



### Making drugs from T cells: Quantitative Analysis of CAR-T Pharmacology

**FOC**IS Cancer Immunity & Immunotherapy Course

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## **Pharmacometrics:** Quantitative Pharmacokinetics & Pharmacodynamics (PKPD)

Pharmacokinetics (PK): Dose-Exposure



Pharmacodynamics (PD): Exposure-Response



PKPD: Dose regimen optimization



 $C_{max}$ : maximal concentration  $T_{max}$ : time at Cmax AUC : Area under the Curve CL: Clearance rate (~half-life<sup>-1</sup>)

Effect =  $E_{max} \cdot \left(\frac{C^k}{C^k + EC50^k}\right)$ Therapeutic index: EC50(efficacy) – EC50(tox)

How do we apply these quantitative metrics to adoptive T cell therapy?

# Adoptive T cell therapy: what drives exposure/response?



Mueller KT, Waldron ER, Grupp SA, et al (2018) Clinical Pharmacology of Tisagenlecleucel in B-Cell Acute Lymphoblastic Leukemia.

Clin Cancer Res 24(24):6175-6184

#### Distribution

- Where do T cells go?
- Does proliferation/expansion occur in tissues or blood?

#### **Cell Expansion**

- Memory vs. exhaustion phenotype...sometimes
- Intrinsic proliferative capacity of the cells
- CAR design & expression
- Patient cytokine levels
- Tumor burden

#### **Contraction & Clearance/Persistence**

- · Memory cell generation following antigen clearance
- Competition from host T cells for 'space'
- Allogeneic elimination (host vs. graft)

#### Anti-tumor efficacy & toxicity (CRS)

- Exposure (Cmax / AUC)
- Intrinsic cytotoxic potency
- CAR design & expression
- Tumor Microenvironment inflammatory/anti-inflammatory signals
- Tumor homing/penetration\*\*



Qi T, McGrath K, Ranganathan R, et al (2022) Cellular kinetics: A clinical and computational review of CAR-T cell pharmacology. Adv Drug Deliver Rev 188:114421.



# Adoptive T cell therapy: what drives exposure/response?



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## Outline

- 1. What pharmacometrics predict patient response?
  - *Empirical* pharmacokinetic (PK) modelling
- 2. What cell-intrinsic properties of the CART product underly the wide clinical variability?
  - Mechanistic PKPD modelling of Tcell:tumor interactions
  - Machine learning model for predicting response
- 3. What patient-intrinsic factors mediate response?
  - A. T cell bio-distribution\*
  - B. Tumor inflammation
  - C. Lympho-depletion regimen & patient response
  - D. Host vs. Graft (allogeneic clearance)



# 1. What CAR-T pharmacometrics predict response?



## CAR-T pharmacokinetic ("cellular kinetics") model Developed for Kymriah (TISAGENLECLEUCEL-T) BLA



#### Empirical model quantifies PK curves PK simulations vs. clinical data

#### 

#### Internal model simulations



#### **Model parameters**

\*Kalos et al (2011) Sci Transl Med 3:73-95.

PAARAMETR	THETA (mean)	ETA (variance)
Cmax	24000 (counts/ug)	0.65
Tmax	9.3 (day)	0.38
foldX (Cmax/C <sub>0</sub> )	3900	2.4
Fb (fraction Tm at tmax)	0.0079	0.8
Alpha (contraction)	0.16 day <sup>-1</sup>	0.91
Beta (persistence)	0.0032 day <sup>-1</sup>	0.86



Stein AM, Grupp SA, Levine JE, et al (2019) Tisagenlecleucel Model-Based Cellular Kinetic Analysis of Chimeric Antigen Receptor–T Cells. CPT Pharmacometrics Syst Pharmacol 8:285–295.

# CAR-T exposure-response analyses

Abecma in Multiple Myeloma



Connarn JN, Witjes H, Geffen M van Z, et al (2023) Characterizing the exposure–response relationship of idecabtagene vicleucel in patients with relapsed/refractory multiple myeloma. Cpt Pharmacometrics Syst Pharmacol. https://doi.org/10.1002/psp4.12922



### Inter-individual variability (IIV) washes out dose-responses Kymriah in DLBCL





Awasthi R, Pacaud L, Waldron E, et al (2020) Tisagenlecleucel cellular kinetics, dose, and immunogenicity in relation to clinical factors in relapsed/refractory DLBCL. Blood 8 Adv 4:560–572.

