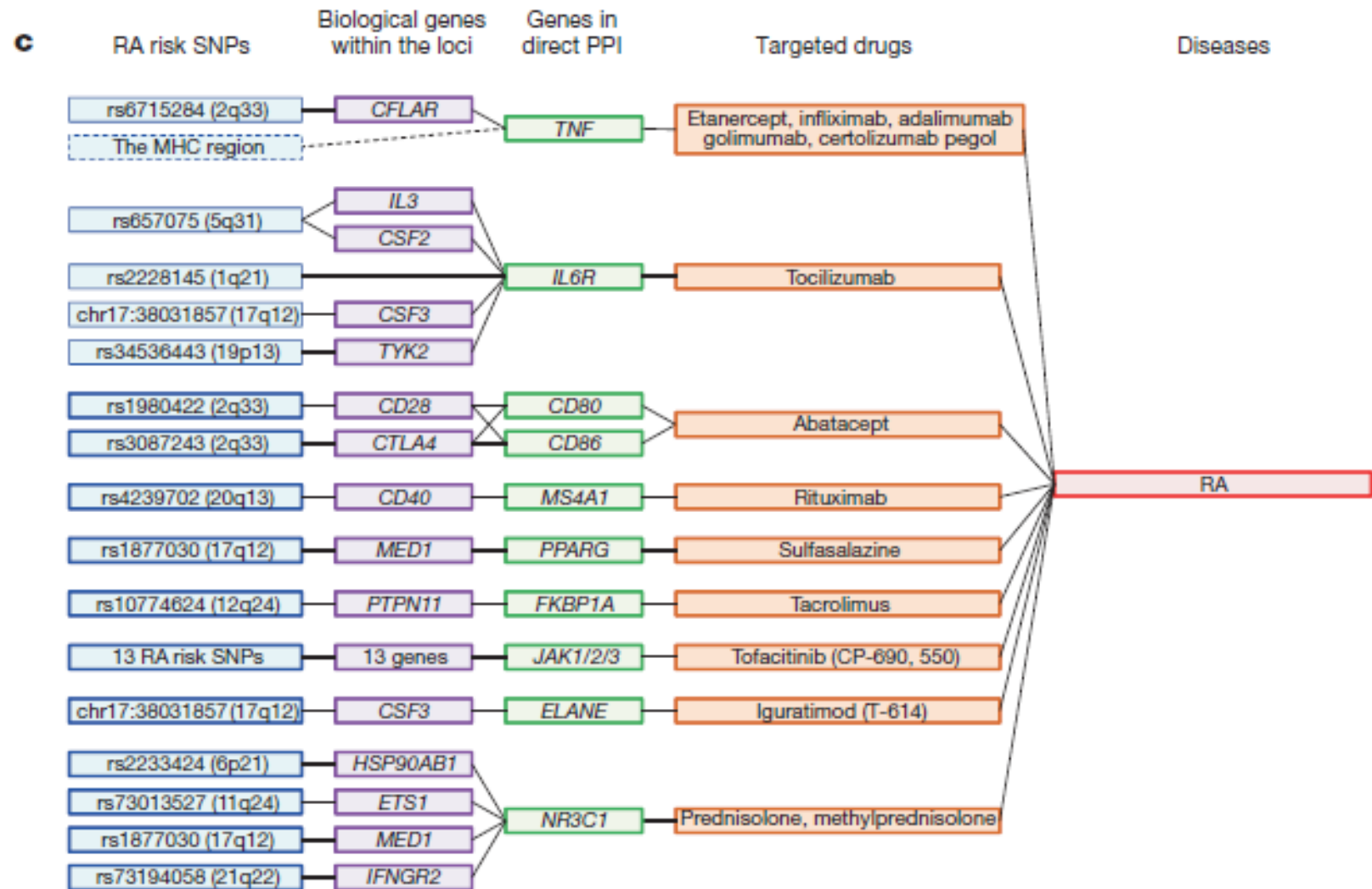
- 98 biological RA risk genes (score ≥ 2)
- +2,322 genes in PPI



➔ Strong enrichment for RA approved drugs (extracted from DrugBank TherapeuticTargets Database - TTD)

Mapping RA risk SNPs to drug targets

Example: PPI link to known RA drugs

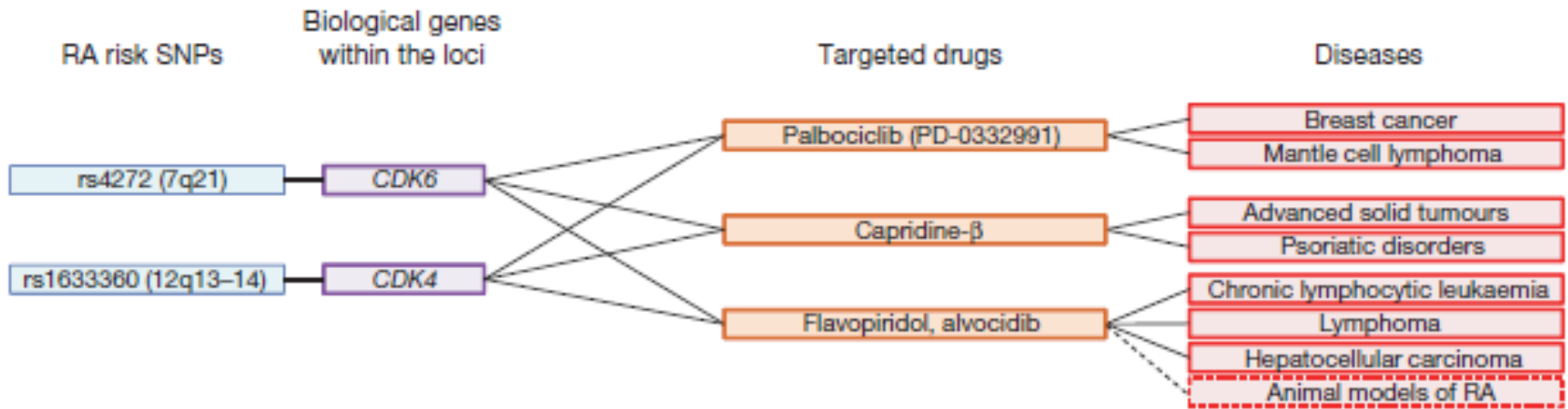


- 98 biological RA risk genes (score ≥ 2)

Drug Repurposing

Connections between RA genes and drugs indicated for other diseases

d



Study Summary

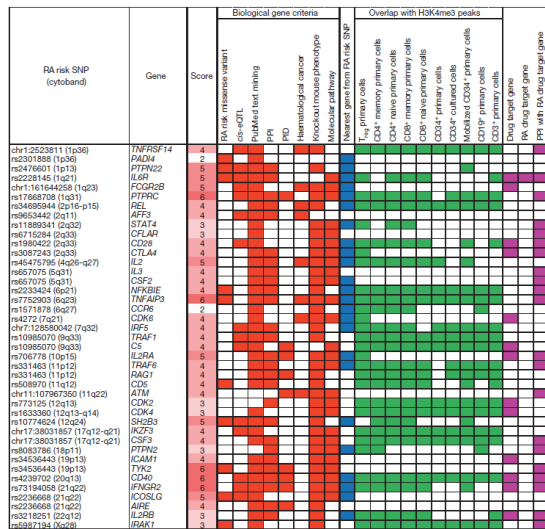
- Comprehensive genetic study with 100,000 subjects:
 - identified 42 novel RA risk loci
 - provided novel insights into RA pathogenesis.
 - demonstrated role of genetics for drug discovery
- Systematic approach to derive disease biological insights and novel drug candidates by integrating human genetic data with different layers of orthogonal information

Take Home Messages

- GWAS can be used to identify candidate regions and genes associated with risk of RA
- Testing millions of hypotheses implies hunting for very low p-values
- Using a series of strategies and additional data to understand the functions of associated genes to disease and prioritize them as possible targets
 - Epigenetics
 - PPI Network analysis
 - Link genetics to intermediate phenotypes (e.g. eQTLs)
- Enrichments for existing RA and other drugs supports the pipeline
- If new candidate RA genes can be targeted by existing drugs, drug repositioning opportunities can be evaluated

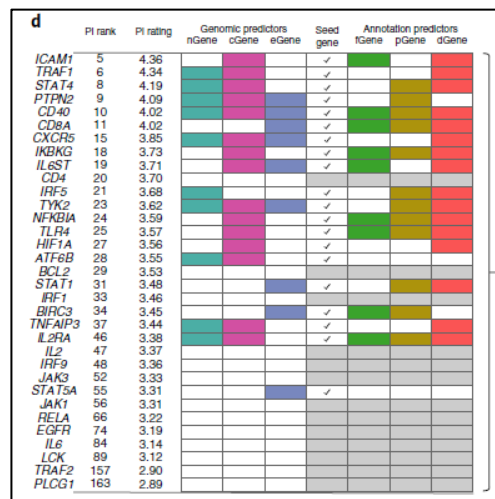
Similar Approaches to Data Integration

Genetics, functional genomics, network connectivity. RA



R. Plenge Nature 2014

Genetics, functional genomics, immune-related annotations, network connectivity. 30 immune traits



J. Knight Nat Genetics 2019

Genetics, animal models, text-mining, druggability, animal models, text mining, pathways. All diseases



Open Targets

<https://www.targetvalidation.org/>

SLE Molecular Immune Monitoring – Study Workflow

Personalized Immunomonitoring Uncovers Molecular Networks that Stratify Lupus Patients

Romain Banchereau,^{1,7} Seunghee Hong,^{1,7} Brandi Cantarel,¹ Nicole Baldwin,¹ Jeanine Baisch,¹ Michelle Edens,¹ Alma-Martina Cepika,¹ Peter Acs,¹ Jacob Turner,¹ Esperanza Anguiano,¹ Parvathi Vinod,¹ Shaheen Khan,² Gerlinde Obermoser,¹ Derek Blankenship,¹ Edward Wakeland,² Lorien Nassi,^{2,3} Alisha Gotte,^{2,3,4} Marilyn Punaro,^{2,3} Yong-Jun Liu,^{1,5} Jacques Banchereau,⁶ Jose Rosello-Urgell,¹ Tracey Wright,^{2,3} and Virginia Pascual^{1,3,4}