# 

# 2. What cell-intrinsic properties underly clinical variability and response?



McLane LM, Abdel-Hakeem MS, Wherry EJ (2015) CD8 T Cell Exhaustion During Chronic Viral Infection and Cancer. Annu Rev Immunol 37:1–39.



DC Kirouac, C Zmurchok, A Deyati, J Sicherman, C Bond, PW Zandstra. Deconvolution of clinical variance in CAR-T pharmacology and response. Nat Biotech 2023

# "Toggle switch" model structure and assumptions



- $T_M$ : memory T cells
- T<sub>E</sub>: effector T cells
- T<sub>X</sub>: exhausted T cells
- B: B cells (tumor)
- B<sub>A</sub>: B cell antigen

### T cell differentiation toggle switch





### Model training data: Kymriah in Chronic Lymphoblastic Leukemia PKPD profiles, CAR-T product transcriptomes and immuno-phenotypes vs. response



#### Population mean PKPD: Kymriah in Chronic Lymphoblastic Leukemia (CLL)

- Can we recapitulate the pharmacokinetics & tumor dynamics (PKPD) based on T cell biology?
- What kinetic parameters / molecular features distinguish robust vs. poor responding patients?

#### Pre-infusion CAR-T transcriptomes



CR=5, PR =5, NR=21





Fraietta JA, Lacey SF, Orlando EJ, et al (2018) Determinants of response and resistance to CD19 chimeric antigen receptor (CAR) T cell therapy of chronic lymphocytic leukemia. Nat Med 24:563–571.

# Model development and validation workflow



## Model calibration & analysis What features (model parameters) separate clinical outcomes?



What differentiates CR vs. NR?

#### CAR-T products in CR vs. NR show:

- Heightened memory cell turnover ( $\mu_M$ ,  $d_M$ )
- Heightened cytotoxic potency (*TK50*)
- Little difference in Tmem/Texh frequency 3



\*Assume Dose = 10<sup>8</sup> cells, Tumor burden = 10<sup>10</sup> cells (median reported); Estimate parameters using PSO: simulations represent 90% confidence intervals



Scale counts/ug to cell/uL using data from: Kalos, M. et al. T Cells with Chimeric Antigen Receptors Have Potent Antitumor Effects and Can Establish Memory in Patients with Advanced Leukemia. Sci Transl Med 3, 95ra73-95ra73 (2011).

# 'Validation' of model inferences via single-cell transcriptomes

Mathematical inferences assessed in an additional blood cancer: Acute Lymphoblastic Lymphoma

T cell composition (memory vs. exhausted cells) does not substantially vary by response category



**\*ProjecTILS annotation**: Andreatta M, Corria-Osorio J, Müller S, et al (2021) Interpretation of T cell states from single-cell transcriptomics data using reference atlases. Nat Commun 12:2965.

**CART Dysfunction**: Good CR, Aznar MA, Kuramitsu S, et al (2021) An NK-like CAR T cell transition in CAR T cell dysfunction. Cell.



# 'Validation' of model inferences via single-cell transcriptomes

Mathematical inferences assessed in an additional blood cancer: Acute Lymphoblastic Lymphoma

#### T memory cells from NR patients display intrinsic functional deficits analogous to T cell exhaustion





\*ProjecTILS annotation: Andreatta M, Corria-Osorio J, Müller S, et al (2021) Interpretation of T cell states from single-cell transcriptomics data using reference atlases. Nat Commun 12:2965.

CART Dysfunction: Good CR, Aznar MA, Kuramitsu S, et al (2021) An NK-like CAR T cell transition in CAR T cell dysfunction. Cell.



Tem, Tmem cells from NR samples appear functionally exhausted \*Tem: defined via ProjecTILs algorithm \*Tmem: defined via CD8+CD45RO-CD27+ CITEseg tags

Cell-intrinsic differences



Data Source: Bai Z, Woodhouse S, Zhao Z, et al (2022) Single-cell antigen-specific landscape of CAR T infusion product identifies determinants of CD19-positive relapse in patients with ALL. Sci Adv 8:.

# CAR-T clinical response prediction

Are pre-infusion CAR-T transcriptomes predictive of clinical response (CR vs. NR)?