¹Baylor Institute for Immunology Research, Dallas, TX 75204, USA

²UT Southwestern Medical Center, Dallas, TX 75235, USA

³Texas Scottish Rite Hospital for Children, Dallas, TX 75219, USA

⁴Vanderbilt University School of Medicine, Nashville, TN 37232, USA

⁵MedImmune, Gathersburg, MD 20878, USA

⁶The Jackson Laboratory for Genomic Medicine, Farmington, CT 06030, USA

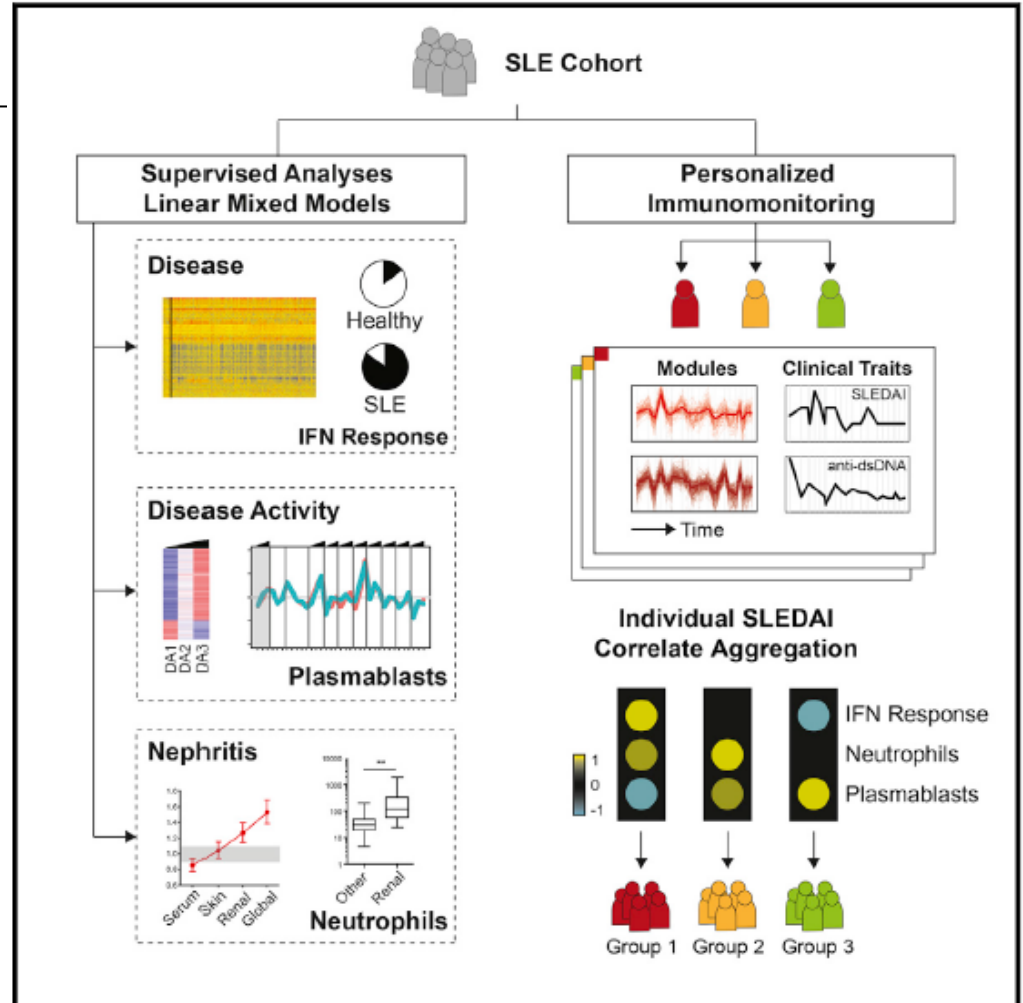
⁷Co-first author

*Correspondence: virginia.pascual@bwhhealth.org

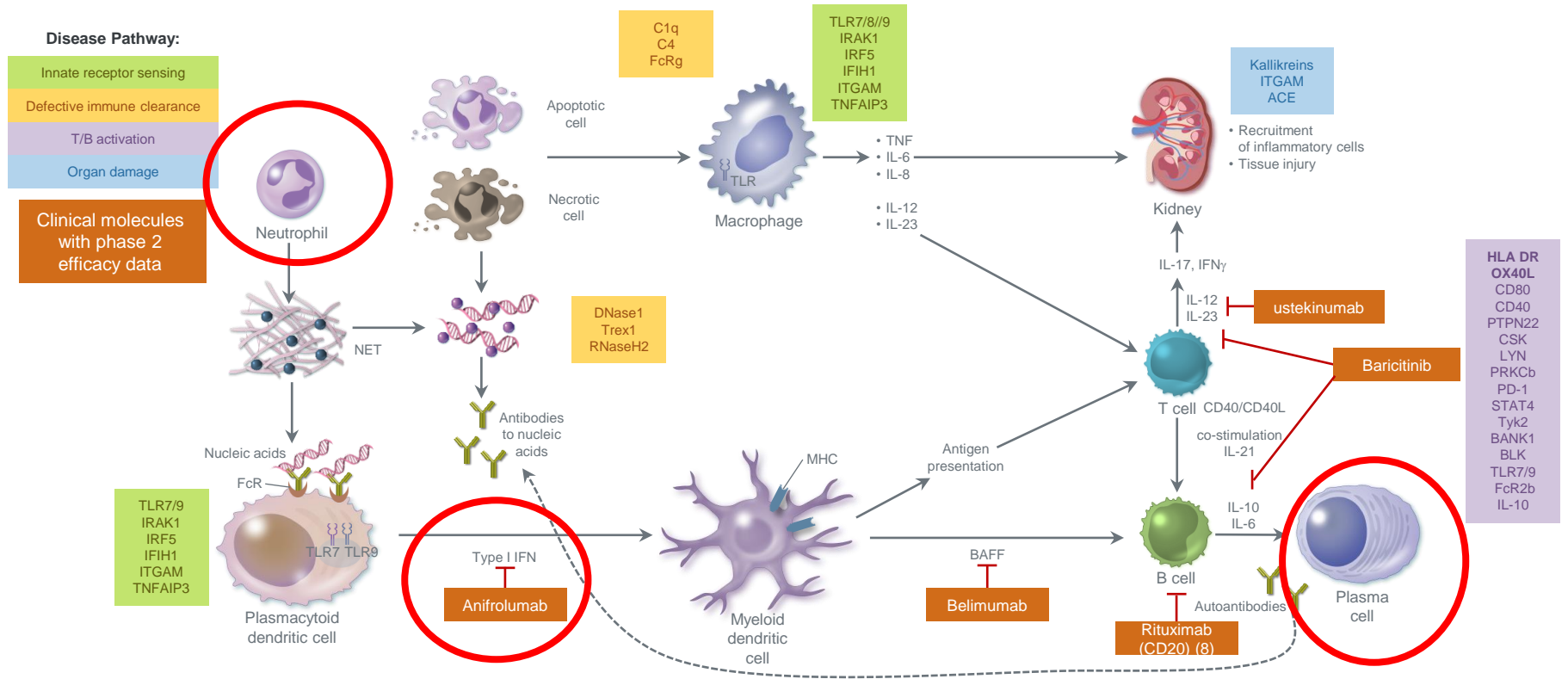
<http://dx.doi.org/10.1016/j.cell.2016.03.008>

Clinical and transcriptional profiling of 158 lupus pediatric patients, up to a period of 4 years

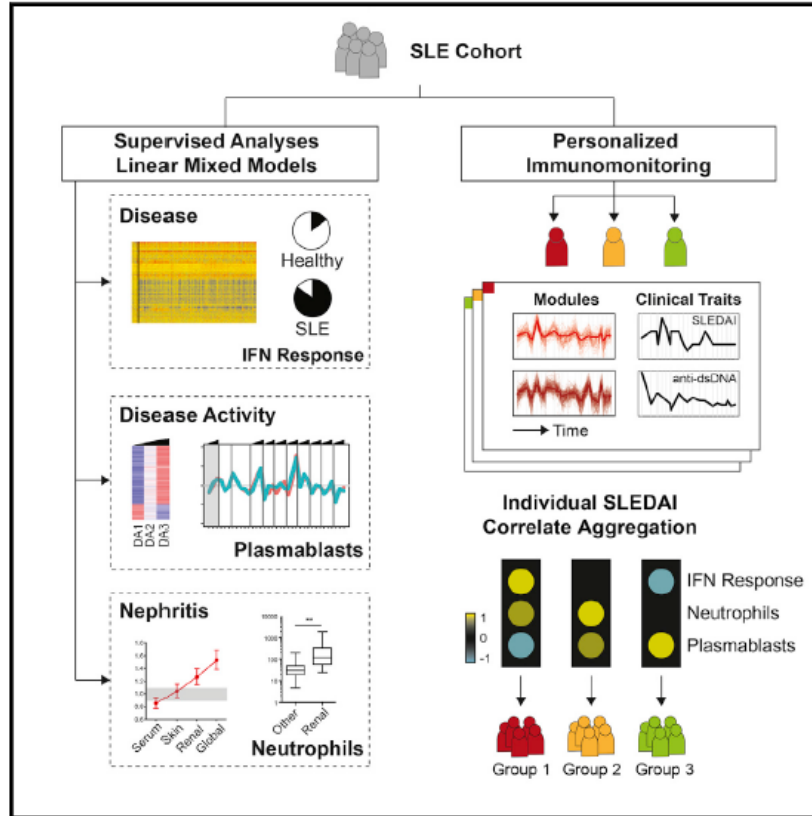
Cell. 2016 Apr 21;165(3):551-65



Cells and Pathways Driving SLE



Analytical Study Workflow



Genes associated to SLE, DA, Race, Treatment, disease subtypes

From genes to blood modules

WGCNA modules for each patient and correlation with SLEDAI

Linking WGCNA to blood modules

Stratification of Patients into Groups

- **Gene Expression Analysis**
 - Multivariate linear regression modelling
 - Heat maps and Hierarchical Clustering
 - Aggregating gene expression through modules
 - WGCNA modules
 - Blood modules
 - Gene Set Enrichment Analysis



SLE Blood Transcriptional Fingerprint

Genes associated to
SLE, DA, Race,
Treatment, disease
subtypes

From genes to "blood
modules"

WGCNA
modules for
each patient and
correlation with
SLEDAI

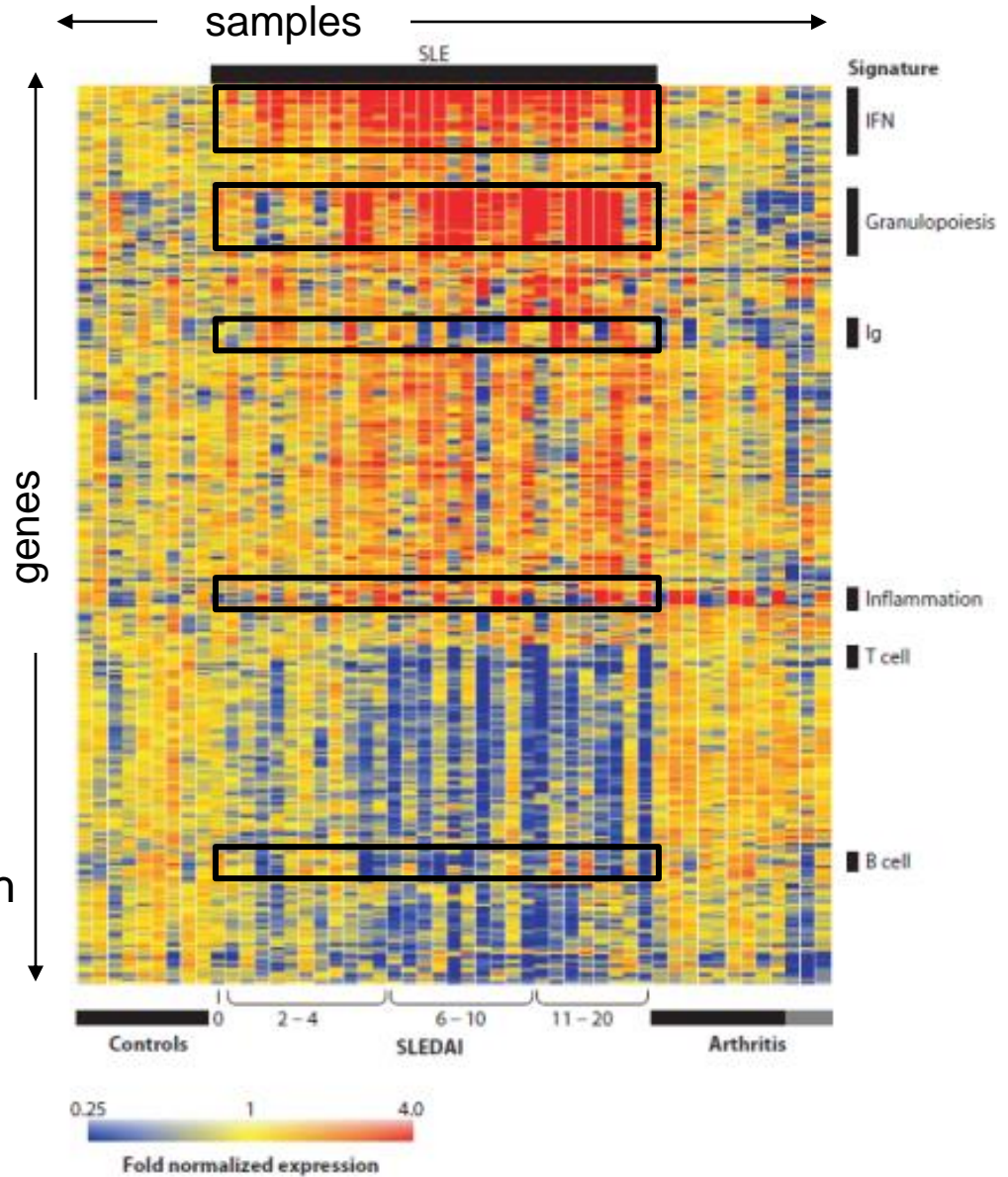
Linking WGCNA
to blood modules
for inference of
biological
function

Stratification of
patients into
groups

Gene Signatures



	Sample 1	Sample 2	Sample 3	Sample 4	Sample 5	Sample 6
Gene 1	1235	546	943	263	136	314
Gene 2	1266	32	556	435	687	2718
Gene 3	947	2829	389	3820	2039	1414
Gene 4	392	2398	84	829	4392	512
...



- Group of genes whose expression values, altogether, are associated with a given feature.
- Genes in signatures often show coordinated expression levels although this is not a requirement.

Annu. Rev. Immunol. 2010. 28:535–71

Differential Expressed Genes: Multivariate Linear Model



Example: 2 groups – SLE and Healthy Controls

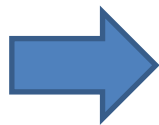
$$y_{gi} = \beta_0 + \beta_1 x_{1i} + \beta_2 x_{2i} + \dots + \beta_p x_{pi} + \epsilon_g$$



Explanatory Variables (e.g. SLE/Healthy, Disease Activity, Treatment etc.)



Expression of gene A



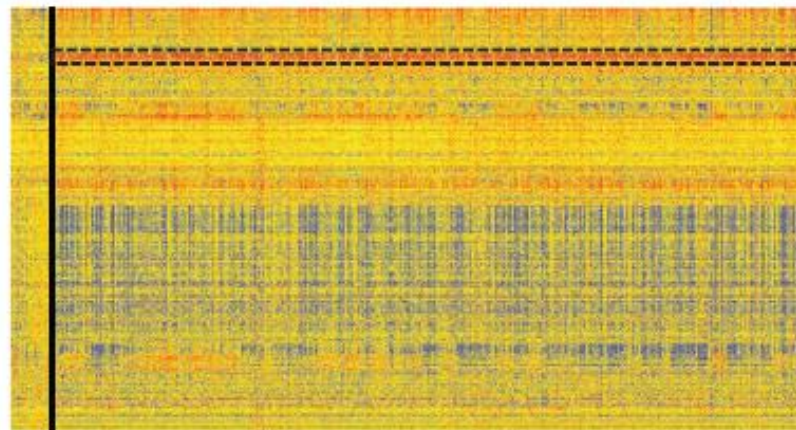
- Values of B coefficients
- P-value of B coefficients being different than 0

Genes Associated with SLE

Interferon response

A

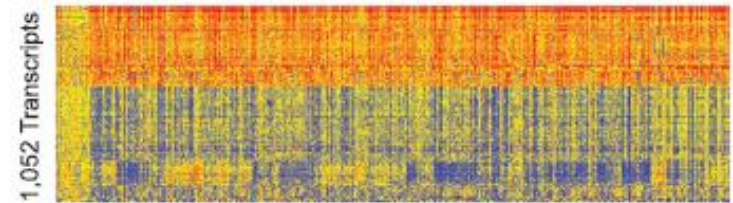
Dallas Pediatric SLE Cohort
 158 SLE subjects (SLE)
 48 healthy controls (H)
 972 samples



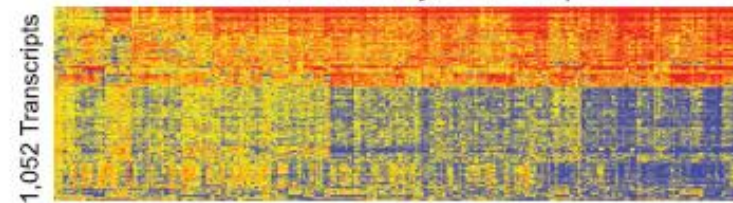
DDX60
 DHX58
 GBP1
 IFI27
 IFI35
 IFIH1
 IFIT1
 IFIT2
 IFIT3
 IFIT5
 IFITM1
 IFITM3
 IRF7
 IRF9
 ISG15
 ISG20
 LAMP3
 MX1
 OAS1
 OAS2
 OAS3
 OASL
 OTOF
 SP140
 STAT1
 STAT2
 TAP1
 TRIM22
 TRIM5
 USP18
 USP41

B

Training Set
 106 SLE / 31 healthy / 649 samples

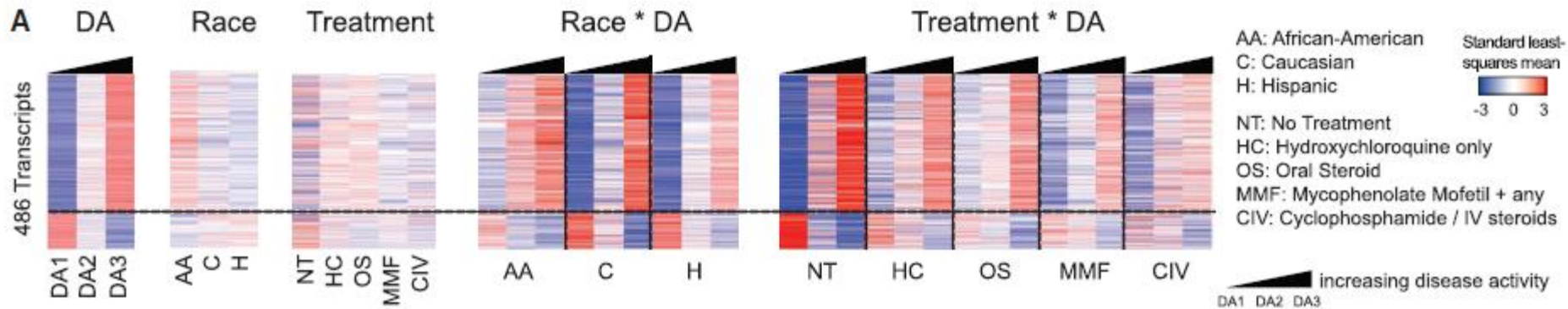


Test Set
 52 SLE / 17 healthy / 323 samples



➔ 1.052 genes differentially expressed in SLE vs Healthy

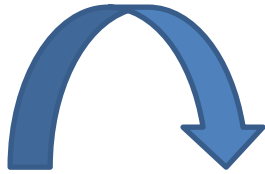
Genes Associated With SLE Disease Activity



- 486 Transcripts Differentially Expressed between DA1 (SLEDAI: 0-2) and DA3 (SLEDAI >7)
- Results stratified by Race, Treatment

SLE Blood Transcriptional Fingerprint

- % of genes up/down
- QuSage fold-change



Genes associated to
SLE, DA, Race,
Treatment, disease
subtypes

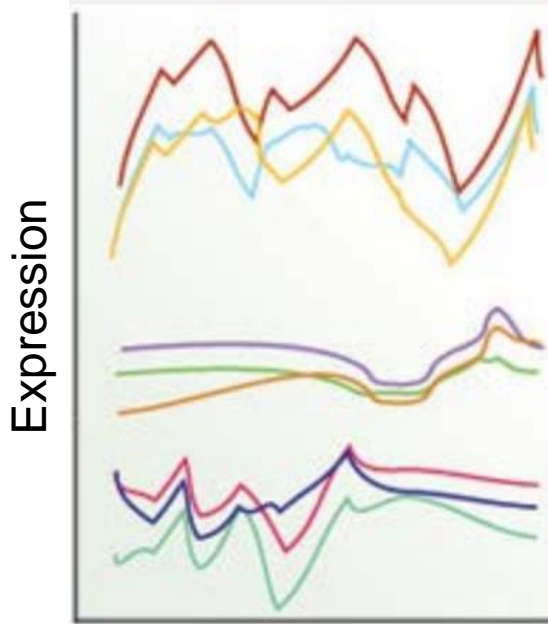
From genes to “Blood
Modules”

WGCNA
modules for
each patient and
correlation with
SLEDAI

Linking WGCNA
to blood modules
for inference of
biological
function

Stratification of
patients into
groups

Building Transcriptional Blood Modules



Blood Samples (239)

Blood Module 1

Blood Module 2

Blood Module 3

239 Blood Samples:

- systemic juvenile idiopathic arthritis (n = 47)
- systemic lupus erythematosus (n = 40)
- type I diabetes (n = 20)
- metastatic melanoma (n = 39)
- acute infections (Escherichiacoli [n = 22]
- Staphylococcus aureus [n = 18], Influenza A [n = 16])
- liver-transplant recipients undergoing immunosuppressive therapy (n = 37).

260 blood modules identified

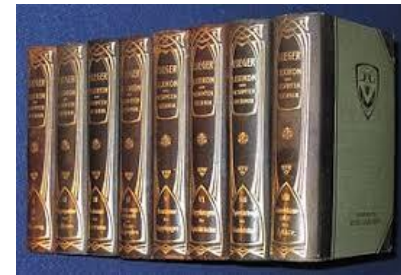
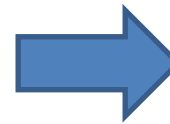
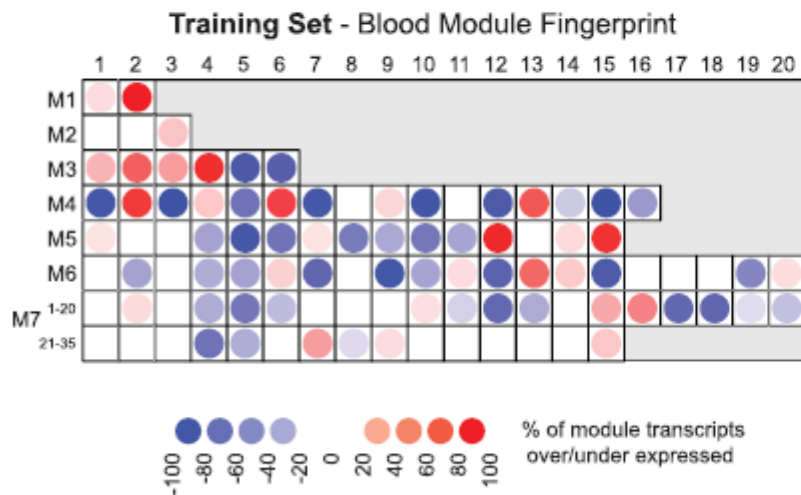


Table 1. Functional Interpretation of Transcriptional Modules

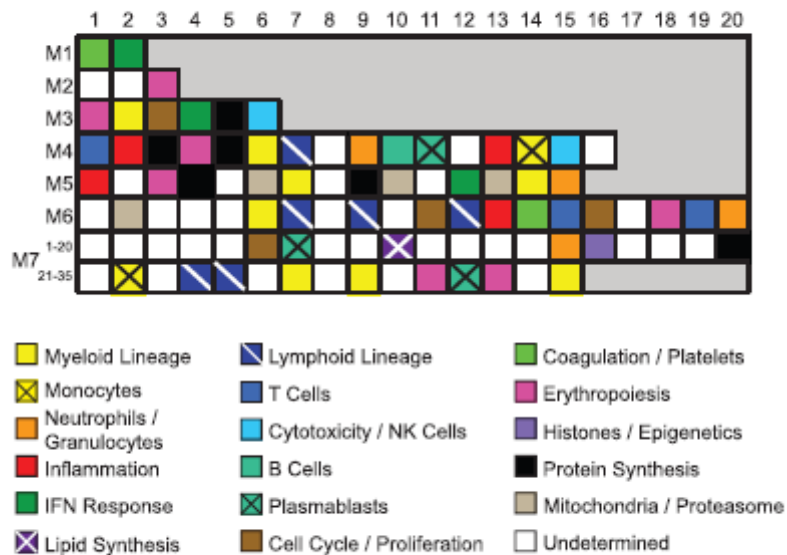
Module I.D.	Number of Probe Sets	Keyword Selection	Interpretation
M 1.1	76	Ig, Immunoglobulin, Bone, Marrow, PreB, IgM, Mu.	<u>Plasma cells.</u> Includes genes coding for Immunoglobulin chains (e.g., IGHM, IGJ, IGLL1, IGKC, IGHD) and the plasma cell marker CD38.
M 1.2	130	Platelet, Adhesion, Aggregation, Endothelial, Vascular	<u>Platelets.</u> Includes genes coding for platelet glycoproteins (ITGA2B, ITGB3, GP6, GP1A/B) and platelet-derived immune mediators such as PPPB (pro-platelet basic protein) and PF4 (platelet factor 4).

SLE vs Healthy: From Genes to Modules

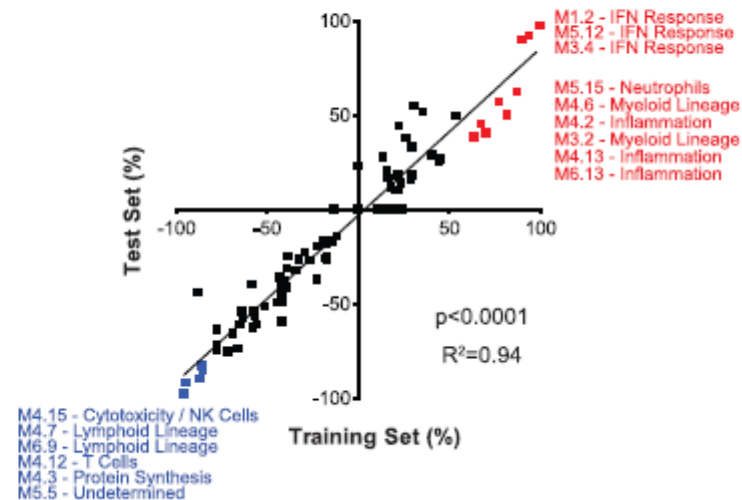
C Modules perturbed in SLE vs Healthy



Blood Module Functional Map



D Blood Module Fingerprint Reproducibility

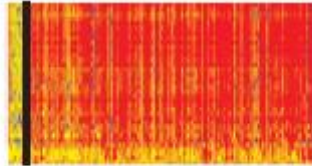


- **Over-expression** of IFN response, neutrophil, inflammation, cell cycle, erythropoiesis, and histone modules.
- **Down-regulation** of NK cell/cytotoxicity, lymphoid lineage, B cells, T cells, and protein synthesis

Over-expression of IFN, plasmablast and neutrophil module genes

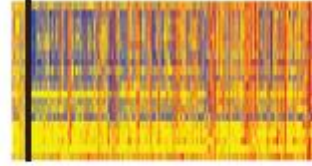
E

M1.2 - IFN Response



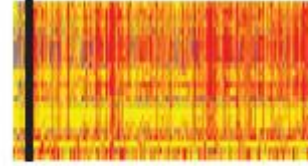
H SLE

M4.11 - Plasmablasts

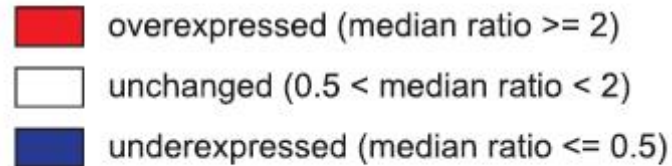
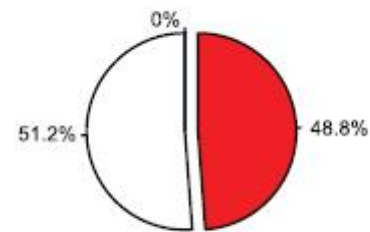
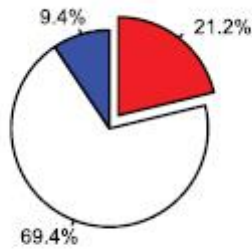
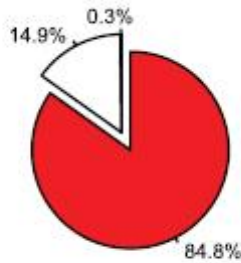


H SLE

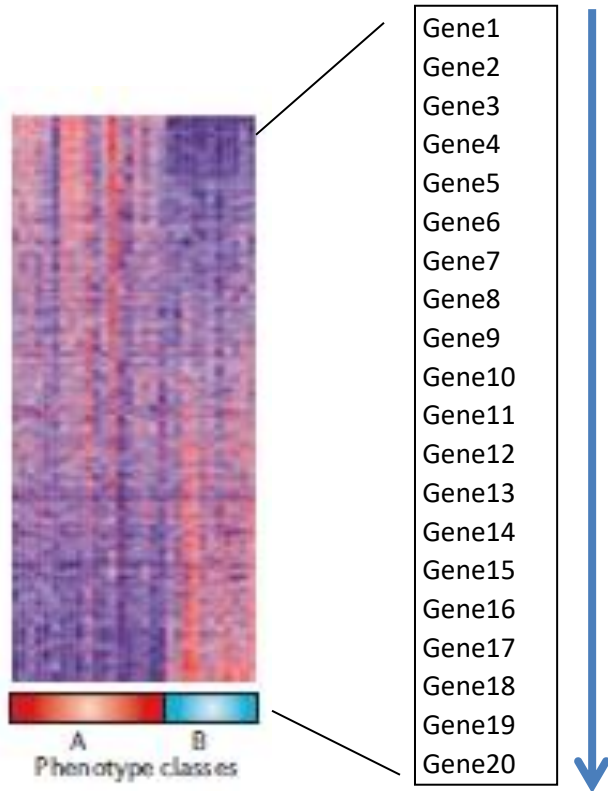
M5.15 - Neutrophils



H SLE

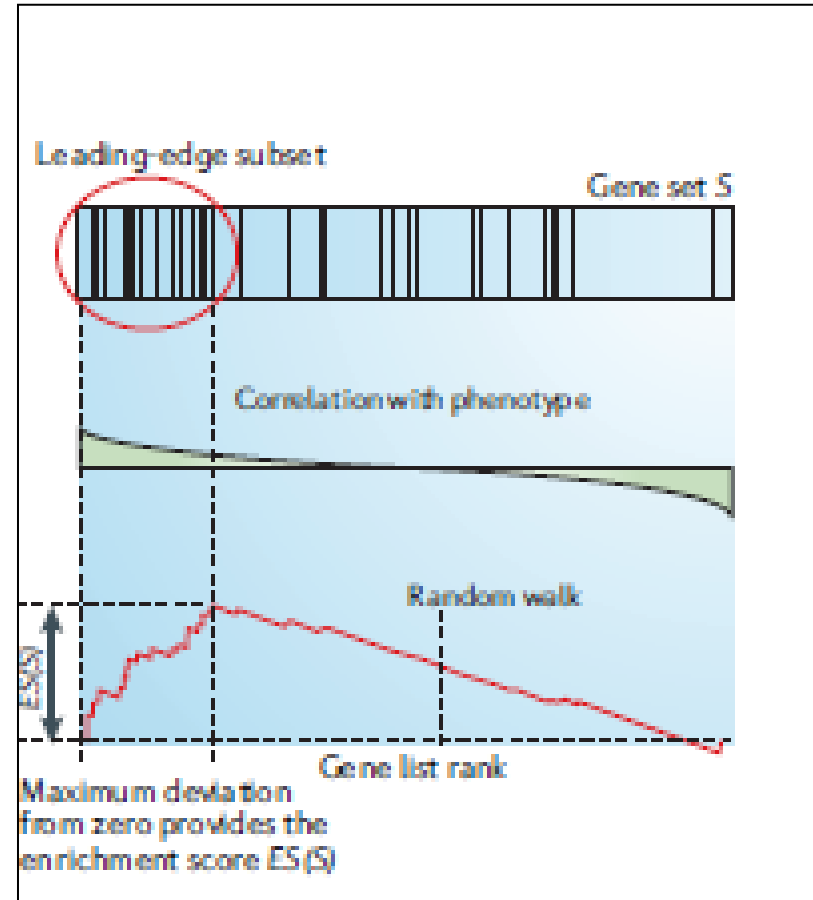
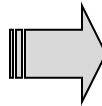


Gene Set Enrichment Analysis (GSEA)



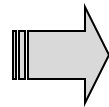
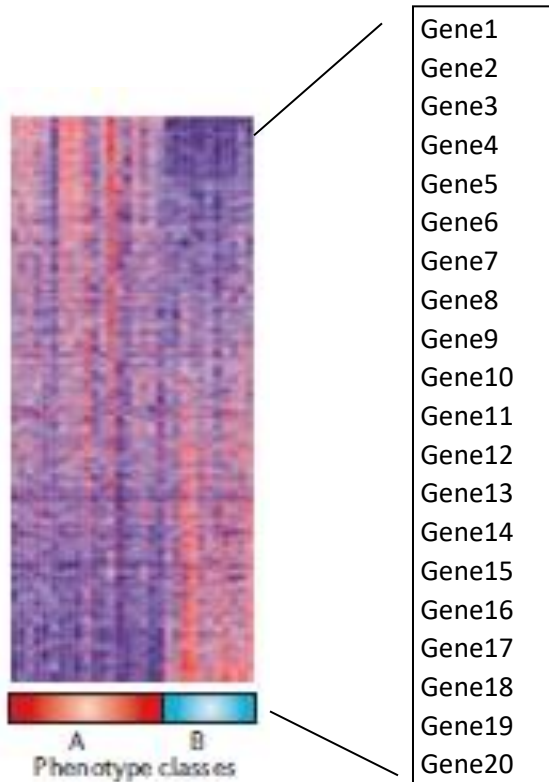
Genes are ranked according to their fold-change

Gene set databases
KEGG
GenMAPP
BioCarta
Expression signatures
Cytogenetic loci
Amplifications
Etc.