Lage P, small N problem: the central challenge in biomedical genomics

### **CAR-T clinical response prediction** Are pre-infusion CAR-T transcriptomes predictive of clinical response (CR vs. NR)?

scRNAseq pre-infusion CAR-Ts **CR/NR/PR** classes **Kymriah in ALL** Yescarta in LBCL Kymriah in LBCL Kymriah in ALL (Bai 2022) (Haradhvala 2022) (Haradhvala 2022) В Transcriptome D С Transcriptome Transcriptome % Tex & Tmem – – – % Tex & Tem – – – % Tex & Tem 2.0 — Null — Null — Null \*\*\* CR = 6 CR = 11 20 CR = 5; NR/RL = 7 **PR/NR = 8** NR = 7 Kymriah in LBCL Density\_ Density Density 1.0 1.0 = 6 : NR = 7 0.5 0.5 Yescarta in LBCBL 0.0 0.00 0.25 0.50 0.75 0.75 1.00 0.25 Accuracy 0.00 0.25 0.50 0.75 1.00 Accuracy Accuracy Accuracy = 80% Accuracy = 80%Accuracy = 71%Tmem, Tex:ProjecTILS Tmem, Tex: CITESeg data Tmem, Tex: ProjecTILS\* 11: NR = 8 CR = 5: NR/RL = 7 CR = 6 ; NR = 7 CR = 11: NR/PR = 8 \*\*\* $P < 10^{-8}$  (rank-sum test) CR NR

> Functional attributes predictive of clinical outcomes are CART-cell-intrinsic & indication-agnostic Transcriptome > 'gold standard' immunophenotyping

Predictive accuracy of response classification using 60:40 train:test splits

### CAR-T clinical response prediction What transcriptional signatures are predictive of CAR-T response?

CAR-T Response Score-card



# 3. Patient-intrinsic factors mediating response

- A. T cell biodistribution
- B. Tumor Inflammation
- C. Response to Lympho-depletion & host-T cell competition
- D. Host vs. Graft response (allogeneic elimination)

### **3A. Adoptive T cell Biodistribution** Where do CAR-Ts go once administered? What happens in tissues vs. Blood?

#### Pharmacokinetics & biodistribution of radio-labelled T cells in mice Whole blood Lungs Heart AUC = 86.3 ± 2.6 AUC = 1727 ± 110.5 $AUC = 43.9 \pm 4.3$ BC = 39 3 BC = 2.010 10 3 0.1 0.01 100 200 100 200 300 10 \* Spleen Liver Tumor AUC = 19646.1 ± 1111.7 AUC = 6457.2 ± 659.6 AUC = 57.1 ± 10.8 1000 1000 BC = 447.6BC = 1.3BC = 147.5CAR-T (cells/uL) 100 10 <sup>0</sup> 200 200 300 100 10 -1 IGLN Bone TDLN AUC = 635 ± 76.6 AUC = 1204.9 ± 305.4 AUC = 711.8 ± 191.6 100 100 BC = 27.5 BC = 14 5 BC = 16.2 10 -2 10 0 **\*BC** = Biodistribution Coefficient.

= AUC of T cells in tissue vs. blood

# Majority of administered T cells distribute to lungs, spleen, liver, kidney & lymph nodes.

**Khot A**, Satoko M, Thomas VA, et al (2019) Measurement and Quantitative Characterization of Whole-Body Pharmacokinetics of Exogenously Administered T Cells in Mice. J Pharmacol Exp Ther 368:jpet.118.252858.

Pharmacology 'accounting' in man vs. mouse





\*ER = Expansion Ratio. How many cells do you detect at Cmax per infused? = Cmax\*Vblood / Dose

Q: Where do the majority of CARTs distribute Q: Where does the 'action' happen (tissue vs. blood)?

## **3B. Tumor inflammation and CAR-T response** Yescarta (CD19-CART) in DLBCL: ZUMA-1 trial

'Immunoscore' (Tumor inflammation) is the most significant patient-intrinsic predictor of CART response Immunoscore (Tumor inflammation) also drives Cmax



Q: How would pre-existing TILs influence CAR-T expansion?

# **3C. Lympho-depletion intensity & response** via IL7 availability?



Q: How does Lympho-depletion intensity affect CAR-T expansion and peak IL7 concentration? Q: Can we *mimic* intense-LDT via cytokine support?