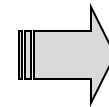


An enrichment score is calculated for each pathway, taking into account the directionality of the input list

From Gene Expression To Gene Set Scores

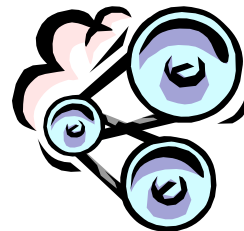


Gene set databases
KEGG
GenMAPP
BioCarta
Expression signatures
Cytogenetic loci
Amplifications
Etc.



- **Apoptosis** (p-val=0.001)
- **Cell Cycle**(p-val=0.00004)

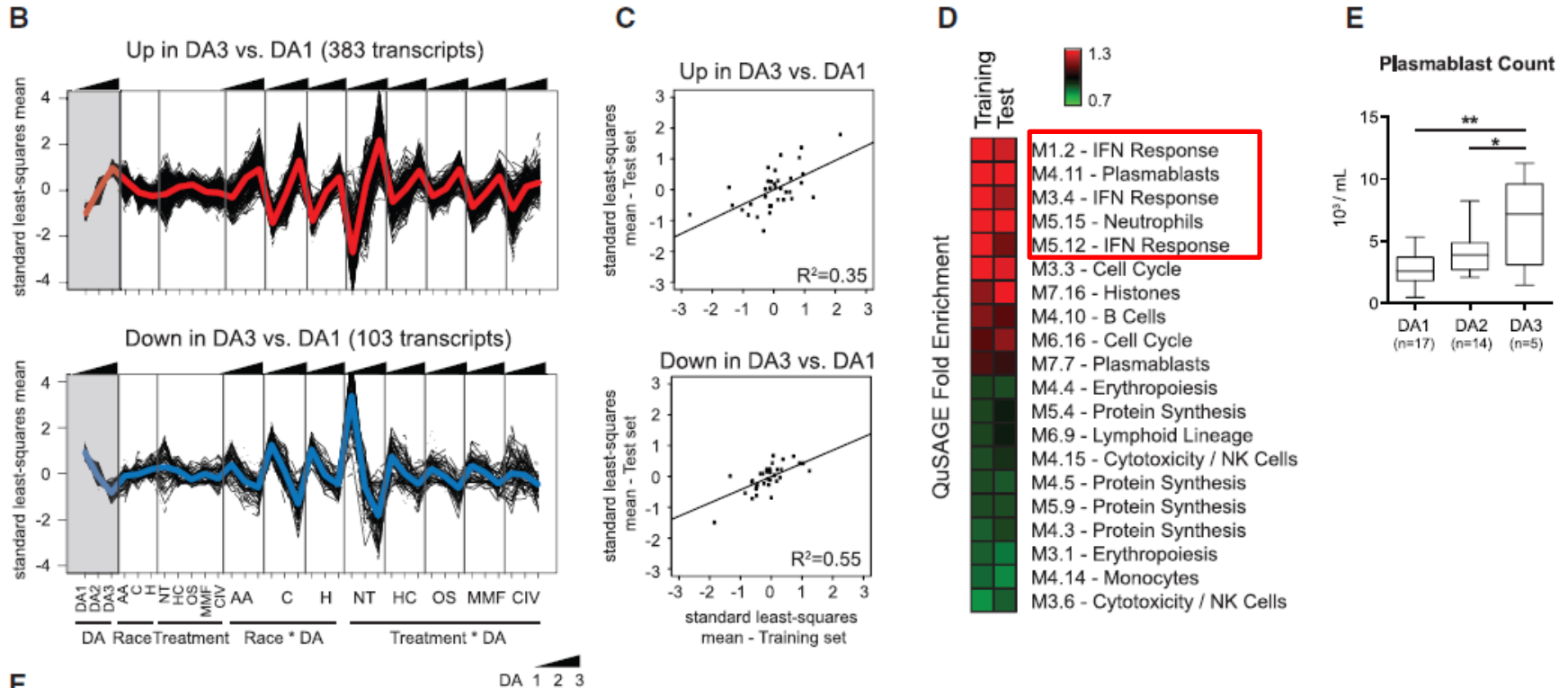
List of perturbed pathways



Bioinformatics analysis using pathway DBs

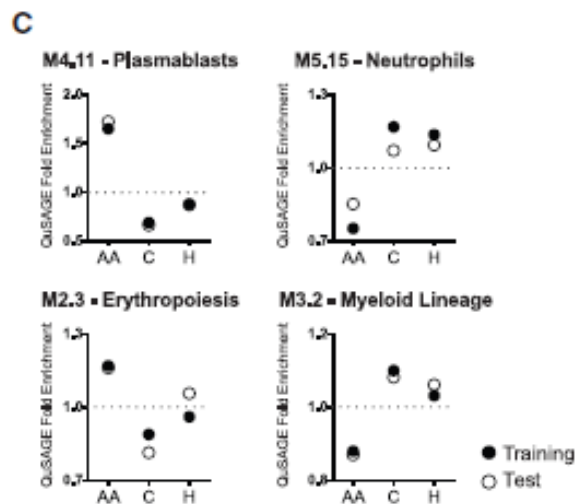
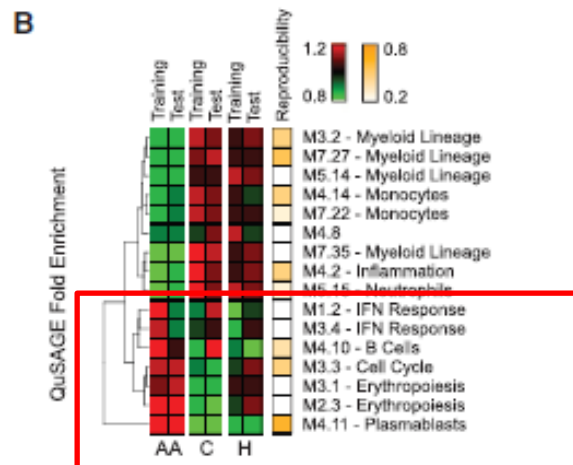
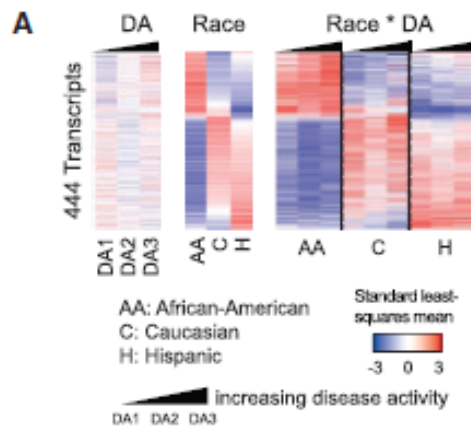
List of perturbed genes (differentially expressed between class A and B)

From Genes to Modules Associated with Disease Activity



→ Genes associated with DA are enriched for **IFN and Plasmablast modules**

Genes and Modules Associated With Race

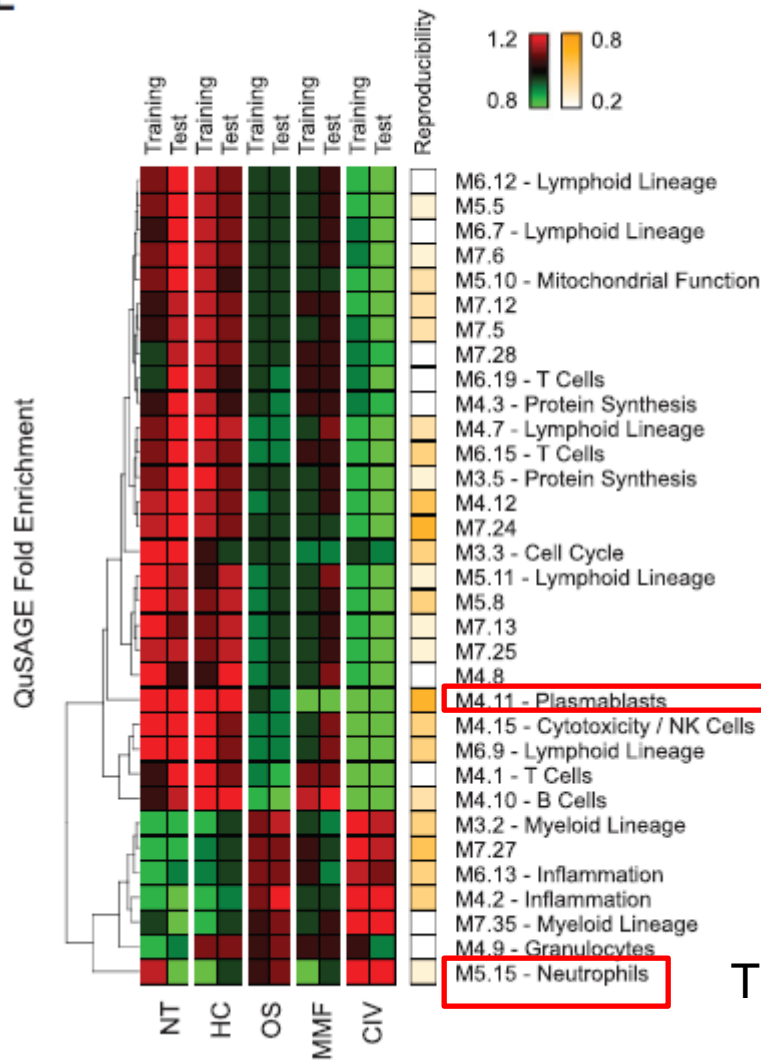


Cell. 2016 Apr 21;165(3):551-65

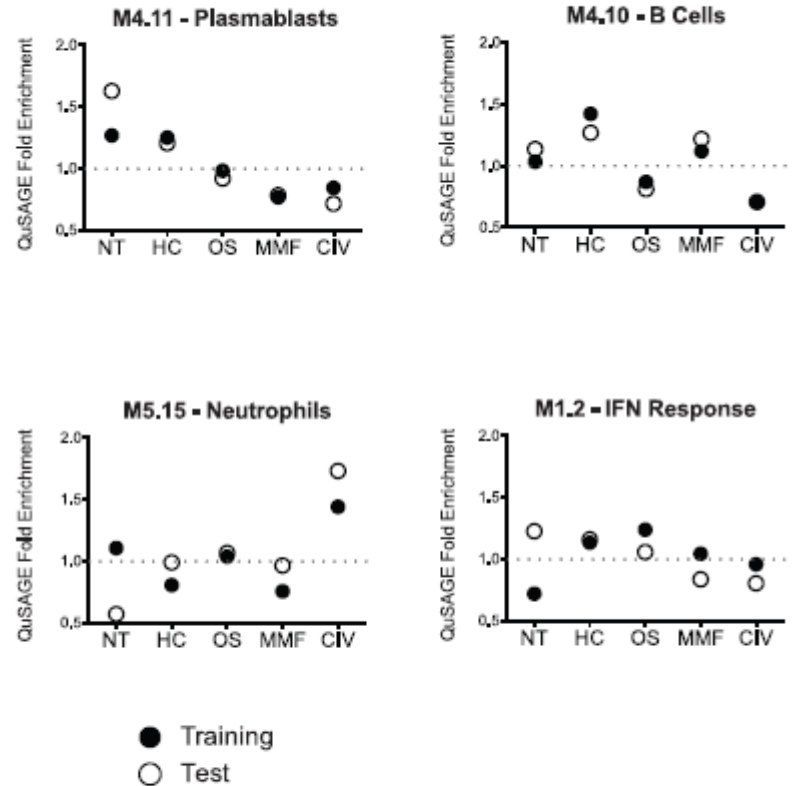
Increased Plasmablast Responses in African-American Patients

Modules Associated With Treatment

E

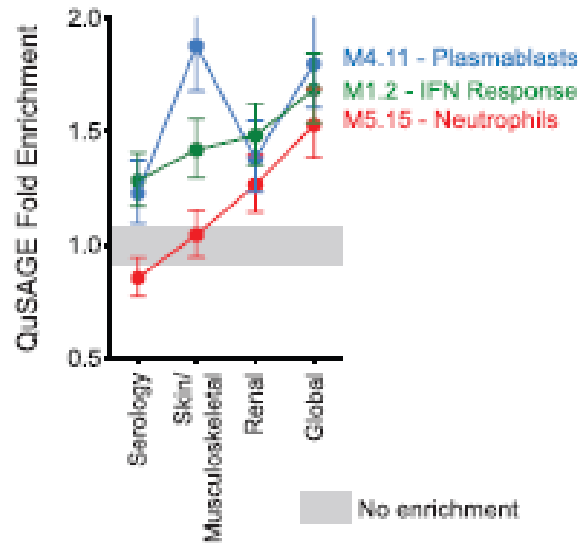
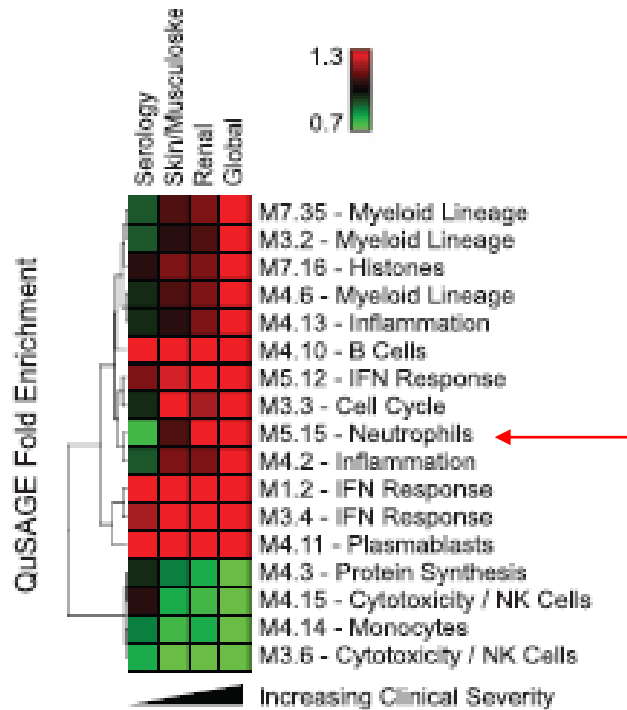


F

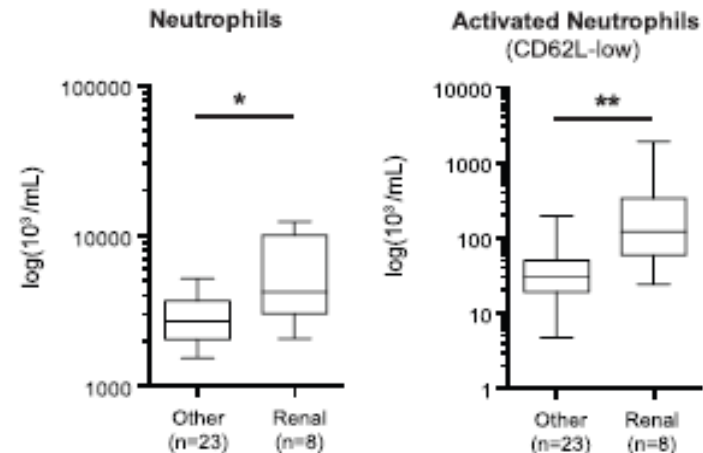


The plasmablast signature was decreased by all treatments compared to NT, but most strongly by (MMF) and CIV, two cytostatic drugs that suppress activated lymphocytes

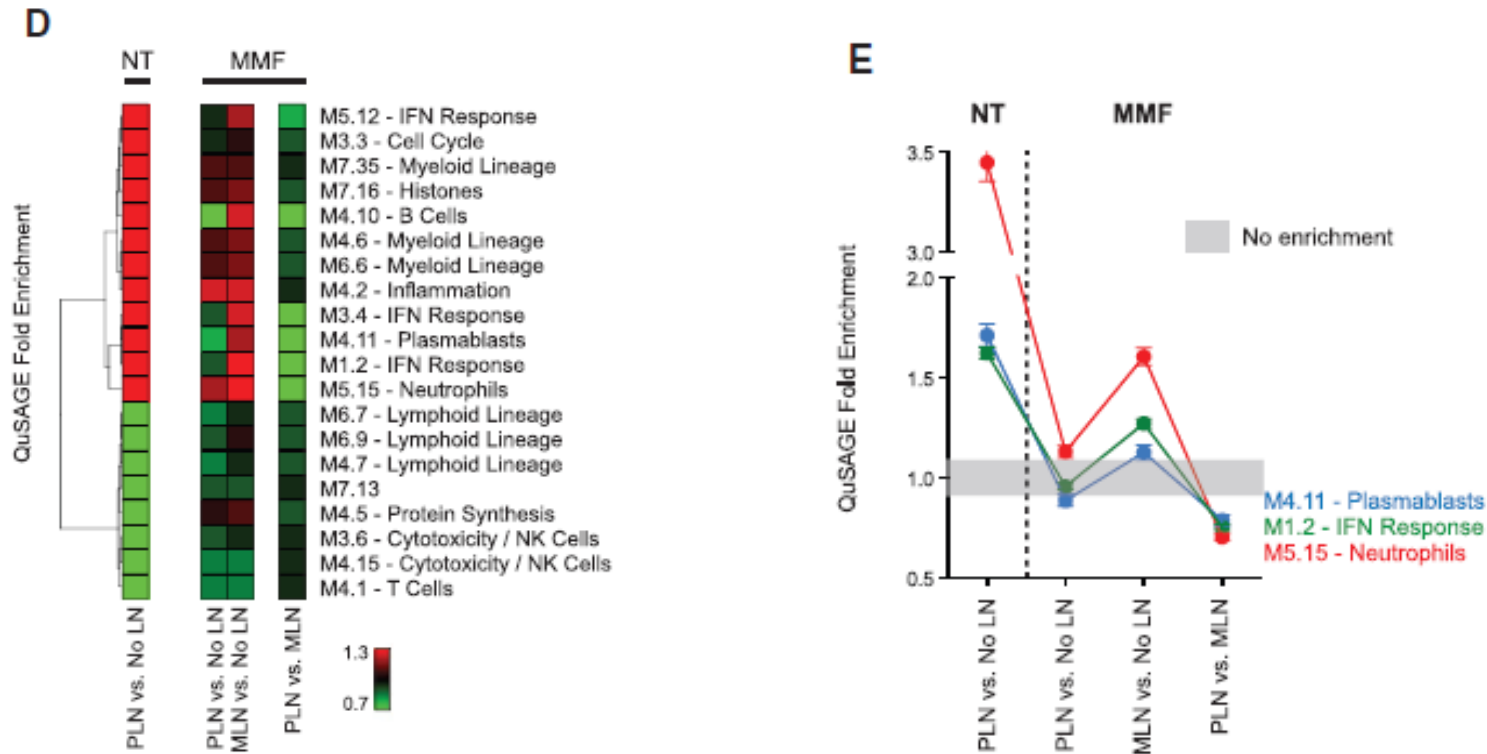
Modules associated with Disease Types



Neutrophil module is associated with Lupus Nephritis



Modules associated with treatment in different nephritis subclasses



Distinct signatures in response to treatment in different nephritis subclasses: PLN (proliferative nephritis) vs MLN (membranous nephritis), treated with MMF (mycophenolate mofetil)

SLE Blood Transcriptional Fingerprint

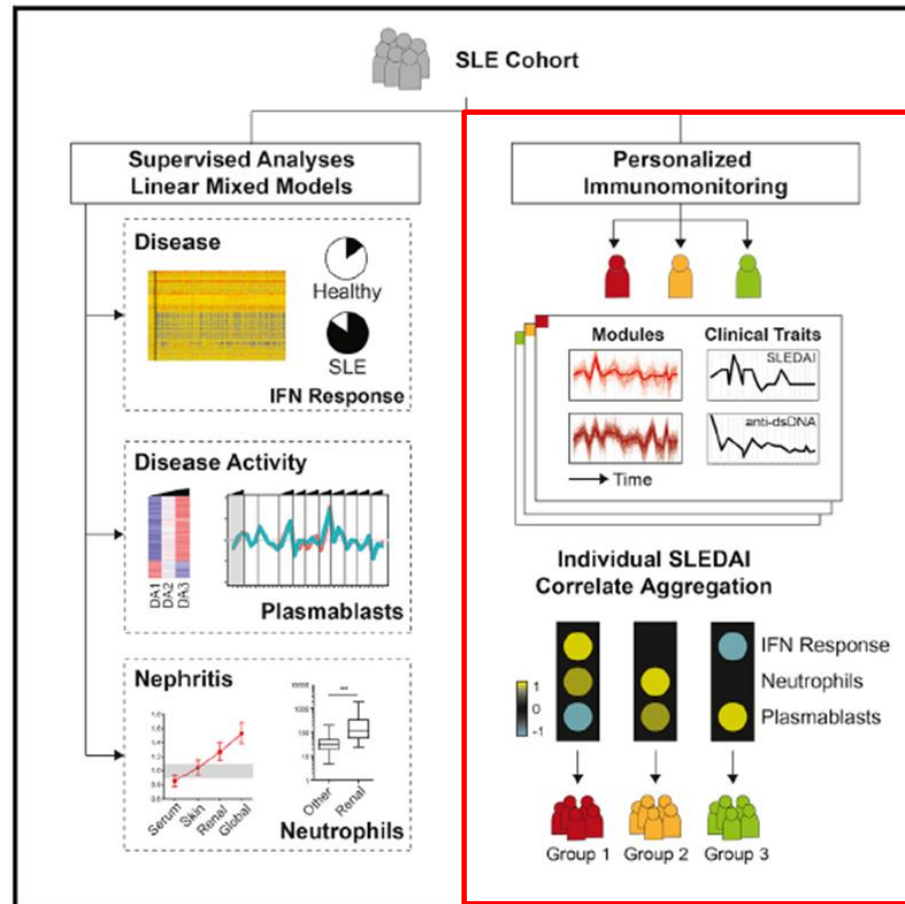
Genes associated to SLE, DA, Race, Treatment, disease subtypes

From genes to "blood" modules

WGCNA modules for each patient and correlation with SLEDAI

Linking WGCNA to blood modules for inference of biological function

Stratification of patients into groups



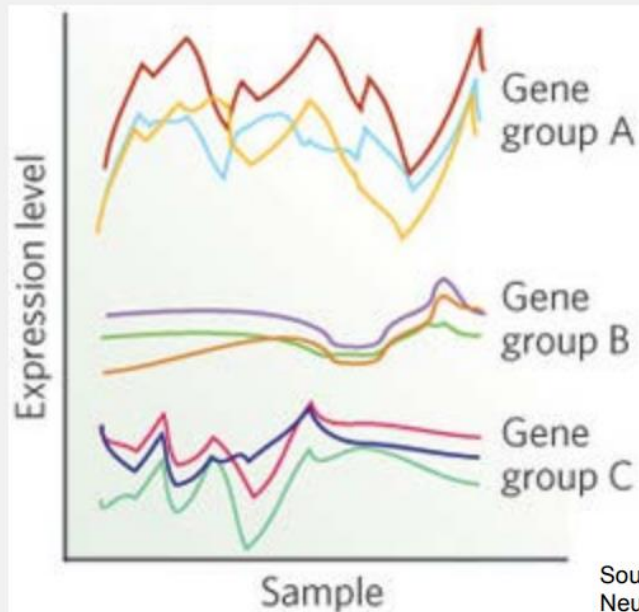
WGCNA – Weighted Gene Correlation Network



Aim of WGCNA: summarizing individual genes into modules, based on correlation

Modules found in WGCNA:

Groups of co-expressed genes (with similar expression profiles over a large group of individuals)



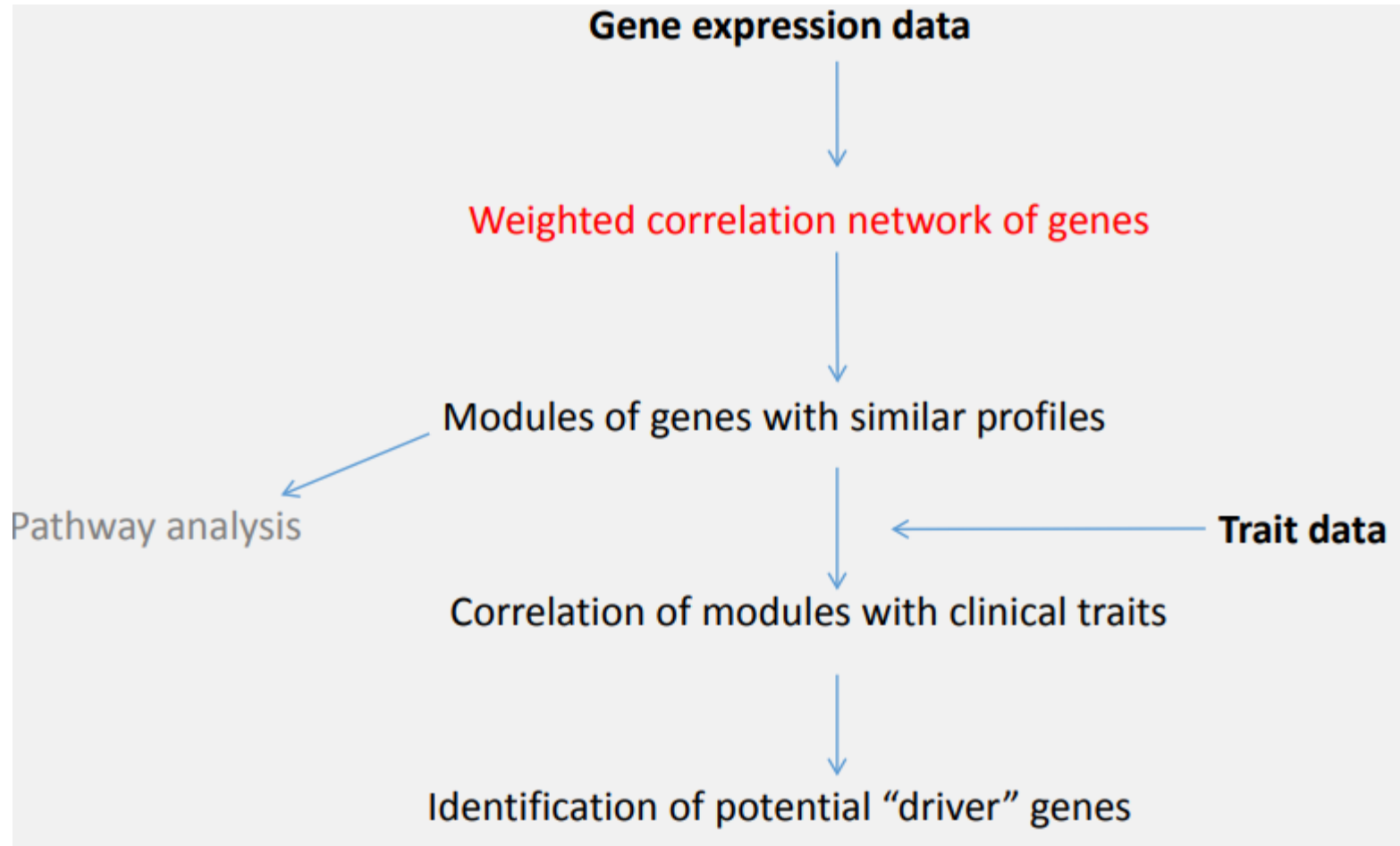
Source: Daniel H. Geschwind & Genevieve Konopka. Neuroscience in the era of functional genomics and systems biology, Nature 461, 908-915

Central Hypothesis:

Genes with **similar expression patterns** are of interest because they may be

- tightly co-regulated
- functionally related
- members of the same pathway

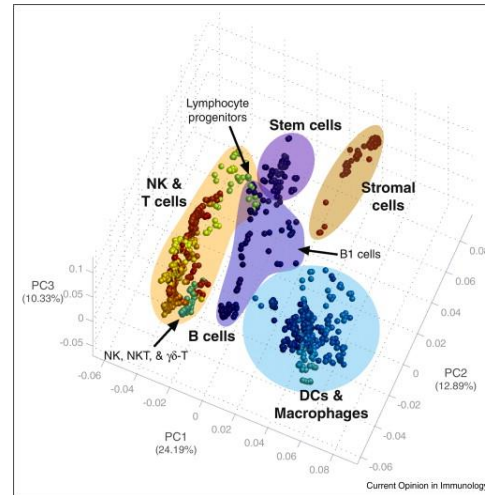
WGCNA – Workflow



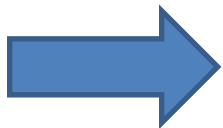
Measuring Modules with One Metrics: Eigengenes



	WGCNA MODULE X							
	Gene 1	Gene 2	Gene 3	Gene 4	Gene 5	Gene 6	Gene 7	Eigengene Value
sample 1	17	55	80	41	3	70	70	A
sample 2	43	100	56	91	72	22	2	B