Hirayama AV, Gauthier J, Hay KA, et al (2019) The response to lymphodepletion impacts PFS in patients with aggressive non-Hodgkin lymphoma treated with CD19 CAR T cells. Blood 133:1876–1887.

# Competition between Adoptive vs. Patient T cells

#### **Model structure**



#### Model fitting: Yescarta in LBCL (ZUMA-1) CAR-T and Host-T cell kinetics





Kimmel GJ, Locke FL, Altrock PM (2021) The roles of T cell competition and stochastic extinction events in chimeric antigen receptor T cell therapy. Proc Royal Soc B 288:20210229.

### **3D. Host vs. Graft response (allogeneic elimination)** Host T cells actively clear (allogenic) T cell grafts

UCART19 in B-ALL: The first reported allogeneic CAR-T clinical data CD19-CART, allogeneic (healthy donor-derived) T cells, *TRAC*<sup>-/-</sup>



• Deeper LDT & slower T cell reconstitution ~ greater allogenic CART exposure Q: How would additional gene edits (i.e. MHC-knock out) affect allo-clearance rates

**Data digitized from:** Derippe T, Fouliard S, Marchiq I, et al (2022) Mechanistic modeling of the interplay between host immune system, interleukin 7 and UCART19 allogeneic CAR-T cells in adult B-cell acute lymphoblastic leukemia. Cancer Res Commun 2:1532–1544.



**Kymirah PK simulations:** Stein AM, Grupp SA, Levine JE, et al (2019) Tisagenlecleucel Model-Based Cellular Kinetic Analysis of Chimeric Antigen Receptor–T Cells. Cpt Pharmacometrics Syst Pharmacol 8:285–295.

# The next frontier: iPSC-derived CAR-Ts

FT819: The first reported clinically tested iPSC-derived CART CD19-CART, allogeneic (iPSC-differenentiated) T cells, *TRAC*<sup>-/-</sup>



Both robust cell expansion + persistence (AUC) is required for clinical activity

**Q:** Why are (FT819) iPSC-CARTs incapable of persistence - *Cell intrinsic* deficit vs. *allogeneic*-clearance?



# Summary

- 1. Empirical PKPD models
- Cmax predicts response
- High variability makes doseoptimization infeasible



# 2. Mechanistic modelling & machine learning

- Product intrinsic-proliferation of memory cells is important for clinical response
- Predictive features are buried in CART transcriptomes



#### 3. Patient-intrinsic effects

 Biodistribution, inflammatory state, lympho-depletion response, and Host vs Graft affect PK and response



Mathematical models can enable CAR-T design, optimization and data interpretation Quantitative data is required to translate measurements to kinetic parameters



### **Thank You!**

### Vancouver, BC

- ✓ Developmental immunology
- ✓ Systems Biology and T cell pharmacology

### Seattle, WA

- Protein and genome engineering
  Translational sciences
- ✓ Cancer biology



#### Avisek Deyati, Jordan Sicherman Cole Zmurchok Peter Zandstra, Chris Bond, Gregory Block Irja Elliott Donaghue

# Toronto, ON✓ GMP iPSCs and gene editing

T cell manufacturingQA/QC



# THERAPEUTICS

- Confidential -

# What value does modelling bring to drug development?

The biological mechanisms underlying experimental data are often complex and non-intuitive



The number of possible experiments to conduct is infinite



### **3D.** Host vs. Graft response (allogeneic elimination) Host T cells actively clear (allogenic) T cell grafts

UCART19 in B-ALL: The first reported allogeneic CAR-T clinical data CD19-CART, allogeneic (healthy donor-derived) T cells, TRAC-/-



Dupouy S, Marchiq I, Derippe T, et al (2022) Clinical Pharmacology and Determinants of Response to UCART19, an Allogeneic Anti-CD19 CAR-T Cell Product, in Adult B-cell Acute Lymphoblastic Leukemia. Cancer Res Commun 2:1520-1531.



Host T cell reconstitution limits CAR-T expansion

### Initial expansion (Cmax) predicts response for multiple CAR-Ts Clearance does not (for autologous products)



Liu C, Ayyar VS, Zheng X, et al (2020) Model-based Cellular Kinetic Analysis of Chimeric Antigen Receptor-T Cells in Humans. Clin Pharmacol Ther.