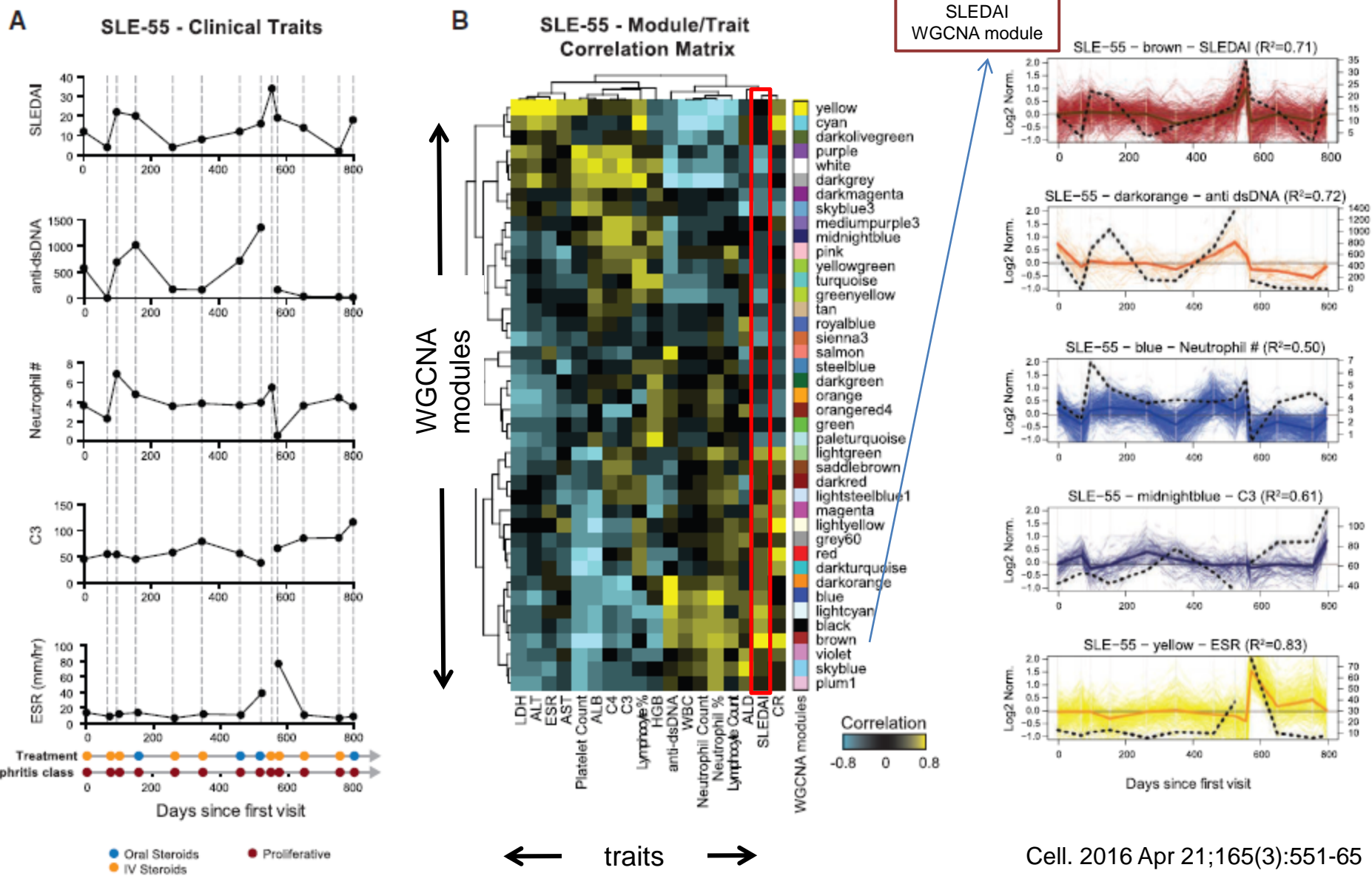
Kim and Lanier – Curr Opin Immunol 2013



- Module's "eigengene" is the first principal component of the expression matrix of the corresponding module
- It can be used as summary score for a vector of genes in a sample

Patient 55: Linking WGCNA modules to clinical traits by correlation

Patient- specific modules of co-expressed transcripts over time are identified by WGCNA. Analysis run on WGCNA modules (eigengenes) correlated with continuous clinical traits



Patient 55: Linking WGCNA Modules to Blood Modules

Genes associated to SLE, DA, Race, Treatment, disease subtypes

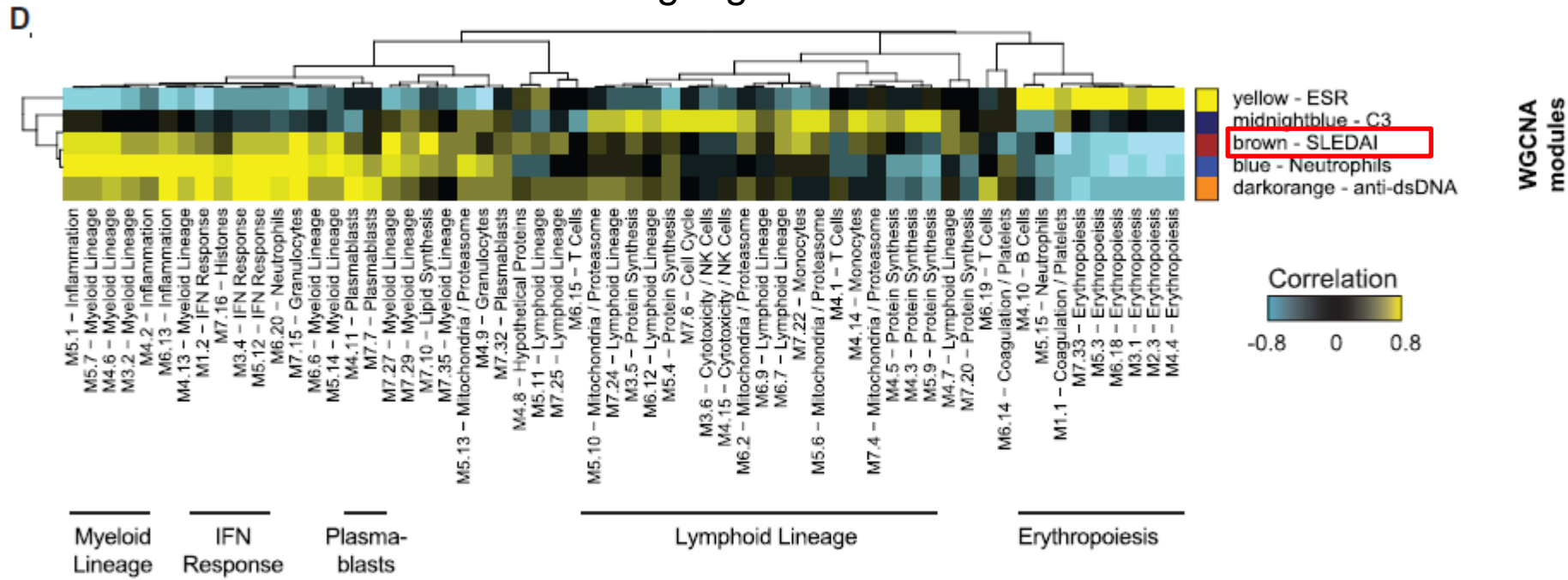
From genes to "blood modules"

Extract WGCNA modules for each patient and correlation with SLEDAI

Linking WGCNA to blood modules for inference of biological function

Stratification of patients into groups

Correlation between patient SLE-55 WGCNA and blood modules using eigengenes



SLE Blood Transcriptional Fingerprint

Genes associated to
SLE, DA, Race,
Treatment, disease
subtypes

From genes to “blood”
modules

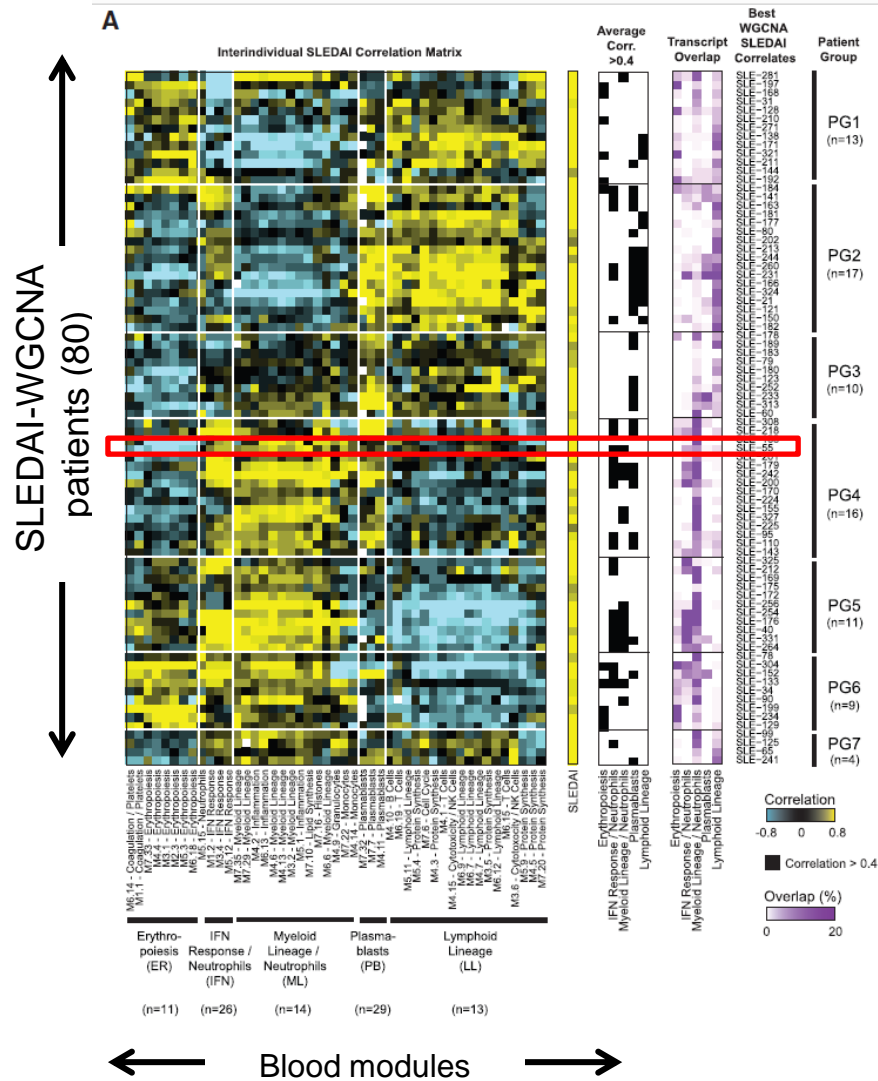
WGCNA
modules for
each patient and
correlation with
SLEDAI

Linking WGCNA
to blood modules
for inference of
biological
function

Stratification of
patients into
groups

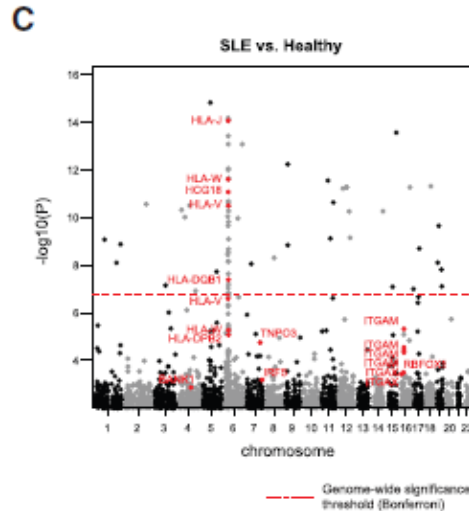
Stratification of SLE Patients Based on Transcriptional Correlates of SLEDAI

matrix of correlation between the SLEDAI WGCNA and blood modules, for all patients

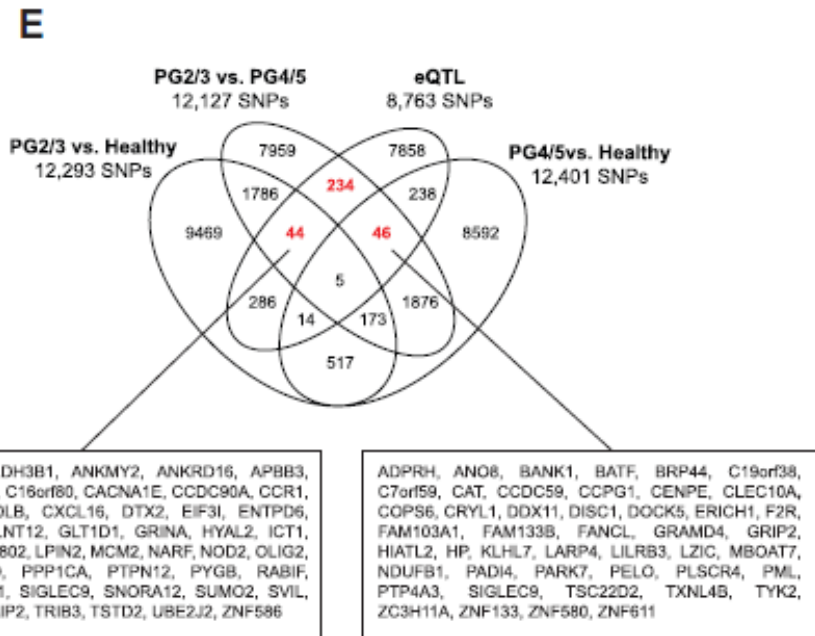
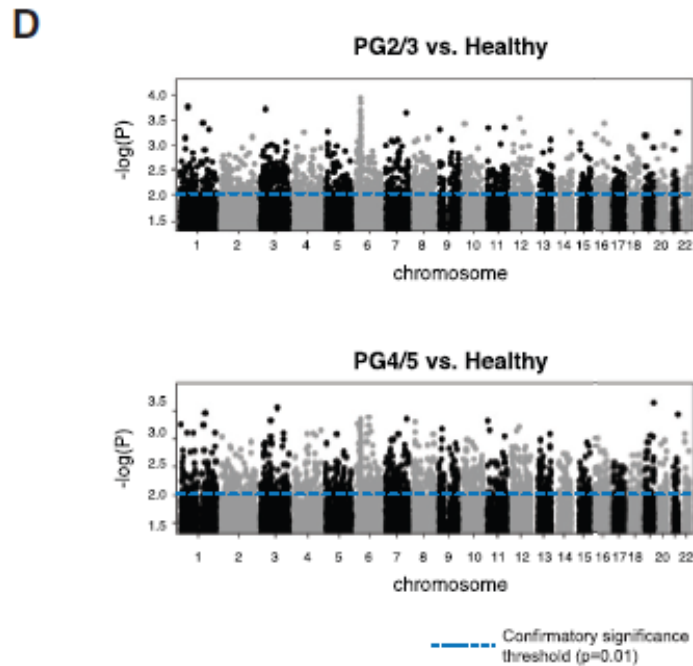


➤ 7 patient groups identified based on the combination of blood immune signatures that best correlate with the SLEDAI-WGCNA.