Cell Kinetic model to data from 7 CART trials (Jansen)





## Model-based insights into clinical response: cell dose & tumor burden

Virtual Populations vs. Yescarta in LCBCL (ZUMA-1) В С А Dose/B0 Random Param. 0.75 0.75 0.75 Yescarta Response 0.5 0.5 0.5 \*\*\*\*\* 0.25 0.25 0.25 10<sup>-2</sup>  $2.7 \times 10^{10}$ 10<sup>-2</sup> 10<sup>-1</sup> 10<sup>0</sup>  $10^{2}$  $8.5 \times 10^{8}$  $10^{1}$  $10^{3}$  $10^{0}$  $10^{-4}$ Tumor Burden (cells) Cmax/Tumor Burden (dimensionless) Cmax (cells/uL)

Predicted covariates of response: Cmax vs. Tumor Burden

**Data source:** Locke FL, Rossi JM, Neelapu SS, et al (2020) Tumor burden, inflammation, and product attributes determine outcomes of axicabtagene ciloleucel in large B-cell lymphoma. Blood Adv 4:4898–4911.

Mechanism-based models can *predict* biological processes underlying clinical observations



#### Model training: Ph1 Abecma dose escalation (BCMA, Multiple Myeloma)

#### Predicted sub-population dynamics: Ph1 dose escalation





# Lympho-depletion intensity & response via IL7 availability?

Cycolophosphamide (Cy) vs. Cy + Fludarabine (Flu): CD19-CART therapy in B-ALL



Turtle CJ, Hanafi L-A, Berger C, et al (2016) CD19 CAR–T cells of defined CD4+:CD8+ composition in adult B cell ALL patients. J Clin Invest 126:2123–2138.

Q: How does Lympho-depletion intensity affect CAR-T expansion and peak IL7 concentration?

Q: Can we *mimic* intense-LDT via cytokine support?

#### High vs. Low-intensity Cy+Flu: CD19-CART therapy in NHL



<sup>\*60</sup> vs. 30 mg/kg cyclophosphamide

Hirayama AV, Gauthier J, Hay KA, et al (2019) The response to lymphodepletion impacts PFS in patients with aggressive non-Hodgkin lymphoma treated with CD19 CAR T cells. Blood 133:1876–1887.



## **3B. Tumor inflammation and CAR-T response** Yescarta in DLBCL: ZUMA-1



T cell inflamed tumors ~ improved survival

#### **Tumor inflammation ~ Cmax (CART expansion)**



### Immunoscore is the most significant "co-variate"

Cox-regression (statistical) model

Variable	Ν		HR (95% CI)	P value
Immunoscore				
High	17	<b>•</b>	Reference	
Low	12		412.45 (2.63, 64,685.	90)0.020
Gender			No. 22 No.	
Female	13		Reference	
Male	16	⊢∎	0.51 (0.05, 5.27)	0.569
Subtypest				
GCB	18	<b>•</b>	Reference	
ABC	4	$\leftarrow$ $\stackrel{!}{\longrightarrow}$	• 0.00 (0.00, Inf)	0.999
N/A	1	$\leftrightarrow$	0.00 (0.00, Inf)	1.000
Unknown	6	⊢⊨∎−−1	2.00 (0.12, 34.57)	0.634
IPI				
Low	6	ŧ	Reference	
Intermediate	10	⊢ <b></b>	0.64 (0.02, 16.94)	0.787
High	13		0.06 (0.00, 16.40)	0.327
Baseline tumor burden (SPD)	29	•	1.00 (1.00,1.00)	0.018
BCL2 overexpression				
Yes	16		Reference	
No	8	<b>∉</b> _ <u>+</u>	0.07 (0.00, 2.16)	0.127
Unknown	5	$\leftrightarrow$	0.00 (0.00, Inf)	1.000
c-MYC overexpression				
Yes	10		Reference	
No	14	· · · · · · · · · · · · · · · · · · ·	202.19 (3.98, 10,283.	23)0.008
Unknown	5	$\leftrightarrow$	0.00 (0.00, Inf)	1.000
BCL6 overexpression				
Yes	15		Reference	
No	10	- <b>-⊞</b>	6.52 (0.65, 65, 49)	0.111
Unknown	4	$\leftrightarrow$	Inf (0.00, Inf)	0.999
		·····	(111)	
		0, 10,00,000		
		1, 00,		
		HB (95% CI)		

#### **Q: How would pre-existing TILs influence CAR-T expansion?**