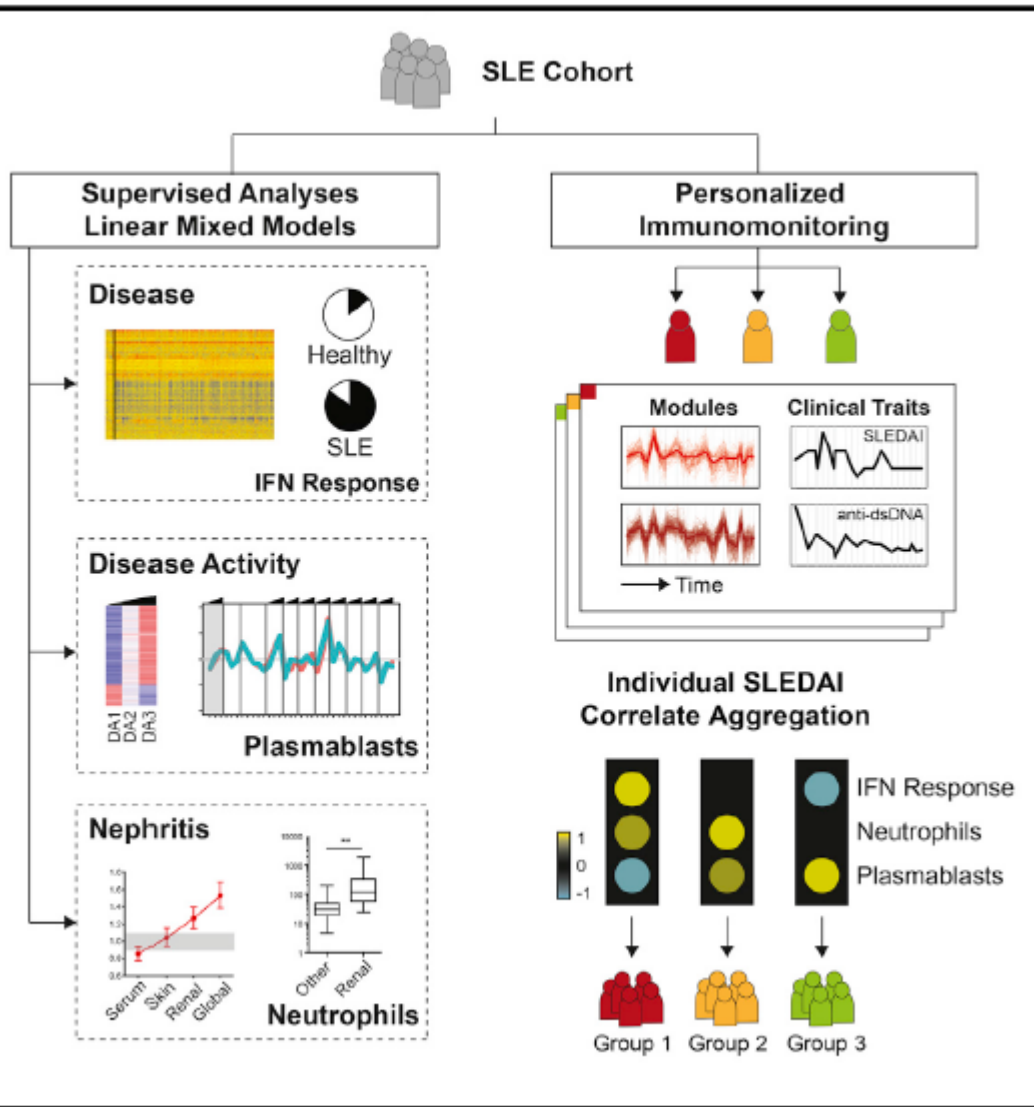
Genetic Analysis of Patients Groups



- To find a genetic basis for these clusters, SNP analysis was conducted (135 patients + HCs)
- SNPs differentiating between PG2/3 and PG4/5 were found
- Intersection with eQTL SNPs (SNPs associated with DA genes expression) points to IFN-inducible genes



Let's Recap

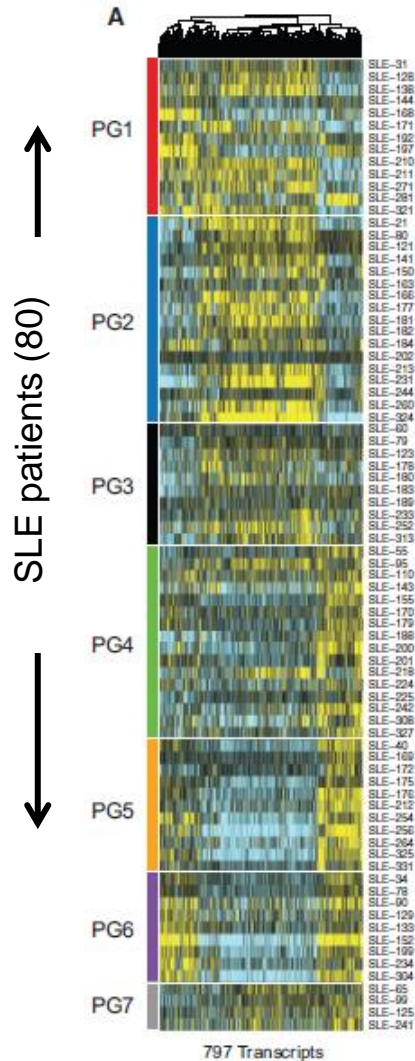


- Supervised analysis, linear models and blood module analyses led to identification of:
 - i) IFN response module in SLE
 - ii) Plasmablasts associated to DA
 - iii) neutrophils associated to nephritis
- Different immune signatures correlating with SLEDAI (7 groups)
- Groups supported by different genotypes
- Supports the development of customized treatment strategies.

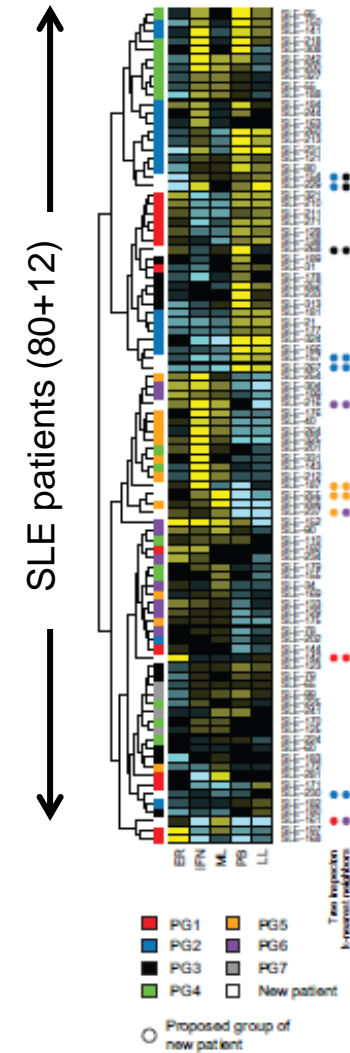
A Targeted Panel for SLE Patient Stratification

Can we find a gene panel which allows direct patients stratification ?

- Hierarchical clustering of the 797 transcripts **differentially correlating with SLEDAI** between the seven patient groups



- Assignment of novel 12 patients to each of the groups "guilt-by-association"



Study Summary

- Clinical and transcriptional profiling of 158 lupus patients up to a period of 4 years
- IFN, Plasmablast and Neutrophil signatures driving SLE
- Neutrophil-related signatures associate with progression to active nephritis
- Molecular correlates of disease activity stratify patients into seven major groups
- Molecular stratification may improve the outcome of clinical trials in SLE

Take Home Messages

- Interpreting biological meaning of ~25K genes in a heterogeneous sample such as blood – starting from bulk data – is extremely complex!
- One way to address this is to aggregate this information into functional building blocks (i.e. dimensionality reduction):
 - “blood modules” : of general use, obtained from previous analysis of 239 disease blood samples
 - “WGCNA modules”: patient specific and obtained from correlation analysis of the SLE study data set
- Blood modules can be (tentatively) assigned a biological meaning looking at their gene content
- Patient-specific WGCNA modules can be associated to clinical phenotypes based on longitudinal correlation
- For each sample, each module can be measured with one number (e.g. % of genes up- or down-regulated in the module, or eigenvalue). This approach has the statistical advantage of reducing number of tests
- Patients’ SLEDAI can be associated with different modules. On this basis patients can be stratified into classes, presumably having different underlying biology and needing different treatments

Q & A



My Best Reference for Systems Immunology: ImmunoSCOPE



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@Immunoscope

Building an immune compendium #immunology

📍 Paris, France 📅 Joined May 2018

1,328 Following 7,792 Followers

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Tweets & replies

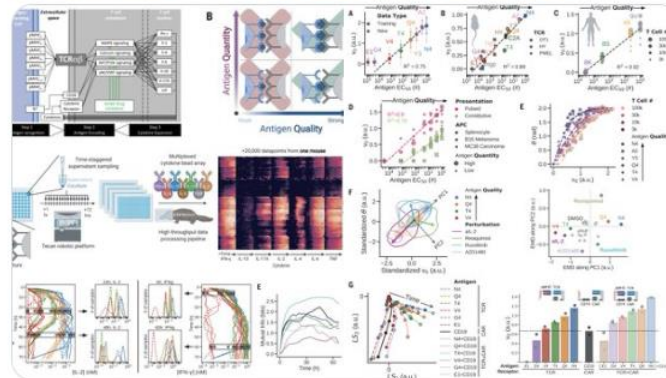
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ImmunoSCOPE @Immunoscope · May 19

Importance of antigen quality rather than quantity in predicting dynamic T cell responses | #CD8 #TCR #antigen #timecourse | Grégoire Altan-Bonnet @g_altanbonnet @p_r_francois @trademaker17 @McGillUPhysics @NCIResearchCtr | Science @ScienceMagazine bit.ly/3wwJUko



4

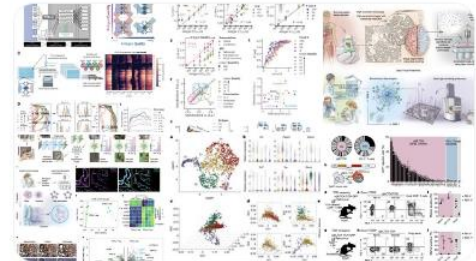


36



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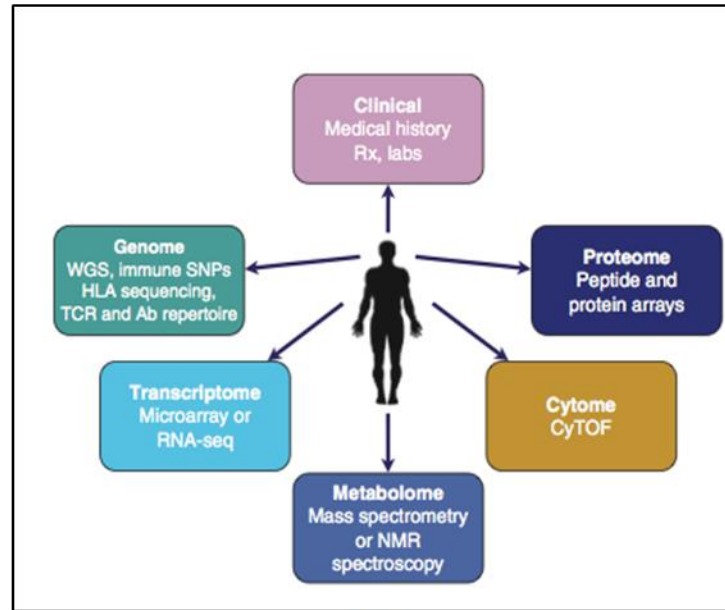
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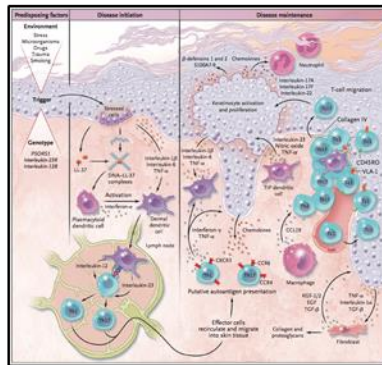
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Sign up

Computational Immunology



Kidd et al. Nat. Immunology 2014



*Understanding
Molecular Aetiology*

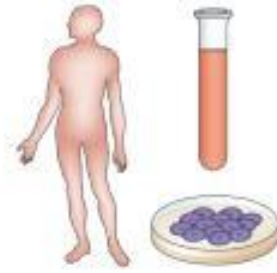
$$P \sim f(m)$$

*Explaining clinical phenotype as
function of molecular data.
Finding biomarkers*



*Identifying Novel
therapeutic targets*

Immune Phenotypes



Cell state or material

Resting

Cytokine stimulation

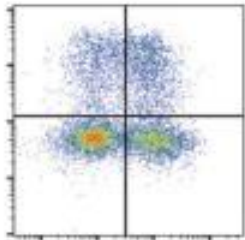
Non-antigenic stimulation

Antigenic stimulation

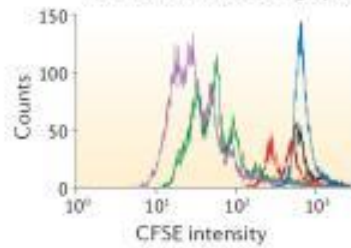
Blood plasma

Technique

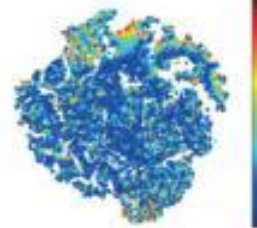
Flow cytometry



CFSE
(fluorescent cell-staining dye)



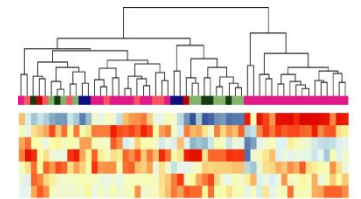
Mass cytometry



Luminex



RNA-Seq
Microarrays

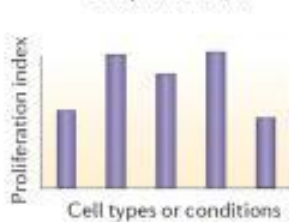


Measurement

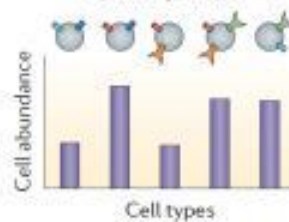
Signalling response



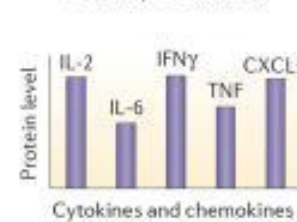
Cell proliferation



Cell frequencies



Serum protein levels



- RNA-levels
- Cell frequencies
- Cell clusters

The last two decades of single-cell

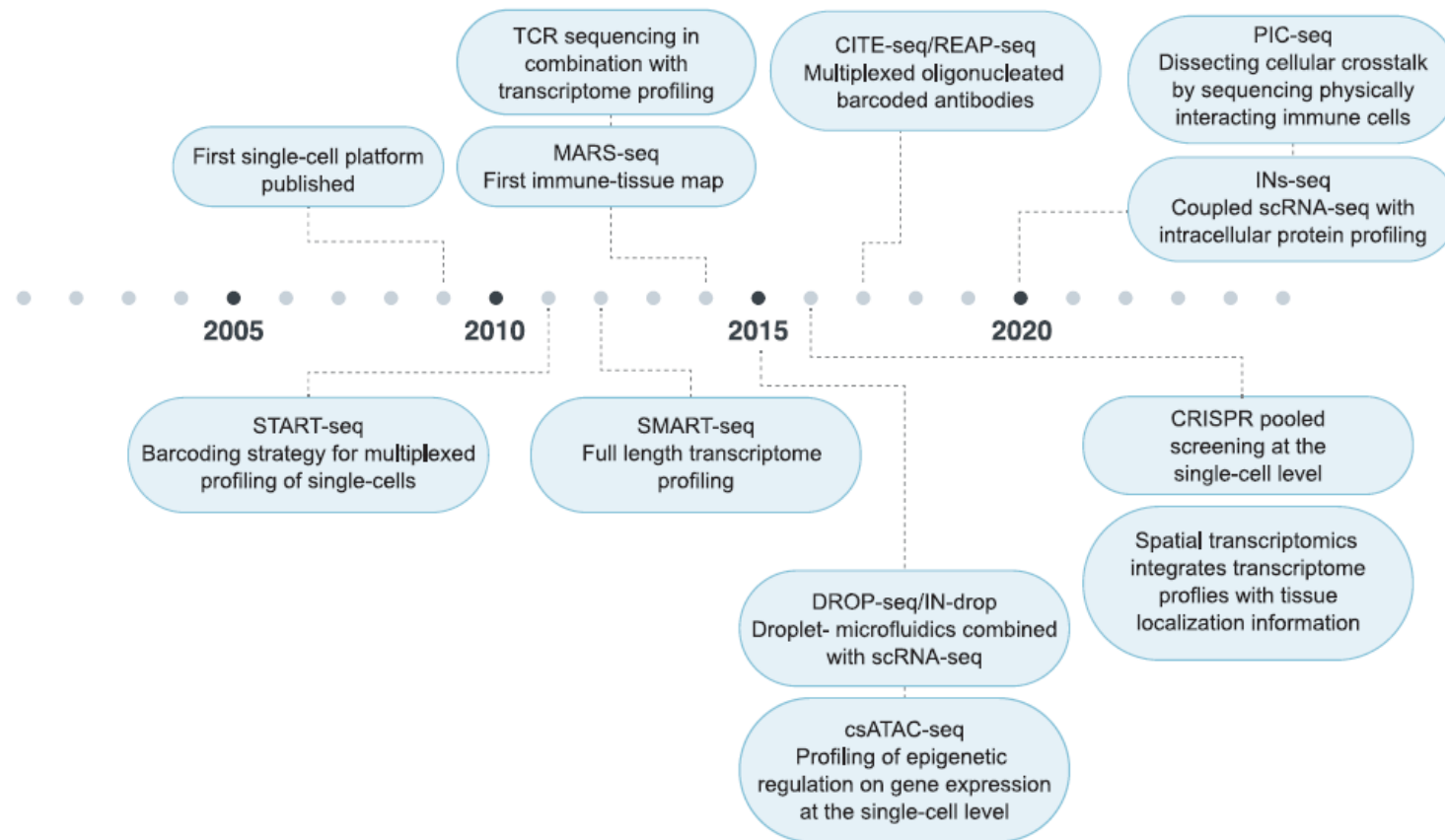
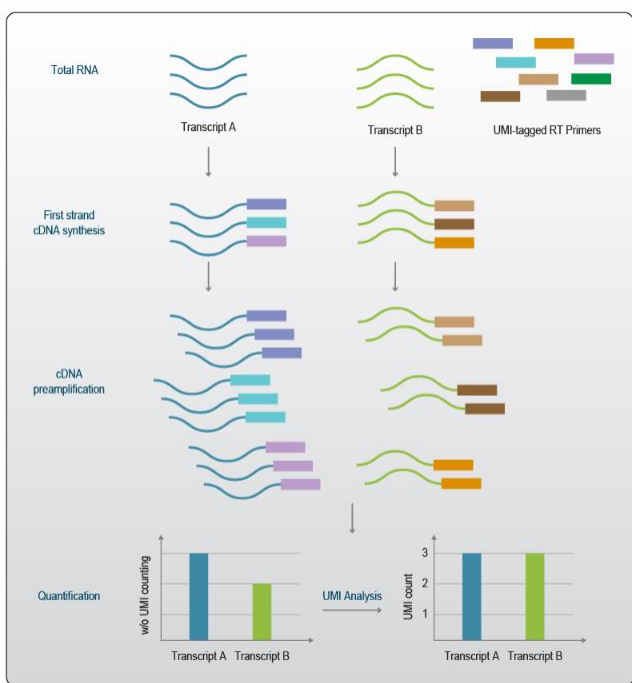
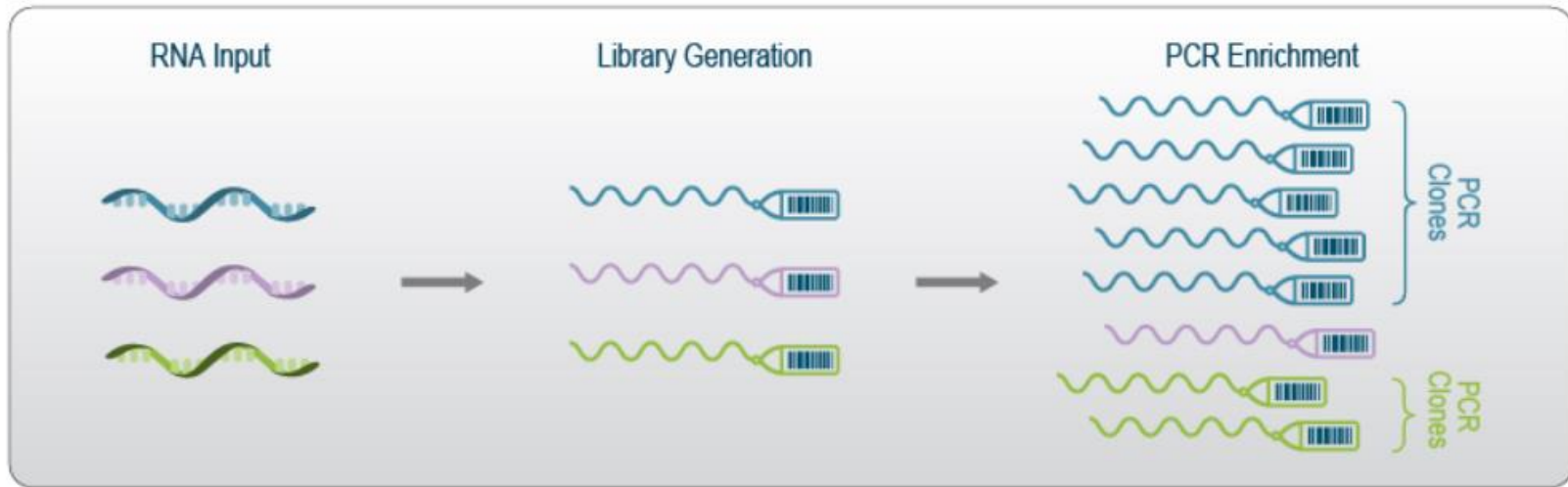


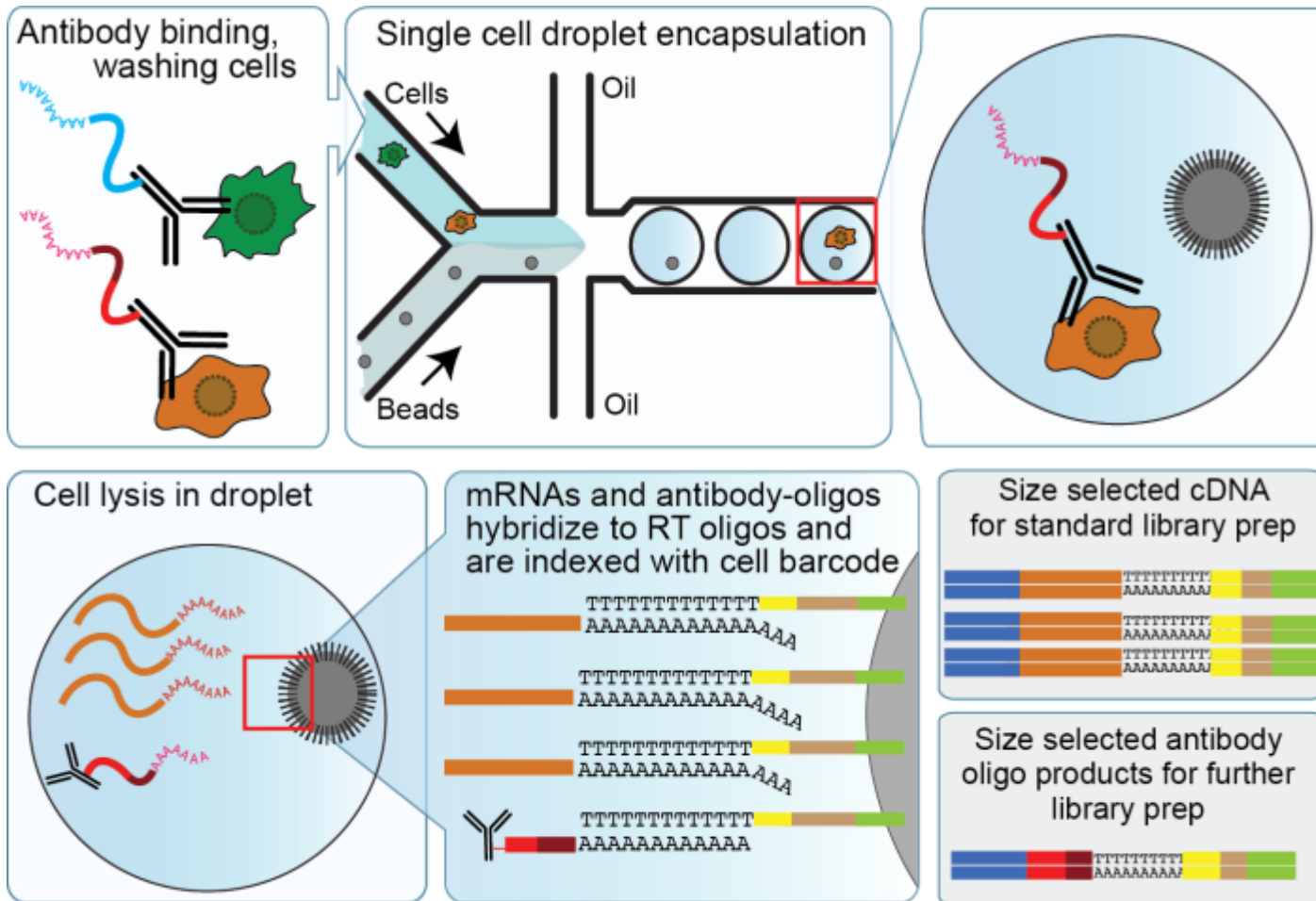
Figure 2. Timeline describing emerging single-cell technologies and their application in immunology research

What are Unique Molecular Identifiers (UMIs) and Why do We Need Them?



Transcript level quantification with UMIs. Transcripts or cDNAs are tagged with UMIs in an early step of library generation. The UMI sequences can then be used for quantification of the number of molecules that were originally present in the sample. UMIs can thus control for amplification biases associated with PCR-based sample preparation

CITE-Seq



Experimental methods for unimodal and multimodal single-cell measurements

Table 1 | Current experimental methods for unimodal and multimodal single-cell measurements

Data types	Method name	Feature throughput	Cell through put	Refs
Unimodal				
mRNA	Drop-seq	Whole transcriptome	1,000–10,000	4
	InDrop	Whole transcriptome	1,000–10,000	5
	10X Genomics	Whole transcriptome	1,000–10,000	6
	Smart-seq2	Whole transcriptome	100–300	30
	MARS-seq	Whole transcriptome	100–300	3
	CEL-seq	Whole transcriptome	100–300	1
	SPLIT-seq	Whole transcriptome	≥50,000	8
	sci-RNA-seq	Whole transcriptome	≥50,000	7
Genome sequence	SNS	Whole genome	10–100	9
	SCI-seq	Whole genome	10,000–20,000	10
Chromatin accessibility	scATAC-seq	Whole genome	1,000–2,000	13
	sciATAC-seq	Whole genome	10,000–20,000	14
	scTBS-seq	Whole genome	10,000–20,000	15
DNA methylation	scBS-seq	Whole genome	5–20	17
	snmC-seq	Whole genome	1,000–5,000	16
	sci-MET	Whole genome	1,000–5,000	19
	scRRBS	Reduced representation genome	1–10	18
Histone modifications	scChIP-seq	Whole genome + single modification	1,000–10,000	24
Chromosome conformation	scHi-C-seq	Whole genome	1–10	26
Multimodal				
Histone modifications + spatial	NA	Single locus + single modification	10–100	23
mRNA + lineage	scGESTALT	Whole transcriptome	1,000–10,000	32
	ScarTrace	Whole transcriptome	1,000–10,000	33
	LINNAEUS	Whole transcriptome	1,000–10,000	34
Lineage + spatial	MEMOIR	NA	10–100	27
mRNA + spatial	osmFISH	10–50 RNAs	1,000–5,000	35
	STARmap	20–1,000 RNAs	100–30,000	31
	MERFISH	100–1,000 RNAs	100–40,000	300
	seqFish	125–250 RNAs	100–20,000	29
mRNA + cell surface protein	CITE-seq	Whole transcriptome + proteins	1,000–10,000	20
	REAP-seq	Whole transcriptome + proteins	1,000–10,000	21
mRNA + chromatin accessibility	sci-CAR	Whole transcriptome + whole genome	1,000–20,000	40
mRNA + DNA methylation	scMGT-seq	Whole genome	50–100	46
mRNA + genomic DNA	OGT-seq	Whole genome + whole transcriptome	50–200	44
mRNA + intracellular protein	NA	96 mRNAs + 38 proteins	50–100	50
	NA	82 mRNAs + 75 proteins	50–200	49
DNA methylation + chromatin accessibility	scNOME-seq	Whole genome	10–20	11

Stuart T, Satija R. Nat Rev Genet. 2019